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U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER

**TRANSMITTAL LETTER TO THE UNITED STATES  
DESIGNATED/ELECTED OFFICE (DO/EO/US)  
CONCERNING A FILING UNDER 35 U.S.C. 371**

**PG3606USW**

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR

**09/857123**

INTERNATIONAL APPLICATION NO.

**PCT/EP99/09284**

INTERNATIONAL FILING DATE

**30 November 1999**

PRIORITY DATE CLAIMED

**1 December 1998**

TITLE OF INVENTION

**HUMAN VANILLOID RECEPTORS AND THEIR USES**

APPLICANT(S) FOR DO/EO/US

**DELANY, Natalie Samantha**

**TATE, Simon Nicholas**

**SANSEAU, Philippe**

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This is an express request to begin national examination procedures (35 U.S.C. 371(f)). The submission must include items (5), (6), (9) and (24) indicated below.
4. ☒ The US has been elected by the expiration of 19 months from the priority date (Article 31).
5. ☒ A copy of the International Application as filed (35 U.S.C. 371 (c) (2))
  - a. ☐ is attached hereto (required only if not communicated by the International Bureau).
  - b. ☒ has been communicated by the International Bureau.
  - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☒ An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)).
  - a. ☐ is attached hereto.
  - b. ☒ has been previously submitted under 35 U.S.C. 154(d)(4).
7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371 (c)(3))
  - a. ☐ are attached hereto (required only if not communicated by the International Bureau).
  - b. ☐ have been communicated by the International Bureau.
  - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
  - d. ☒ have not been made and will not be made.
8. ☐ An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☐ An oath or declaration of the inventor(s) (35 U.S.C. 371 (c)(4)).
10. ☐ An English language translation of the annexes of the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371 (c)(5)).
11. ☒ A copy of the International Preliminary Examination Report (PCT/IPEA/409).
12. ☒ A copy of the International Search Report (PCT/ISA/210).

**Items 13 to 20 below concern document(s) or information included:**

13. ☒ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
14. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
15. ☒ A **FIRST** preliminary amendment.
16. ☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
17. ☐ A substitute specification.
18. ☐ A change of power of attorney and/or address letter.
19. ☐ A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 35 U.S.C. 1.821 - 1.825.
20. ☐ A second copy of the published international application under 35 U.S.C. 154(d)(4).
21. ☐ A second copy of the English language translation of the international application under 35 U.S.C. 154(d)(4).
22. ☒ Certificate of Mailing by Express Mail
23. ☒ Other items or information:

**Copy of Request (PCT/RO/101)**

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR

097/857123

INTERNATIONAL APPLICATION NO.

PCT/EP99/09284

ATTORNEY'S DOCKET NUMBER

PG3606USW

24. The following fees are submitted:

**BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)) :**

- ☐ Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO ..... \$1000.00
- ☒ International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO ..... \$860.00
- ☐ International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO ..... \$710.00
- ☐ International preliminary examination fee (37 CFR 1.482) paid to USPTO but all claims did not satisfy provisions of PCT Article 33(1)-(4) ..... \$690.00
- ☐ International preliminary examination fee (37 CFR 1.482) paid to USPTO and all claims satisfied provisions of PCT Article 33(1)-(4) ..... \$100.00

**ENTER APPROPRIATE BASIC FEE AMOUNT =****CALCULATIONS PTO USE ONLY**

\$860.00

Surcharge of \$130.00 for furnishing the oath or declaration later than ☐ 20 ☐ 30 months from the earliest claimed priority date (37 CFR 1.492 (e)).

\$0.00

CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE
Total claims	51 - 20 =	31	x \$18.00
Independent claims	5 - 3 =	2	x \$80.00

\$558.00

\$160.00

Multiple Dependent Claims (check if applicable). ☐

\$0.00

**TOTAL OF ABOVE CALCULATIONS =**

\$1,578.00

Applicant claims small entity status. (See 37 CFR 1.27). The fees indicated above are reduced by 1/2.

\$0.00

**SUBTOTAL =**

\$1,578.00

Processing fee of \$130.00 for furnishing the English translation later than ☐ 20 ☐ 30 months from the earliest claimed priority date (37 CFR 1.492 (f)).

\$0.00

**TOTAL NATIONAL FEE =**

\$1,578.00

Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) (check if applicable). ☐

\$0.00

**TOTAL FEES ENCLOSED =**

\$1,578.00

Amount to be:  
refunded \$  
charged \$

- a. ☐ A check in the amount of \_\_\_\_\_ to cover the above fees is enclosed.
- b. ☒ Please charge my Deposit Account No. 07-1392 in the amount of \$1,578.00 to cover the above fees. A duplicate copy of this sheet is enclosed.
- c. ☒ The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 07-1392. A duplicate copy of this sheet is enclosed.
- d. ☐ Fees are to be charged to a credit card. **WARNING:** Information on this form may become public. **Credit card information should not be included on this form.** Provide credit card information and authorization on PTO-2038.

**NOTE:** Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:



23347

PATENT TRADEMARK OFFICE

SIGNATURE

Frank Grassler

NAME

31,164

REGISTRATION NUMBER

DATE

June 1, 2001

## IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re: Application of: DELANY et al.  
International Application No.: PCT/EP99/09284  
International Filing Date: 30 November 1999  
Title: HUMAN VANILLOID RECEPTORS AND THEIR USES

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Honorable Commissioner of Patents  
Washington, D.C. 20231

## FIRST PRELIMINARY AMENDMENT

Dear Sir:

The above-identified application is being transmitted herewith for entry in the US National Phase under Chapter II of the PCT for the purpose of adding the priority information. Please amend the application as follows:

In the Abstract:

The Abstract has been placed on a separate sheet of paper according to US practice, as required under 37 CFR 1.72(b).

In the Specification:

On the first line of the specification, after the Title, please add:

--This application is filed pursuant to 35 U.S.C. §371 as a United States National Phase Application of International Application No. PCT/EP99/09284 filed 30 November 1999, which claims priority from Great Britain Application No. 9826359.3 filed 1 December 1998.--

In the Claims

14. (Amended) An expression vector comprising a nucleotide sequence according to claim 6, which is capable of expressing an hVR protein or a variant thereof.

23. (Amended) An antibody specific for a human vanilloid receptor (hVR) protein or a variant thereof as claimed in claim 1.

26. (Amended) A method for identification of a compound which exhibits hVR modulating activity comprising contacting a human vanilloid receptor (hVR) protein or a variant thereof according to claim 1 with a test compound and detecting modulating activity or inactivity.

45. (Amended) A method of producing an hVR protein or a variant thereof according to claim 1 comprising introducing into an appropriate cell line a suitable vector comprising a nucleotide sequence encoding for an hVR protein or a variant

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thereof, under conditions suitable for obtaining expression of the hVR protein or variant thereof.

50. (Amended) A human vanilloid receptor (hVR) protein according to claim 48, which is hVR1 or a variant thereof.

51. (Amended) A human vanilloid receptor (hVR) protein according to claim 48 which is hVR3 or a variant thereof.

#### REMARKS

Applicants have attached an abstract on a separate sheet of paper as required by US practice. Applicants have amended the specification for purposes of adding the priority information. Claims have been amended to eliminate multiple dependencies.


Attached hereto is a marked-up version of the changes made to the specification by the current amendment. The attached page is captioned "Version with markings to show changes made." Applicant respectfully requests the entry of the above preliminary amendments.

Examiner is invited and encouraged to contact the undersigned if such contact would facilitate prosecution of this application.

No fee is believed due in connection with this Amendment, however the Commissioner is hereby authorized to charge any under-payment to Deposit Account No. 07-1392.

Respectfully submitted,

Date: June 1, 2001

  
Frank P. Grassler  
Attorney of Record, Reg. No. 31,164

GlaxoSmithKline  
Corporate Intellectual Property Department  
Five Moore Drive, PO Box 13398  
Research Triangle Park, NC 27709-3398  
Telephone: 919-483-3934  
Facsimile: 919-483-7988

CERTIFICATE OF EXPRESS MAILING (37 CFR 1.10)  
I hereby certify that this paper (along with any referred to as being attached or enclosed) is being deposited with the United States Postal Service "Express Mail Post Office to Addressee" service under 37 CFR 1.10 in an envelope addressed to: Assistant Commissioner of Patents Washington, D.C. 20231 on 6/10/01

  
Marilyn Eldridge



**Version with markings to show changes**

14. An expression vector comprising a nucleotide sequence according to [any one of] claim[s] 6 [to 13].
23. An antibody specific for a human vanilloid receptor (hVR) protein or a variant thereof as claimed in [any one of] claim[s] 1 [to 5].
26. A method for identification of a compound with exhibits hVR modulating activity comprising contacting a human vanilloid receptor (hVR) protein or a variant thereof according to [any one of] claim[s] 1 [to 5] with a test compound and detecting modulating activity or inactivity.
45. A method of producing an hVR protein or a variant thereof according to [any one of] claim[s] 1[-5] comprising introducing into an appropriate cell line a suitable vector comprising a nucleotide sequence encoding for an hVr protein or a variant thereof, under conditions suitable for obtaining expression of the hVR protein or variant thereof.
50. A human vanilloid receptor (hVR) protein according to claim 48 [or 49] which is hVR1 or a variant thereof.
51. A human vanilloid receptor (hVR) protein according to claim 48 [or 49] which is hVR3 or a variant thereof.

## HUMAN VANILLOID RECEPTORS AND THEIR USES

### Field of the Invention

5 The present invention relates to human vanilloid receptor (hVR) proteins and to related nucleotide sequences, expression vectors, cell lines, antibodies screening methods, compounds, methods of production and methods of treatment, as well as other related aspects.

### Background of the Invention

10 Capsaicin, the irritant in hot peppers and a member of the vanilloid family activates a sub-group of sensory neurons: the nociceptors. These neurons transmit nociceptive and thermoceptive pain information back to pain-processing centres in the central nervous system such as the spinal cord and the brain. They are also sites for the release of pro-inflammatory mediators in the periphery (1). Nociceptors show heterogeneity in their sensitivity to capsaicin. Excitation and prolonged exposure of these neurons to capsaicin is followed by a refractory state known as desensitisation (2) when they become insensitive to capsaicin and other noxious stimuli (3). The long-term response to insensitivity could be explained by death of the nociceptors or destruction of its peripheral terminals (4). Because of the desensitisation phenomenon, capsaicin has been used therapeutically for decades as an analgesic agent for the treatment of pain in a range of disorders (5).

20 It has been speculated that the endogenous target for capsaicin plays an important function in the detection of painful stimuli. It has been shown by electrophysiological and biochemical studies that capsaicin induces a flux of cations in dorsal root ganglion (DRG) neurons (6,7). Because other vanilloid derivatives show responses in a dose dependent manner (8,9) a receptor is the most likely candidate to explain the mechanism. Therefore, based on indirect evidence it has been anticipated that these actions of capsaicin (excitation / desensitisation) are mediated by a specific membrane-bound receptor named vanilloid receptor (10).

35 Evidence for the existence of a vanilloid receptor came from binding experiments with resiniferatoxin (RTX), a capsaicin analog (11), and a competitive antagonist

of capsaicin, capsazepine (12). Vanilloid receptors have been visualised by using ( $[^3\text{H}]$ -RTX) autoradiography in dorsal root ganglia (DRG) and spinal cord of different species including man (13,14).

5 Recently, a rat vanilloid receptor termed VR1 has been identified using an expression-cloning strategy to isolate the complementary DNA (cDNA) encoding the corresponding protein from a rat DRG cDNA library (15). The cDNA clone was completely sequenced. The rat VR1 cDNA has an open reading frame of 2,514 nucleotides and encodes for a protein of 838 amino acids with a predicted relative molecular mass of 95,000. Analysis of the amino acid sequence identified 6 potential transmembrane regions with a short hydrophobic stretch between the transmembrane regions 5 and 6. The N-terminus (amino terminal) contains three ankyrin repeat domains. No motifs have been identified at the C-terminus (carboxy terminal).

10 It has been noted that rat VR1 transfected cells exhibit an increase in calcium levels after heat treatment and it has been suggested that *in vivo* VR1 and vanilloid receptors are involved in detection of noxious heat (but not innocuous heat). It has also been proposed that protons could act as modulators of the vanilloid receptors (16, 17, 18).

15 While it has been recognised that the rat capsaicin receptor, VR1, is a member of the family of non-selective ion channels that are gated by ligands and that it is involved in pain sensation, the natural ligand of VR1 remains unknown. It is therefore suggested that human vanilloid receptor sub-types may provide targets for the development of novel analgesic agents (agonists and antagonists) and agents which may interact with other disorders.

20 Accordingly, it is an object of the present invention to locate and characterise human vanilloid receptors. Other objects of the present invention will become apparent from the following detailed description thereof.

### Summary of the Invention

25 According to one embodiment of the present invention there is provided an isolated human vanilloid receptor (hVR) protein or a variant thereof. Preferably

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the hVR protein is an hVR1 or hVR3 protein or a variant thereof. In a particularly preferred aspect of the invention the hVR protein has an amino acid sequence as shown in figure 3 or in figure 18.

5 According to another aspect of the invention, there is provided a human vanilloid receptor (hVR) protein or a variant thereof, preferably hVR1 or hVR3 or a variant thereof, for use in a method of screening for agents useful in the treatment or prophylaxis of a disorder which is responsive to the modulation of hVR activity, preferably hVR1 or hVR3 activity, in a human patient. Preferably the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), a respiratory disorder such as asthma or chronic obstructive pulmonary disease (COPD), a urological disorder such as diabetic neuropathy, incontinence and interstitial cystitis, or an inflammatory disorder.

10 According to another aspect of the invention there is provided a nucleotide sequence encoding a human vanilloid receptor (hVR) protein or a variant thereof as hereinbefore described, or a nucleotide sequence that is complementary thereto. Preferably the nucleotide sequence encodes an hVR1, hVR3 protein or variant thereof or a nucleotide sequence which is complementary thereto. Particularly preferably the nucleotide sequence is as shown in figure 2 and figure 17.

25 According to another aspect of the invention there is provided an expression vector comprising a nucleic acid sequence as referred to above which is capable of expressing an hVR protein as hereinbefore described or a variant thereof, preferably hVR1 or hVR3 or a variant thereof. Preferably the expression vector is as displayed in figure 6 or figure 20.

30 According to another aspect of the invention there is provided a stable cell line comprising an expression vector as referred to above which is capable of expressing an hVR protein as hereinbefore described or a variant thereof, preferably hVR1 or hVR3 or a variant thereof. The stable cell line is preferably a

modified mammalian cell line, preferably HEK293, CHO, COS, HeLa or BHK although transient expression may be preferred in *Xenopus* oocytes.

According to another aspect of the invention there is provided an antibody specific for an hVR protein as hereinbefore described or a variant thereof, preferably specific for hVR1 or hVR3 or a variant thereof.

According to another aspect of the invention there is provided a method for identification of a compound which exhibits hVR modulating activity, comprising contacting an hVR protein as hereinbefore described or a variant thereof, preferably hVR1 or hVR3 or a variant thereof, with a test compound and detecting modulating activity or inactivity.

According to another aspect of the invention there is provided a compound which modulates hVR activity, preferably that of hVR1 or hVR3, identifiable by the method referred to above.

According to another aspect of the invention there is provided a compound which modulates hVR activity, preferably that of hVR1 or hVR3, identifiable by the method referred to above, for use in therapy.

According to another aspect of the invention there is provided the use of a compound which modulates hVR activity, preferably that of hVR1 or hVR3, identifiable by the method referred to above, in the manufacture of a medicament for treatment or prophylaxis of a disorder which is responsive to the modulation of hVR activity, preferably hVR1 activity or hVR3 activity, in a human patient. Preferably the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), a respiratory disorder such as asthma or chronic obstructive pulmonary disease (COPD), a urological disorder such as neuropathy, incontinence or interstitial cystitis, or an inflammatory disorder.

According to another aspect of the invention there is provided a method of treatment or prophylaxis of a disorder which is responsive to modulation of hVR,

preferably hVR1 or hVR3, activity in a human patient which comprises administering to said patient an effective amount of a compound identifiable by the method referred to above. Preferably the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), respiratory disorders such as asthma and chronic obstructive pulmonary disease (COPD) and urological disorders including diabetic neuropathy, incontinence and interstitial cystitis and inflammatory disorders.

According to another aspect of the invention there is provided a compound which modulates hVR activity, preferably that of hVR1 or hVR3, identifiable by the method referred to above, excluding the compounds capsaicin, resiniferatoxin, piperine, zingerone, polydodial, warburganal, aframodial, cinnamodial, cinnamosmolide, cinnamolide, isovelleral, scalaradial, ancistrodial,  $\beta$ -acaridial, scutigeral, merulidial, anandamide and capsazepine.

According to another aspect of the invention there is provided a compound which modulates hVR activity, preferably that of hVR1 or hVR3, identifiable by the method referred to above, excluding the compounds capsaicin, resiniferatoxin, piperine, zingerone, polydodial, warburganal, aframodial, cinnamodial, cinnamosmolide, cinnamolide, isovelleral, scalaradial, ancistrodial,  $\beta$ -acaridial, scutigeral, merulidial, anandamide and capsazepine, for use in therapy.

According to another aspect of the invention there is provided the use of a compound which modulates hVR activity, preferably that of hVR1 or hVR3, identifiable by the method referred to above, excluding the compounds capsaicin, resiniferatoxin, piperine, zingerone, polydodial, warburganal, aframodial, cinnamodial, cinnamosmolide, cinnamolide, isovelleral, scalaradial, ancistrodial,  $\beta$ -acaridial, scutigeral, merulidial, anandamide and capsazepine, in the manufacture of a medicament for treatment or prophylaxis of a disorder which is responsive to the modulation of hVR activity, preferably hVR1 activity or hVR3 activity, in a human patient. Preferably the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic

5 pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), a respiratory disorder such as asthma or chronic obstructive pulmonary disease (COPD), a urological disorder such as neuropathy, incontinence or interstitial cystitis, or an inflammatory disorder.

10 According to another aspect of the invention there is provided a method of treatment or prophylaxis of a disorder which is responsive to modulation of hVR, preferably hVR1 or hVR3, activity in a human patient which comprises administering to said patient an effective amount of a compound identifiable by the method referred to above, excluding the compounds capsaicin, resiniferatoxin, piperine, zingerone, polydodial, warburganal, aframodial, cinnamodial, cinnamosmolide, cinnamolide, isovelleral, scalaradial, ancistrodial,  $\beta$ -acaridial, scutigeral, merulidial, anandamide and capsazepine. Preferably the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), respiratory disorders such as asthma and chronic obstructive pulmonary disease (COPD) and urological disorders including diabetic neuropathy, incontinence and interstitial cystitis and inflammatory disorders.

20 According to another aspect of the invention there is provided a compound identified by the method referred to above.

25 According to another aspect of the invention there is provided a compound identified by the method referred to above, for use in therapy.

30 According to another aspect of the invention there is provided the use of a compound identified by the method referred to above in the manufacture of a medicament for treatment or prophylaxis of a disorder which is responsive to the modulation of hVR activity, preferably hVR1 activity or hVR3 activity, in a human patient. Preferably the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), a respiratory disorder such as asthma or chronic

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obstructive pulmonary disease (COPD), a urological disorder such as neuropathy, incontinence or interstitial cystitis, or an inflammatory disorder.

According to another aspect of the invention there is provided a method of treatment or prophylaxis of a disorder which is responsive to modulation of hVR, preferably hVR1 or hVR3, activity in a human patient which comprises administering to said patient an effective amount of a compound identified by the method referred to above. Preferably the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), respiratory disorders such as asthma and chronic obstructive pulmonary disease (COPD) and urological disorders including diabetic neuropathy, incontinence and interstitial cystitis and inflammatory disorders.

According to another aspect of the invention there is provided a method of producing an hVR protein as hereinbefore described or a variant thereof, preferably hVR1 or hVR3 or a variant thereof, comprising introducing into an appropriate cell line a suitable vector comprising a nucleotide sequence encoding for an hVR protein or a variant thereof, preferably hVR1 or hVR3 or a variant thereof, under conditions suitable for obtaining expression of the hVR protein or a variant thereof, preferably hVR1 or hVR3 or a variant thereof.

#### **Brief Description of the figures**

Figure 1 is an alignment of hVR1 *in silico* derived clusters with rat VR1.

Figure 2 displays the human VR1 nucleotide sequence including the 5'UTR (nt - 773 to nt 0), coding region (nt 1 to 2517) and 3'UTR (nt 2518 to nt 3560).

Figure 3 illustrates the nucleotide and encoded amino acid sequence of the human VR1 sequence.

Figure 4 depicts the amino acid sequence of the hVR1 gene, the shading denotes predicted trans-membrane regions (boxed) and the ankyrin binding domains (unboxed). The predicted phosphorylation sites are underlined.

Figure 5 is a comparison of the amino acid sequences of the rat (rVR1) and human (hVR1) vanilloid receptors.



Figure 6 illustrates constructs pBluescriptSK(+) (A) and pCIN5-new (B) with the full length hVR1 gene cloned via NotI and EcoRI restriction sites.

Figure 7 shows a Slot Blot hybridisation with hVR1 probe with positive labelling of both rat and human DRG mRNA.

5 Figure 8 displays a Western blot probed with anti-VR1 antibodies with the arrow indicating the VR1 specific protein.

Figure 9 shows localisation of VR1 in rat DRG tissue sections, the arrow points to VR1 expressing small diameter (<25µm) neurone cell bodies.

Figure 10 depicts the *in situ* localisation of VR1 in human DRG sections (A) and human skin (B).

Figure 11 illustrates the functional response to capsaicin and blockade by capsazepine (CPZ) (A) with the current voltage relationship plotted in (B) on human VR-1 channels, transiently expressed in HEK293T cells.

Figure 12 shows capsaicin-induced desensitisation of human VR-1 channels in the presence of 2mM external calcium (A), maximum current (65mV) against time (B) and current voltage relationship in the absence of Ca<sup>2+</sup> (C).

Figure 13 shows the influx of calcium into transiently transfected HEK293T cells over a time course in the presence of agonist capsaicin, anandamide and resiniferatoxin in the absence (A, B, D and F) or presence (C, E, G) of the antagonist, capsezipine.

Figure 14 illustrates a graphical presentation the results shown in figure 13 examining the response of hVR1 transfected HEK293T cells over time before and after exposure to agonists: capsaicin, anandamide and resiniferatoxin in the absence (A, B, D and F) or presence (C, E, G) of the antagonist, capsezipine.

25 Figure 15 displays the proposed assay strategy to carry out drug screening.

Figure 16 displays an alignment of *in silico* derived hVR3 specific clusters with rat VR1.

Figure 17 depicts the hVR3 nucleotide sequence including the 5' UTR (nt -686 to nt 0) Coding region (nt1 to nt 2889), 3'UTR (nt 2890 to nt 3418).

30 Figure 18 shows the nucleotide and amino acid sequence of hVR3.

Figure 19 is of the amino acid sequence of hVR3, the shading denotes predicted trans-membrane regions (boxed) and the ankyrin binding domains (unboxed).

Figure 20 displays constructs pBluescriptSK(+) (A) and pCDNA3.1 (+) (B) with the full length hVR3 gene cloned via NotI and XhoI restriction sites.

Figure 21 illustrates a multiple comparison of the amino acid sequences of the rat VR1 and the human vanilloid receptors: hVR1, hVRL-1 and hVR3.

Figure 22 Northern Blot hybridisation with hVR3 probe with strong signals detected in trachea (A), prostate (B), placenta, kidney and pancreas (C).

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### Detailed Description of the Invention

Throughout the present specification and the accompanying claims the words "comprise" and "include" and variations such as "comprises", "comprising", "includes" and "including" are to be interpreted inclusively. That is, these words are intended to convey the possible inclusion of other elements or integers not specifically recited, where the context allows.

As referred to above, the present invention relates to isolated human vanilloid receptor (hVR) proteins, and in particular to the human vanilloid receptors which will be termed respectively human vanilloid receptors 1 and 3 (hVR1, and hVR3), sequence information for which is provided in figures 2 (hVR1) and 17 (hVR3). In the context of this invention the term "isolated" is intended to convey that the receptor protein is not in its native state, insofar as it has been purified at least to some extent or has been synthetically produced, for example by recombinant methods. The term "isolated" therefore includes the possibility of the receptor protein being in combination with other biological or non-biological material, such as cells, suspensions of cells or cell fragments, proteins, peptides, organic or inorganic solvents, or other materials where appropriate, but excludes the situation where the receptor protein is in a state as found in nature.

Routine methods, as further explained in the subsequent experimental section, can be employed to purify and/or synthesise the receptor proteins according to the invention. Such methods are well understood by persons skilled in the art, and include techniques such as those disclosed in Sambrook, J. *et al.* (28), the disclosure of which is included herein in its entirety by way of reference.

By the term "variant" what is meant throughout the specification and claims is that other peptides or proteins which retain the same essential character of the human vanilloid receptor proteins for which sequence information is provided, are also intended to be included within the scope of the invention. For example,

other peptides or proteins with greater than about 80%, preferably at least 90% and particularly preferably at least 95% homology with the sequences provided are considered as variants of the receptor proteins. Such variants may include the deletion, modification or addition of single amino acids or groups of amino acids within the protein sequence, as long as the peptide maintains the biological functionality of a human vanilloid receptor. This biological functionality can of course be assessed by conducting binding studies with known vanilloid modulators such as capsaicin, capsazepine (12) and resiniferatoxin (11).

Human VR1 is preferentially expressed in human dorsal root ganglia (DRG) and relative to hVR3 has the highest sequence homology with the rat VR1. Therefore, hVR1 is likely to be the human orthologue to rat VR1. hVR3 is less similar to rat VR1 and is expressed in a wider range of tissues. Nucleotide sequence analysis of hVR1 reveals a 2517bp open reading frame which encodes an 839 amino acid protein (see figures 2, 3 and 4). This deduced hVR1 protein sequence is 86 % identical to the rat VR1 (15) and shares many of its characteristics such as 6 transmembrane regions with an hydrophobic stretch between transmembrane 5 and 6 and an N-terminus which contains 3 ankyrin repeat domains. Similarly hVR3 has an open reading frame of 2889bp open reading frame which encodes a 963 amino acid protein (see figures 17, 18 and 19). The deduced hVR3 protein is 46 % identical to rat VR1 and 44 % identical to hVR1 sharing many of VR1's characteristics such as 6 transmembrane regions with an hydrophobic stretch between transmembrane 5 and 6 and an N-terminus which contains 3 ankyrin repeat domains.

The invention also includes nucleotide sequences which encode for human vanilloid receptor proteins or variants thereof as well as nucleotide sequences which are complementary thereto. Preferably the nucleotide sequence is a DNA sequence and most preferably, a cDNA sequence. Preferably the proteins are hVR1, hVR3 or variants thereof. Such nucleotides can be isolated or synthesised according to methods well know in the art. See reference 28, the disclosure of which is included herein in its entirety by way of reference.

The present invention also includes expression vectors which comprise nucleotide sequences encoding for the hVR, preferably hVR1 or hVR3, receptor

5 proteins or variants thereof. A further aspect of the invention relates to an expression vector comprising nucleotide sequences encoding for hVR1 or hVR3 receptor proteins or variants thereof. Such expression vectors are routinely constructed in the art of molecular biology and may for example involve the use of plasmid DNA and appropriate initiators, promoters, enhancers and other elements, such as for example polyadenylation signals which may be necessary, and which are positioned in the correct orientation, in order to allow for protein expression. Suitable vectors for use in practicing the present invention include pBluescript (Stratagene), pCR-Script (Stratagene), pCR2.1-TOPO (Invitrogen), pCRII-TOPO (Invitrogen), pCR-Blunt (Invitrogen), with vectors such as pCIN (32) (available from Clontech as pIRES-neo), pCDNA 3.1 (Invitrogen) or pCIneo (Promega) required for mammalian expression. Appropriate methods can be effected by following protocols described in many standard laboratory manuals (28, 29).

10 The invention also includes cell lines which have been modified to express the novel receptor. Such cell lines include transient, or preferably stable higher eukaryotic cell lines, such as mammalian cells or insect cells, lower eukaryotic cells, such as yeast or prokaryotic cells such as bacterial cells. Particular examples of cells which have been modified by insertion of vectors encoding for the receptor proteins according to the invention include HEK293T cells and *Xenopus* oocytes. Preferably the cell line selected will be one which is not only stable, but also allows for mature glycosylation and cell surface expression of the inventive receptor. Representative examples of appropriate hosts include animal cells such as HEK293, CHO, COS, HeLa and BHK.

15 It is also possible for the receptors of the invention to be transiently expressed in a cell line or on a membrane, such as for example in a baculovirus expression system. Such systems, which are adapted to express the receptors according to the invention, are also included within the scope of the present invention.

20 In particular, the functional hVR protein may include hVR receptor proteins selected from hVR1 and hVR3 and thereof or even other hVR protein subtypes or splice variants which have not yet been identified.

According to another aspect, the present invention also relates to antibodies, preferably monoclonal antibodies, which have been raised by standard techniques and are specific for the receptor proteins or variants thereof according to the invention. Such antibodies could for example be useful in purification; isolation or screening involving immuno precipitation techniques and may be used as tools to further elucidate hVR, preferably hVR1 or hVR3, protein function, or indeed as therapeutic agents in their own right. Antibodies may also be raised against specific epitopes of the receptors according to the invention.

An important aspect of the present invention is the use of receptor proteins according to the invention in screening methods designed to identify compounds which act as receptor ligands and which may be useful to modulate receptor activity. In general terms, such screening methods will involve contacting the receptor protein concerned, preferably hVR1 or hVR3, with a test compound and then detecting modulation in the receptor activity, or indeed detecting receptor inactivity, which results. For further details on the screening strategy refer to figure 15. The present invention also includes within its scope those compounds which are identified as possessing useful hVR, preferably hVR1 or hVR3, modulation activity, by the screening methods referred to above. The screening methods comprehended by the invention are generally well known to persons skilled in the art. High throughput screens may include fluorescence based assays using the Fluorometric Imaging Plate Reader (FLIPR) with calcium sensitive dyes, and reporter gene assays using calcium sensitive photoproteins that emit light on the influx of calcium and can be detected using an Imaging system. Secondary screens may involve electrophysiological assays utilising patch clamp technology to identify small molecules, antibodies, peptides, proteins or other types of compounds that interact with hVR, preferably hVR1 or hVR3, to modulate activity. Tertiary screens may involve the study of modulators in well characterised rat and mouse models of pain. These models of pain include, but are not restricted to, intraplantar injection of inflammatory agents such as carageenan, formalin and complete freunds adjuvant (CFA). Models of neuropathic pain such as loose ligature of the sciatic nerve are also included. Other screens may involve the study of modulators in human volunteers subject to topically applied capsaicin.

Another aspect of the present invention is the use of compounds which have been identified by screening techniques referred to above in the treatment or prophylaxis of disorders which are responsive to modulation of hVR, preferably hVR1 or hVR3, receptor activity, in a human patient. By the term "modulation" what is meant is that there will be either agonism or antagonism at the receptor site which results from ligand binding of the compound at the receptor. By the term "modulation" what is meant is that there will be either agonism or antagonism at the receptor site which results from ligand binding of the compound at the receptor excluding the compounds capsaicin, resiniferatoxin, piperine, zingerone, polydodial, warburganal, aframodial, cinnamodial, cinnamosmolide, cinnamolide, isovelleral, scalaradial, ancistrodial,  $\beta$ -acaridial, scutigeral, merulidial, anandamide and capsazepine. hVR, preferably hVR1 and hVR3, proteins have been implicated in disorders of the central nervous system (CNS), gastrointestinal (GI) tract, lungs and bladder and therefore modulation of hVR, preferably hVR1 or hVR3, receptor activity in these tissues will result in a positive therapeutic outcome in relation to such disorders. In particular, the compounds which will be identified using the screening techniques according to the invention will have utility for treatment and/or prophylaxis of disorders such as pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, IBS, respiratory disorders such as asthma and COPD, urological disorders including diabetic neuropathy, incontinence and interstitial cystitis, and inflammatory disorders. It is to be understood however, that the mention of such disorders is by way of example only, and is not intended to be limiting on the scope of the invention.

The compounds which are identified according to the screening methods outlined above may be formulated with standard pharmaceutically acceptable carriers and/or excipients as is routine in the pharmaceutical art, and as fully described in Remington's Pharmaceutical Sciences, Mack Publishing Company, Eastern Pennsylvania, 17th Ed, 1985, the disclosure of which is included herein in its entirety by way of reference.

The compounds may be administered via enteral or parenteral routes such as via oral, buccal, anal, pulmonary, intravenous, intraarterial, intramuscular, intraperitoneal, topical or other appropriate administration routes.

- 5 The present invention will be further explained, by way of examples, in the appended experimental section. Reference examples are provided.

### Experimental details

10 **Reference Example A: Identification of related human ESTs (Expressed Sequence Tags) (19) to the rat VR1 sequence by *in silico* analysis**

15 The full-length rat VR1 amino acid sequence (15) was used as a query sequence using the tBlastn (20) alignment program to identify related human genes in the dbEST (21) and Incyte (Incyte Pharmaceuticals, Inc., 3174 Porter Drive, Palo Alto, California 94304, USA) databases. Several human ESTs were identified and those with similarities greater than 50% selected for further analysis. One of these ESTs was T12251 previously shown to have 68% amino-acid identity and 84% similarity over a region of 70 amino acids (15). Full-length  
20 cloning and functional characterisation of the gene represented by this cluster has been completed (30). This gene was denoted hVRL-1 and encoded a protein of 764 amino acid protein that was 48 % identical to the rat VR1 protein. All human ESTs from both databases were clustered to identify overlapping identical ESTs belonging to the same transcript. The GCG package (Wisconsin Package Version 9.0, Genetics Computer Group (GCG), Madison, Wisconsin)  
25 and a program developed in house termed ESTBlast (22) were used to build up these clusters. In total, forty-three ESTs derived from different tissue sources and both EST databases were clustered into ten groups, one of these clusters represented hVRL-1. The remaining nine clusters have been named hVRa, hVRb, hVRc, hVRd, hVRe, hVRf, hVRg, hVRh and hVRi. For each EST the  
30 tissue source was assigned according to the annotations in the dbEST and Incyte databases. Since no obvious starting codon was present and the cluster sequences were shorter than the rat VR1 transcript none of these clusters were likely to represent a full-length vanilloid receptor transcript. Finally hVRg, hVRh and hVRi collapsed into a single contig. Sequence analysis has shown that  
35

these cDNAs are likely to be chimeric. The 5' end has weak similarities with the rat VR1 gene but the 3' end is identical to a DNA binding protein. No more work was pursued with that transcript.

5 **Reference Example B: Isolation of the human orthologue to the rat VR1 gene (reference examples B1-B4):**

**Reference Example B1: *In silico* assembly of human VR1**

10 The consensus nucleotide sequences from the ten clusters were searched with the tBlastx program (20) against the rat VR1 sequences to identify the most likely open reading frames. Frame shifts were corrected when the sequence trace files were available. Each cluster was aligned against the rat VR1 amino-acid sequence according to the Blastx results. The Blastx alignment program (20) was used to compare the full-length rat VR1 protein with the amino-acid sequences of the ten clusters. The three clusters with the highest homology, displayed in figure 1, were aligned with the rat VR1 gene.

20 Cluster hVRa shared a high homology (70% identity and 75% similarity over a stretch of 107 amino acids) with the 5' of the rat VR1 sequence but did not seem to have a potential start codon. It contained two ESTs (EST1 and EST2) derived from the same tissue, bladder, and from the same patient. These two ESTs were selected for further investigation since this cluster was the most 5', had high homology with rat VR1 and the bladder tissue could be contaminated with sensory neurones. Both cDNA clones were ordered but only clone EST1 was received as EST2 failed the recovery procedure.

30 Cluster hVRb composed of two EST's (EST3 and EST4), with 89% identity and 92% similarity over 90 residues, showed the highest degree of homology to the rodent sequence. The overlap between both sequences was located towards the middle of the gene.

35 hVRc (EST5) also while having high homology (71% identity and 75% similarity over 65 residues) with rat VR1 was closely related to the C-terminus of the rat protein sequence.



### Reference Example B2: Sequencing of clones

5 All DNA sequences were determined by automated DNA sequencing based on the dideoxy chain-termination method using the ABI 373A / 377 sequencers (Applied Biosystems). Sequence-specific primers were used with the 'Big-Dye' Terminator Cycle Sequencing kit (Applied Biosystems). The nucleotide sequence was analysed using programs from the University of Wisconsin Genetics Computer Group package.

10 More specifically when sequencing an EST clone, the following protocol was adopted. The EST1 clone was grown using standard procedures and DNA was isolated using Qiagen columns. SP6 (5' ATTTAGGTGACACTATAG) and T7 (5' TAATACGACTCACTATAGGG) primers flanking the cloning site were used to sequence both ends. Plasmid DNA (0.6 pmol) was used with 10.0 pmol of each primer for the dye terminator reaction. The SP6 end corresponded to the *in silico* derived EST sequence (identical to EST1). The T7 end did not have homologies with VR1 nor did it possess a long open reading frame or a polyadenylation motif. The size of the insert was determined by enzyme digestion of the DNA with the endonucleases NotI and EcoRI and calculated to be approximately 3kb.

15 Plasmid DNA (50ng) was used to amplify the insert by Polymerase Chain Reaction (PCR) with T7 and SP6 as primers. The PCR conditions included an initial hot-start at 94°C for 2 minutes, followed by 35 cycles at 94°C for 45 seconds, 50°C for 45 seconds and 72°C for 1 minute and terminated by 5 minutes at 72°C. The resulting PCR amplicon was separated on a 1.2% agarose gel and shown to be of ~3kb in size.

20 To fully sequence the PCR product the nuclease-Bal-31 technique was used where both strands of duplex DNA are degraded from both ends (23). After ethanol precipitation of the PCR product, the pellet was re-suspended in 30ml of 1X Bal-31 buffer (add buffer composition). A time-course digest with 2 units of Bal-31 enzyme (Roche Molecular Biochemicals) was carried out with 12 time points taken over 90 minutes (30 seconds, 1, 2, 3, 5, 7, 10, 15, 25, 45, 75 and 90 minutes). Three pools were made respectively from digests 1 to 4, 5 to 8 and 9

to 12. Each pool was blunt-ended and sub-cloned into the pCR-Script SK (+) plasmid from Stratagene at the SrfI site. After transformation, 16 colonies from each pool were screened by PCR with the flanking Reverse (5' GGAAACAGCTATGACCATG) and M13-20 (5' GTAAACGACGGCCAGT) primers. The amplicons of 6 positive colonies per pool were subjected to direct sequencing (24) using the T3 (5' AATTAACCCTCACTAAAGGG) and T7 primers. The DNA sequences obtained were assembled using the GCG package, translated and aligned against the rat VR1 gene using the Blast tools. After analysis, the 3079bp amplicon was shown to have 2 introns of 603bp and 1221bp. The latter intron was located at the 3'end of the PCR product. The coding sequence covered 1255 bp and was separated by the former intron. Therefore the clone EST1 was likely to be a partially spliced and incomplete cDNA.

The clone belonging to cluster 1b (EST3) and derived from a kidney cDNA library was ordered and sequenced using the Bal-31 technique. After assembly of the sequences using the GCG package an identical overlap was identified with the DNA sequence of the cluster hVRc. Moreover a 3'end with a polyadenylation signal and tail was identified. The complete sequence of the combined hVRb Bal-31 derived sequence and hVRc was 2063 bp (1020 bp of coding and 1043 bp of 3' untranslated sequence).

### Reference Example B3: Amplification of the middle section of hVR1 using the Polymerase Chain Reaction

We formulated the hypothesis that both sequences (hVRa and hVRb/c) were part of a common transcript. If the human and rat VR1 were going to be similar, the 2 contigs should be separated by a gap of approximately 275bp. Primers were designed on both sides of the gap to amplify mRNA from brain tissues in order to clone the gap. A smear was obtained with the sense primer (5' TCTACTTCGGTGAAGTGGCC) and antisense (5' ACGGCAGGGAGTCATTCTTC). For specificity 50ng of the PCR product were amplified with the nested sense (5' CTGCAGAACTCCTGGCAGA) and antisense (5' GTCACCACCGCTGTGGAAAA) primers. The 900bp nested amplicon was sequenced and shown to be identical to hVRa at one end and

hVRb/c at the other end. The middle part of the PCR product was homologous to the rat VR1 sequence. This region corresponded to 91 amino acids. When the sequences of hVRa, hVRb/hVRc and the internal amplicon are combined the total length of the Open Reading Frame (ORF) is 824 amino acids followed by a 3' untranslated sequence of 1043 bp. The human amino acid sequence is 87% identical to the rat sequence over that part of the coding region. This sequence was termed hVR1 because of its high degree of identity with the rat VR1 sequence.

#### **Reference Example B4: Isolation of the 5' Terminus of hVR1 by PAC isolation**

Since no start codon was identified at the 5' end an additional strategy was designed to identify the full-length sequence. Two primers, sense (5' TCCTCTGGCTTCCAACCCGTT) and antisense (5' GAACTGGGCAGAAAGTGCCT) were designed to amplify a 150bp product from the first intron mentioned in reference example B2. A P1 Artificial Chromosome (PAC) genomic clone (25) was isolated by PCR screening of a PAC library (Genome Systems, St Louis, Missouri). PAC DNA was recovered by using standard plasmid isolation protocol (26). An anti-sense primer was designed (5' CTGGAGTTAGGGTCTCCATCC) to sequence the genomic clone towards the potential 5' end of the gene. An open reading frame with a starting codon was identified. The gene structure was confirmed by using the GenScan software (27). The complete gene has a nucleotide sequence of 2517bp (figure 2) and encoded a 839 amino acid protein (Figures 3 and 4). The gene was named hVR1. Multiple alignment of the amino acid sequence of hVR1 and rat VR1 shows a remarkable degree of identity and similarities between both sequences (figure 5). The rVR1 and hVR1 amino acid sequences are 86% identical. Moreover after protein analysis 6 trans-membrane domains and 3 ankyrin binding domains were identified in hVR1 as in the rat VR1 gene.

#### **Example 1: Full-length Amplification of hVR1 from human DRG and assembly into cloning vectors**

HVR1 was PCR amplified in three sections from human DRG template. The 5' fragment was amplified using a sense primer encoding a NotI site and a strong

Kozak motif followed by gene specific sequence (5' GTCATAGCGGCCGCGCCGCCACCATGAAGAAATGGAGCAGCAC) and an antisense primer (5' AGGCCCACTCGGTGAACTTC). The thermo-cycling conditions used for this amplification included a hot start at 94°C for 4 mins, followed by 35 cycles of 94°C for 1 min, 54°C for 1 min and 72°C for 1 min. A final extension step of 72°C for 5 min completed the reaction. The resulting PCR products were separated on a 2% agarose gel and cloned into pCR<sup>®</sup>II-TOPO according to the manufacturers instructions supplied with the TOPO<sup>™</sup> TA Cloning<sup>®</sup> kit (Invitrogen). The middle section of hVR1 was PCR amplified using the sense primer: 5' GACGAGCATGTACAATGAGA and antisense primer: 5' GTCACCACCGCTGTGGAAAA. The cycling conditions included a hot start at 94°C for 4 mins, followed by 35 cycles of 1 min at 94°C, 56°C and 72°C. A final extension step of 72°C for 5 min completed the reaction. A band of approximately 870 bp was excised from a 2 % agarose gel and cloned as detailed by the TOPO<sup>™</sup> TA Cloning<sup>®</sup> kit into the vector pCR2.1<sup>®</sup>-TOPO. Finally the 3' end was PCR amplified with the sense primer: 5' TGTGGACAGCTACAGTGAGA and the antisense primer: 5'TGCACTGAATTCGAGCACTGGTGTTCCTCAG which encoded an EcoRI site for cloning. The PCR conditions included a 90 sec hot start at 94°C followed by 35 cycles of 94°C for 50 sec, 50°C for 50 sec and 72°C for 50 sec. The cycling was completed with a 72°C step for 5 min. PCR products were separated on a 2% agarose gel and cloned into the vector pCR2.1<sup>®</sup>-TOPO.

Resulting clones for each of the three hVR1-fragments were taken for sequence analysis and separate clones coding a consensus sequence were used in the full length assembly of the gene. The NotI/DraIII (New England Biolabs) digested 5' end fragment ligated together with the middle DraIII/EcoRI fragment into a NotI/EcoRI restricted pBluescript SK (+) vector (Stratagene). Finally, the remaining 3' fragment was introduced into the resulting construct via MscI and EcoRI restriction sites, a map of the resulting construct is displayed in figure 6A.

Several clones were selected for sequence analysis to confirm that constructs still encoded the hVR1 consensus sequence. These were then digested with NotI/EcoRI and ligated into the mammalian expression vector pCIN5-new (a modified version of pCIN1 (32) having an IVS deletion as well as a 36 bp

deletion repositioning the start codon of neomycin phosphotransferase immediately after the upstream EMVC IRES) as illustrated in figure 6B.

### Example 2: Chromosomal Localisation

The primers used to isolate the PAC clone (reference example B4) were selected for PCR on the G3 radiation hybrid panel from Stanford commercially available from Research Genetics (Huntsville, Alabama). The positive lanes and negative patterns were analysed using the public web server at Stanford University (<http://www-sghc.stanford.edu>). After analysis the hVR1 gene appears to be located on human chromosome 17 around marker SHGC-36073 (lod score=9.55).

### Example 3: mRNA Distribution

The tissue distribution of hVR1 was established by slot-blot hybridisation. RNA was transferred onto a sheet of GeneScreen hybridisation transfer membrane (DUPONT) sandwiched in a slot blotter by suction via a vacuum pump. Once the membrane was rinsed in 2x SSC (3M sodium chloride and 0.3M sodium citrate pH7) for 2 min it was exposed to UV using an Ultraviolet crosslinker (Amersham Life Science) for 1min at 15000uW/cm<sup>2</sup> thus enabling cross-linkage of the RNA onto the membrane. The amounts of RNA on the blot are unknown. The probe was obtained by PCR amplification of a 260 bp product of the coding region of hVR1 with the following two primers: 5' TGTGGACAGCTACAGTGAGA and 5' GTGGAAAACCCGAACAAGA. Membranes were hybridised for 4 hr shaking at 60°C in a 10% dextran sulphate, 1% SDS (sodium dodecyl sulphate) and 1M NaCl solution. The probe was labelled with [ $\alpha$ 32P]dCTP (Amersham) using the Rediprime™ DNA labelling system (Amersham), so as to obtain approximately 500,000cpm of the labelled probe per ml of prehybridisation solution. Briefly 100ng of probe was boiled for 3 minutes (denaturization) and then cooled on ice for 2 minutes in a total volume of 45 $\mu$ l. This was added to the labelling tube from the kit together with 3 $\mu$ l of 32P dCTP followed by an incubation at 37°C for 30 minutes. 400 $\mu$ l of Herring Sperm DNA (Sigma) at a concentration of 8 $\mu$ g/ml was added to the labelled probe and heated at 99°C for 3 minutes followed by rapid cooling on ice. The labelled probe was added and mixed well in pre-hybridisation solution. The membranes were hybridised overnight at 55°C.

The membranes were then washed, first at room temperature in 2xSSC and 1% SDS for 5 minutes, followed by 2x SSC and 1% SDS for 30 min at 50°C. If necessary further washes with 1x SSC and 0.5% SDS or 0.1xSSC and 0.1% for 30 mins at the same temperature were carried out. The membranes were then exposed to Scientific Imaging Film AR (Kodak) using intensifying screens at – 70°C overnight and the film developed.

The results are shown on figure 7. Strong signals were observed with the positive controls (slots 4B and 5B). Signals are detected on the human DRG slots (1A and 1B). No signals were detected with the water control (slot 3B). Three multi-tissue northern blots (Clontech) with a wide range of tissues have also been hybridised with the same probe, however no signals were detected. RT-PCR was performed on various tissues with the primer combination used to amplify the probe. A strong band was detected in DRG RNA. Taken together these hybridisations suggest that hVR1 is specifically expressed in neuronal tissue and DRG in particular.

#### **Example 4: Design and production of Anti-hVR1 Antibody**

The peptides CHFTTTRSRTRLFGKGDSEEASC (peptide68) and CGSLKPEDAEVFKDSMVPGEK (peptide69) were synthesised by standard solid phase techniques and purified by gel filtration chromatography. These peptides were conjugated via their Cys residues to the carrier protein, Tuberculin PPD (purified protein derivative) using sulpho-SMCC (sulfosuccinimidyl 4-[N-maleimidomethyl]-cyclohexan-1-carboxylate). Rabbits, previously sensitised to Bacillus Calmette Guerin (BCG), were inoculated with the resulting conjugates emulsified in incomplete Freund's adjuvant at approx monthly intervals. Serum was prepared from blood samples taken 7 days after each immunisation. The specific antibody response was followed by indirect enzyme-linked immunosorbent assay (ELISA) using free peptide as antigen. Immunoglobulins were purified from high titre sera using immobilised peptide affinity columns (sulpholink Pierce). Rabbits designated M143, 144 and 145 received peptide68 conjugate, rabbits M146, 147 and 148, peptide69 conjugate.

The antibodies have been validated by specific staining of the recombinant protein expressed in HEK293 cells. Whole cell lysates were prepared in Sample

Buffer (4 ml dH<sub>2</sub>O, 1 ml 0.5 M Tris-HCl, pH 6.8, 0.8 ml glycerol, 1.6 ml 10 % w/v SDS, 0.4 ml 2-β mercaptoethanol and 0.2 ml of 0.05 % w/v bromophenol blue) and proteins separated by SDS-PAGE and transferred to a nitrocellulose filter by electroblotting. Following incubation with the antisera, bound immunoglobulins were revealed using HRP-conjugated secondary antibodies and enhanced chemiluminescence (ECL) detection. The antisera showed specific binding to a protein(s) of the appropriate molecular weight(s) in extracts of VR1 transfected cells, but not in control extracts, this is illustrated in figure 8.

#### **Example 5: *Insitu* localisation of hVR1 using specific antibody**

The purified immunoglobulins have been used for immunohistochemical staining of rat DRG tissue sections. Fixed cryosections of DRG were incubated with antibodies for 48h at 4°C at concentrations between 0.1 to 0.5µg/ml. Following a washing step, bound antibodies were detected by indirect immunofluorescence. The antibodies recognised exclusively small diameter cell bodies of the peripheral sensory neurones as displayed in figure 9. This observation has been extended to human DRG tissues for the anti-peptide68 peptide antibodies demonstrating cross-reactivity with the human sequence as expected. Figure 10A demonstrates labelling of DRG cell bodies with an arrow that points to small diameter neuronal cell body) and in figure 10B the arrow points to labelled neurones innervating human skin.

#### **Example 6: Mammalian Cell Expression (examples 6a-6b)**

##### **Example 6a: Transient expression of hVR1 in mammalian cells**

HEK293 cells were plated onto a 6 well plate, containing poly-L-lysine coated coverslips, at  $5 \times 10^4$  cells per well. Next day, fresh media was added to the cells (50% confluent). CalPhos Mammalian Transfection Protocol (Clontech, K2051-1) was used for DNA transfection. For each well of cells, solution A was made up containing 8ug hVR1pCIN5, 2µg pEYFP-N1 reporter DNA, 12.4 µl calcium solution and water to 100µl. Solution B (hepes buffered saline) was slowly vortexed while solution A was added dropwise. The mixture was incubated at room temperature for 20 minutes, and then added to cells. The plate was slowly rocked to distribute the solution. The cells were incubated at 37°C for 5 hours, and then washed with phosphate buffered saline. Fresh culture

medium was added and the plate was incubated 24-48 hours for functional analysis.

**Example 6b: Stable expression of hVR1 in mammalian cells**

HEK293 cells were plated onto a 6 well plate at  $1 \times 10^5$  cells per well. Next day, fresh media was added to the cells (50% confluent). CalPhos Mammalian Transfection Protocol (Clontech, K2051-1) was used for DNA transfection. For each well of cells, solution A was made up containing 2 $\mu$ g hVR1pCIN5, 12.4 $\mu$ l 2M calcium solution and water to 100 $\mu$ l. Solution B (hepes buffered saline) was slowly vortexed while solution A was added dropwise. The mixture was incubated at room temperature for 20 minutes, and then added to cells. The plate was slowly rocked to distribute the solution. The cells were incubated at 37°C for 5 hours, and then washed with phosphate buffered saline. Fresh culture medium was added and the plate was incubated 48 hours at 37°C, 5% CO<sub>2</sub>. Cells were harvested into 100mm dishes in selection medium containing 800 $\mu$ g/ml geneticin. Cells were then incubated and fed at 4 day intervals. In total around 10 days selection is required for each single cell to multiply into a visible clone. Well-separated clones were each picked (with a gilson tip) into separate wells of a 96 well plate, containing maintenance medium (400 $\mu$ g/ml geneticin). Cells were expanded into flasks for freezing stocks and functional analysis. Stable cells may be plated at  $1 \times 10^5$  cells onto poly-L-lysine coated coverslips in 6 well plate, for calcium imaging next day.

**Example 7: Functional Analysis of hVR1(examples 7a-7c):**

**Example 7a: Electrophysiology using patch clamp methods**

The activation of human VR-1 channels transiently expressed in HEK293T cells by capsaicin was investigated. Cells grown on poly-L-lysine-coated glass coverslips were placed in a recording chamber (0.5ml) and superfused with extracellular solution (2ml min<sup>-1</sup>). The extracellular solution contained: NaCl (140mM), KCl (5mM), MgCl<sub>2</sub> (2mM), CaCl<sub>2</sub> (2mM), 4-(2-hydroxyethyl)-1-piperazineethanesulphonic acid (HEPES, 10mM) and glucose (10mM). The pH was adjusted to 7.4 with NaOH and osmolarity ranged from 310-320mOsm l<sup>-1</sup>. Patch pipettes (borosilicate glass) were pulled using a Sutter P-97 electrode puller. The pipettes were filled with an internal solution consisting of: CsCl



(140mM), ethylene glycol-bis( $\beta$ -aminoethyl ether) *N,N,N',N'*-tetra acetic acid Cs salt (Cs-EGTA, 5mM) and HEPES (10mM). The pH was adjusted to 7.25 using CsOH and the osmolarity ranged from 275-290 mOsm. When filled with this internal solution, patch electrodes had resistances of 2-5 M $\Omega$ . Currents were recorded using standard whole-cell voltage clamp recording techniques (31) at room temperature (21-23°C) using an Axopatch 200A amplifier and signals were sampled at 2 or 0.1 kHz. The majority of series resistance errors (80-85%) were minimized with compensation circuitry. Membrane potentials were not corrected for junction potentials (<4 mV). Voltage pulses and data collection were performed on-line using pClamp8 software (Axon Instruments) interfaced with amplifiers. Membrane potentials were maintained at -60mV between protocols.

Capsaicin or capsazepine (CPZ) were applied, using a 'fast-flow sytem', directly onto the recording cell (<1s to equilibrate). The effects of capsaicin were measured either by application during constant recording while holding the membrane potential at -60mV to elicit an inward current, or applying voltage ramps (-100 to +60mV) in the absence and presence of capsaicin. Similarly both these methods of recording currents evoked by the application of capsaicin were used to demonstrate the blockade by the antagonist (CPZ).

Figure 11A reveals that application of capsaicin (1  $\mu$ M), on human VR1 channels transiently expressed in HEK293T cells, produces an inward current when the membrane was held at a potential of -60mV. This response was abolished by 1 $\mu$ M CPZ and the blockade was partially reversible.

In the presence of 1  $\mu$ M capsaicin, voltage ramps (-100 to +70mV) produced a current-voltage relationship demonstrating a substantial outward rectification. Addition of 1 $\mu$ M CPZ completely blocked the current (figure 11B). Again, only partial recovery was observed, especially for the inward currents evoked by negative potentials.

Capsaicin-induced desensitisation of human VR-1 channels in the presence of 2mM external calcium is illustrated in figure 12. Voltage ramps (-100 to +70) were applied and the addition of capsaicin (1 $\mu$ M) evoked an outwardly rectifying current. Repeated additions of capsaicin resulted in a progressive 'rundown' in

the size of the response (figure 12A). Figure 12B shows a plot of the current elicited at a potential of +65mV against time illustrating the 'rundown' in current amplitude. Voltage ramps were applied every 20s and capsaicin added at 2min intervals for approximately 40s. By the 6th addition the current had reduced about 4-fold.

When the external calcium was replaced with 5mM EGTA the size of the current increased dramatically (figure 12C). However, when calcium was re-applied to the external solution, the current evoked by capsaicin (1 $\mu$ M) was approximately equivalent to that of the 6th addition shown in (figure 12A).

#### **Example 7b: Calcium Imaging with HEK293 expressing hVR1**

HEK293 cells expressing hVR1 transiently or stably, were plated onto poly-l-lysine coated cover slips at  $1 \times 10^5$  cells per well. They were analysed on the following day by calcium imaging (QuantiCell 700, Applied Imaging). On the day of experiment, WASH buffer was prepared by adding  $\text{CaCl}_2$  to extracellular medium (ECM) to a final concentration of 2mM, (ECM contains 125mM NaCl, 5mM KCl, 2mM  $\text{MgCl}_2$ , 0.5mM  $\text{NaH}_2\text{PO}_4$ , 5mM  $\text{NaHCO}_3$ , 10mM Hepes, 10mM glucose, 0.1% BSA, pH7.4). The calcium sensitive dye solution was prepared by adding 50 $\mu$ l 5% pluronic F-127 in DMSO (Molecular Probes) to a vial of fura2-AM (Molecular Probes). After mixing, 20 $\mu$ l of the fura2-AM solution was added to 10ml WASH. 1.5 ml was then added to cells, which were then incubated at 37°C for 30 minutes. The plate was washed three times with WASH. 1ml WASH was added and stored in dark. Agonists and antagonists were prepared in WASH at 5x their required assay concentrations. The reagents and assay temperature was kept at 37°C. For the transiently transfected cells, the YFP reporter DNA fluorescence (490nm excitation) was used to identify the transfected cells. Cells were initially imaged in 400 $\mu$ l WASH (or 300 $\mu$ l WASH plus 100 $\mu$ l antagonist e.g. capsazepine). After approximately 1 min, 100 $\mu$ l agonist (e.g. capsaicin, anadamide or resiniferatoxin) at 5 x the desired concentration was added to give final 1x concentration. A sequence of images (340/380nm excitation) were taken to monitor calcium influx response in cells before (30-60 secs), and after the addition of agonist (2-5 mins). Figure 13 displays time courses taken for each of the tests set up to look at the affect of the different agonists mentioned above in the presence or absence of the rat VR1 antagonist, capsazepine. The Imager

also plots graphs of respective calcium concentration (nM) versus time (seconds) as shown in figure 14. After the addition of agonist (e.g. capsaicin, indicated by the vertical arrow on graph), the cells expressing hVR1 are stimulated to influx calcium. This is shown by the appearance of peak on the trace. The peak height correlates with hVR1 expression level. Varying levels of expression is some times seen depending on which cells are selected for the graph. Similar experiments may be accomplished to examine the response of protons and heat.

#### **Example 7c: Use of a FLIPR assay with VR1**

FLIPR (Fluorometric Imaging Plate Reader) is a high throughput fluorescence-based drug discovery tool for functional cell analysis. Intracellular calcium is monitored with the calcium sensitive dye, fluo3-AM. HEK293 cells stably expressing rat VR1 were plated into a 96 well, poly-L-lysine treated FLIPR plate at  $3 \times 10^4$  cells per well. On the following day, the plate was processed for FLIPR. FBP buffer was prepared (15 $\mu$ M Probenecid (calcium ATPase pump blocker) in 1x FLIPR buffer (145mM NaCl, 5mM KCl, 1mM MgCl<sub>2</sub>, 2mM CaCl<sub>2</sub>, 10mM glucose, 20mM Hepes). FBP buffer pH was then adjusted to 7.4 with NaOH. 400 $\mu$ l DMSO was added to a vial of fluo3-AM (Cambridge Bioscience, F-1241). The fluo3-AM solution was incubated at 37°C for 10 min and vortexed. LOAD was prepared by adding 20 $\mu$ l of fluo3-AM solution and 20 $\mu$ l 20% pleuronic F-127 in DMSO (Cambridge Bioscience, P-3000) into 10 ml FBP. The 96 well plate containing cells was flicked off to remove cell medium. 100 $\mu$ l LOAD was added per well. Cells were then incubated at 37°C for 60 minutes. Capsaicin (a rVR1 agonist) and capsazepine (CPZ, a rVR1 antagonist) were prepared at 10x the desired final assay concentrations in FBP. The plate was flicked to remove LOAD from cells, and 180 $\mu$ l FBP was added per well. The FLIPR machine added 20 $\mu$ l capsaicin per well to give a final 1x concentration. Cells were monitored for 70 seconds after agonist addition. The FLIPR traces (fluorescence change (counts) versus time (seconds)) were produced for each well. Peaks indicate capsaicin-gated calcium influx, by cells expressing rVR1. The peak height correlates with the rVR1 expression level. To measure antagonism of the VR1 response 20 $\mu$ l 10x antagonist CPZ was added into wells to give a final 1x concentration. The plate was incubated for 15 minutes at room temperature prior reading in the FLIPR. The FLIPR traces recorded for each well show that the

peak heights are reduced in cells pre-incubated in CPZ. The same FLIPR assay may be used to monitor the response of human VR1 on exposure to agonists and antagonists.

5     **Example 8: Example of a screen using human VR1.**

FLIPR assay technology may be utilised to screen for hVR1 modulators according to the procedure described in figure 15. Human VR1 may be gated with protons, capsaicin or heat.

10    **Reference Example C: Identification and partial characterisation of additional human vanilloid receptors (reference examples C1-C3):**

15    **Reference Example C1: Identification and characterisation of a novel vanilloid-like receptor, hVR3**

ESTs belonging to the remaining clusters were characterised by *in silico* cloning (reference example A). The following clones were used during this process: - EST6/EST7 (hVRd), -EST8. (hVRe), - EST9/EST10. (hVRf). These EST clusters have been aligned with rat VR1 in figure 16, note that this diagram is not to scale.

20    **Reference Example C2: Sequencing of clones**

Further sequencing, as detailed in reference example B2, and *in silico* cloning, enabled clusters hVRd, hVRe and hVRf to collapse forming a single contig of 583 amino acids. This sequence was named hVR3 and has 49 % identity with the rat VR1 sequence. It was unlikely that this single contig was a full-length vanilloid receptor transcript as no obvious starting codon was present and it was shorter than the rat VR1 transcript.

25    **Reference Example C3: Identification of the 5' terminus of hVR3**

Two primers (sense primer 5' ATGCCACCAGCAGGGTTAC and antisense primer 5' TCTGCCAGGTTCAGCTG) designed to PCR amplify an amplicon stretching the 3' end of hVR3 and its 3'utr were used to isolate a genomic PAC clone (Genome Systems. St Louis, Missouri). The hVR3 specific PAC clone was then used as template to generate a library. This was achieved by sonicating 30    6µg of Qiagen purified PAC construct, size selecting fragmented DNA of 500-  
35

2000bp. These resulting fragments were then blunt ended and cloned into the vector pCR®-Blunt as detailed in the manufacturers protocol supplied with the Zero Blunt™ PCR cloning kit (Invitrogen). Clones were then sequenced (reference example B2) to identify the complete 5' end of the hVR3 transcript.

5 The full-length nucleotide sequence of the hVR3 gene is displayed in figure 17. Figure 18 illustrates both nucleotide and encoded amino acid sequence of the human VR1 and figure 19 depicts the amino acid sequence of the hVR3 gene with shaded regions denoting predicted trans-membrane regions (boxed) and the ankyrin binding domains (unboxed).

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**Example 9: Full-length Amplification of hVR3 from human kidney template**

Human kidney was used as a source of template for the PCR amplification of hVR3. Primers used for amplification were designed to isolate the gene in three fragments. Primers designed to isolate the 5' end included a sense primer encoding a NotI site and a strong Kozak motif followed by gene specific sequence (5' GTCATAGCGGCCGCGCGCCACCATGCCCAGGGTAGTTGGAC and antisense primer (5' CACCTCTTGTTGTCAGTGA). The PCR conditions used were a hot start at 94°C for 4 mins, followed by 35 cycles of 94°C for 1 min, 56°C for 1 min and 72°C for 1 min and finally one cycle at 72°C for 5 min. The resulting PCR products were separated on a 2% agarose gel and cloned into pCR®II-TOPO according to the manufacturers instructions supplied with the TOPO™ TA Cloning® kit (Invitrogen). The middle fragment was PCR generated using sense and antisense primers 5' CAAATCTGCGCATGAAGTTCCAG and 5' GCCACGAGAAGTTCCACGTAGTG respectively in the presence of 5% DMSO.

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PCR thermo-cycling required 35 cycles of 1 min at 94°C, 58°C and 72°C for successful amplification of the fragment which was then excised from a 2% agarose gel for cloning into the pCR®II-TOPO vector. Finally the 3' fragment was amplified with a sense primer 5' GCTGCTCCCATTCTTGCTGA and an antisense primer 5' TGCAGTCTCGAGAAATGAGTGGGCAGAGAAGC encoding a XhoI restriction site. This fragment was successfully amplified using a hot start at 94°C for 4 min followed by 35 cycles of 94°C for 50 sec, 48°C for 50 sec and 72°C for 2 min. The cycling was completed with a 72°C step for 5 min. The amplified fragment was excised from a 2% agarose gel and clone into the pCR®II-TOPO vector.

Resulting clones for each of the three PCR generated hVR3-fragments were taken for sequence analysis and separate clones coding a consensus sequence were used in the full-length assembly of the gene. The DraIII restriction site of the pBluescript SK (+) vector (Stratagene) was firstly abolished by digestion with DraIII followed by a blunt ending step using T<sub>4</sub> DNA polymerase (New England Biolabs). This modified vector was then restricted to enable the ligation of both a NotI/NcoI 5' fragment and NcoI/ EcoRI middle fragment. Finally, the remaining 3' fragment was introduced into the resulting construct via DraIII and XhoI sites (figure 20A).

Several clones were selected for sequence analysis to confirm that the constructs still encoded the hVR3 consensus sequence. These were then digested with NotI/XhoI and ligated into the mammalian expression vector pCDNA3.1 (+) (Invitrogen) as seen in figure 20B. The resulting hVR3 consensus sequence is shown in the multiple alignment along with the full-length sequence of hVR1 and the published hVRL-1 in figure 21.

#### **Example 10: Chromosomal localisation**

The 3' terminus, including the 3' UTR sequence of hVR3 was used to design two primers to amplify a product of 360 bp: sense primer 5' ATGCCACCAGCAGGGTTAC and antisense primer 5' TCTGCCAGGTTCCAGCTG. The G3 radiation hybrid panel from Stanford University (Research Genetics, Huntsville, Alabama) was screened by PCR. The positive and negative lanes were analysed using the public web server at Stanford University (<http://www-sghc.stanford.edu>). After analysis the hVR3 gene appears to be located on human chromosome 12 around markers D12S177E (lod score=15) and D12S1893 (lod score=14).

#### **Example 11: mRNA distribution**

The following primers (5' ACAAGAAGGCGGACATGCGG and 5' ATCTCGTGGCGGTTCTCAAT) were used to obtain a PCR product from the coding region of hVR3. This amplicon was used as a probe on multi-tissue northern blots, the protocol of which is detailed in example 3, to determine the tissue distribution of the gene (figures 22A, 22B and 22C). A transcript of approximately 3.8 kb was detected in the following tissues (the intensities of the

signals are indicated in brackets): trachea (very strong), kidney (strong), pancreas (strong), prostate (strong), placenta (strong), bone marrow (weak), adrenal gland (weak), lymph node (weak), spinal cord (weak), thyroid (weak), stomach (weak), lung (weak) and liver (weak).

Since these commercial blots (Clontech, Palo Alto, California, USA) should have the same amount of RNA it is interesting to note the very strong signal in the trachea lane (figure 22A). This could indicate the potential of hVR3 as a target for respiratory pathologies. It was shown by RT-PCR with the primer combination used to produce the probe that the gene is not expressed in DRG.

#### **Example 12: Riboprobe generation for the in situ localisation of hVR3**

The same probe, which was specific to hVR3 in Northern blot analysis (example 11), was used to generate a riboprobe. This hVR3 specific probe was cloned into the T7 and SP6 encoding pCRII®-TOPO vector (Invitrogen). This construct was then used in the *in vitro* transcription of DIG labelled RNA strands from the vectors promoters as described in the manufacturers instructions as detailed in the DIG RNA labelling kit (Roche Molecular Biochemicals). This riboprobe may be used to identify the cellular localisation of hVR3 present in tissues such as trachea, lung, pancreas, prostate, placenta and kidney.

#### **Example 13: Mammalian Cell Expression of hVR3**

Expression of hVR3 may be accomplished by transfecting a mammalian cell line such as: HEK283T, HEK293, CHO, COS, HeLa and BHK. A detailed method for both transient and stable transfection is detailed in example 6.

#### **Example 14: Functional Analysis of hVR3**

The functional analysis of hVR3 may be studied using the electrophysiology, calcium imaging and FLIPR methods as detailed in examples 7a to 7c.

#### **Example 15: Example of a drug screen using human VR3.**

A stable cell line expressing hVR3 may be used in a drug screen such as a selectivity screen using test compounds that have been identified to have an agonistic or antagonistic action on hVR1. FLIPR assay technology may be utilised to screen for hVR3 modulators as proposed in figure 15.

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CRF Errors Corrected by the STIC Systems Branch

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- ☐ Changed a file from non-ASCII to ASCII
- ☐ Changed the margins in cases where the sequence text was "wrapped" down to the next line.
- ☐ Edited a format error in the Current Application Data section, specifically: \_\_\_\_\_
- ☐ Edited the Current Application Data section with the actual current number. The number inputted by the applicant was ☐ the prior application data; or ☐ other \_\_\_\_\_
- ☐ Added the mandatory heading and subheadings for "Current Application Data".
- ☐ Edited the "Number of Sequences" field. The applicant spelled out a number instead of using an integer
- ☐ Changed the spelling of a mandatory field (the headings or subheadings), specifically: \_\_\_\_\_
- ☐ Corrected the SEQ ID NO when obviously incorrect. The sequence numbers that were edited were: \_\_\_\_\_
- ☐ Inserted or corrected a nucleic number at the end of a nucleic line. SEQ ID NO's edited: \_\_\_\_\_
- ☐ Corrected subheading placement. All responses must be on the same line as each subheading. If the applicant placed a response below the subheading, this was moved to its appropriate place.
- ☐ Inserted colons after headings/subheadings. Headings edited included: \_\_\_\_\_
- ☐ Deleted extra, invalid, headings used by an applicant, specifically: \_\_\_\_\_
- ☒ Deleted: ☒ non-ASCII "garbage" at the beginning/end of files; ☐ secretary initials/filename at end of file.  
☐ page numbers throughout text; ☐ other invalid text, such as \_\_\_\_\_
- ☐ Inserted mandatory headings, specifically: \_\_\_\_\_
- ☐ Corrected an obvious error in the response, specifically: \_\_\_\_\_
- ☐ Edited identifiers where upper case is used but lower case is required, or vice versa.
- ☐ Corrected an error in the Number of Sequences field, specifically: \_\_\_\_\_
- ☐ A "Hard Page Break" code was inserted by the applicant. All occurrences had to be deleted.
- ☐ Deleted *ending* stop codon in amino acid sequences and adjusted the "(A)Length:" field accordingly (error due to a PatentIn bug). Sequences corrected: \_\_\_\_\_
- ☐ Other: \_\_\_\_\_

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## RAW SEQUENCE LISTING

DATE: 10/11/2001

PATENT APPLICATION: US/09/857,123

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## RAW SEQUENCE LISTING

DATE: 10/11/2001

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DATE: 10/11/2001

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DATE: 10/11/2001

TIME: 10:54:47

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L:13 M:270 C: Current Application Number differs, Replaced Application Number  
L:14 M:271 C: Current Filing Date differs, Replaced Current Filing Date

Country	Year	Population (millions)	Urban population (millions)	Urban population (%)	Population density (per sq km)	Urban population density (per sq km)	Population growth rate (%)	Urban population growth rate (%)	Population growth rate (%)	Urban population growth rate (%)	Population growth rate (%)	Urban population growth rate (%)
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Algeria	1985	10.5	4.5	42.9	105.0	262.5	1.8	3.0	1.8	3.0	1.8	3.0
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Algeria	1995	11.5	5.5	47.8	115.0	287.5	2.4	4.0	2.4	4.0	2.4	4.0
Algeria	2000	12.0	6.0	50.0	120.0	300.0	2.7	4.5	2.7	4.5	2.7	4.5
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PCT09

## RAW SEQUENCE LISTING

DATE: 08/30/2001

PATENT APPLICATION: US/09/857,123

TIME: 07:44:20

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Does Not Comply  
Corrected Diskette Needed

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 5 Tate, Simon N  
 6 Delany, Natalie S  
 7 Sanseau, P  
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DATE: 08/30/2001

TIME: 07:44:21

Output Set: N:\CRF3\08302001\I857123.raw

L:14 M:271 C: Current Filing Date differs, Replaced Current Filing Date

L:1772 M:254 E: No. of Bases conflict, LENGTH:Input:1 Counted:20 SEQ:40

[illegible]

Claims

1. An isolated human vanilloid receptor (hVR) protein or a variant thereof.
- 5 2. An isolated human vanilloid receptor (hVR) protein according to claim 1 which is hVR1 or a variant thereof.
3. An isolated human vanilloid receptor (hVR) protein according to claim 1 which is hVR3 or a variant thereof.
- 10 4. An isolated human vanilloid receptor (hVR) protein according to claim 2 having an amino acid sequence as shown in Figure 3.
- 15 5. An isolated human vanilloid receptor (hVR) protein according to claim 3 having an amino acid sequence as shown in Figure 18.
- 20 6. A nucleotide sequence encoding a human vanilloid receptor (hVR) protein or a variant thereof, or a nucleotide sequence which is complementary thereto.
7. A nucleotide sequence according to claim 6 encoding for an hVR1 protein or a variant thereof, or a nucleotide sequence which is complementary thereto.
- 25 8. A nucleotide sequence according to claim 6 encoding for an hVR3 protein or a variant thereof, or a nucleotide sequence which is complementary thereto.
- 30 9. A nucleotide sequence according to claim 6 which is a cDNA sequence.
10. A nucleotide sequence according to claim 7 which is a cDNA sequence
11. A nucleotide sequence according to claim 8 which is a cDNA sequence

12. A nucleotide sequence according to claim 7 as shown in Figure 2.
13. A nucleotide sequence according to claim 8 as shown in Figure 17.
14. An expression vector comprising a nucleotide sequence according to any one of claims 6 to 13, which is capable of expressing an hVR protein or a variant thereof.
15. An expression vector according to claim 14 which is capable of expressing an hVR1 protein or a variant thereof.
16. An expression vector according to claim 14 which is capable of expressing an hVR3 protein or a variant thereof.
17. A stable cell line comprising an expression vector according to claim 14.
18. A stable cell line comprising an expression vector according to claim 15.
19. A stable cell line comprising an expression vector according to claim 16.
20. A stable cell line according to claim 17 which is a modified HEK293, CHO, COS, HeLa or BHK cell line.
21. A stable cell line according to claim 18 which is a modified HEK293, CHO, COS, HeLa or BHK cell line.
22. A stable cell line according to claim 19 which is a modified HEK293, CHO, COS, HeLa or BHK cell line.
23. An antibody specific for a human vanilloid receptor (hVR) protein or a variant thereof as claimed in any one of claims 1 to 5.

24. An antibody according to claim 23 which is specific for hVR1 or a variant thereof.

5 25. An antibody according to claim 23 which is specific for hVR3 or a variant thereof.

10 26. A method for identification of a compound which exhibits hVR modulating activity comprising contacting a human vanilloid receptor (hVR) protein or a variant thereof according to any one of claims 1 to 5 with a test compound and detecting modulating activity or inactivity.

15 27. A compound which modulates hVR activity, identifiable by a method according to claim 26.

28. A compound according to claim 27 for use in therapy.

20 29. The use of a compound according to claim 27 in the manufacture of a medicament for treatment or prophylaxis of a disorder which is responsive to the modulation of hVR activity in a human patient.

25 30. The use according to claim 28 wherein the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algnesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowl syndrome (IBS), a respiratory disorder, asthma, chronic obstructive pulmonary disease (COPD), a urological disorder, neuropathy, incontinence, interstitial cystitis or an inflammatory disorder.

30 31. A method of treatment or prophylaxis of a disorder which is responsive to modulation of hVR activity in a human patient which comprises administering to said patient an effective amount of a compound according to claim 27.

35 32. A method according to claim 31 wherein the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain,

5 rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowl syndrome (IBS), a respiratory disorder, asthma, chronic obstructive pulmonary disease (COPD), a urological disorder, neuropathy, incontinence, interstitial cystitis or an inflammatory disorder.

10 33. A compound which modulates hVR activity, identifiable by a method according to claim 26, excluding the compounds capsaicin, resiniferatoxin, piperine, zingerone, polydodial, warburganal, aframodial, cinnamodial, cinnamosmolide, cinnamolide, isovelleral, scalaradial, ancistrodial,  $\beta$ -acaridial, scutigeral, merulidial, anandamide and capsazepine.

15 34. A compound according to claim 33 for use in therapy.

35. The use of a compound according to claim 33 in the manufacture of a medicament for treatment or prophylaxis of a disorder which is responsive to the modulation of hVR activity in a human patient.

20 36. The use according to claim 35 wherein the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowl syndrome (IBS), a respiratory disorder, asthma, chronic obstructive pulmonary disease (COPD), a urological disorder, neuropathy, incontinence, interstitial cystitis or an  
25 inflammatory disorder.

30 37. A method of treatment or prophylaxis of a disorder which is responsive to modulation of hVR activity in a human patient which comprises administering to said patient an effective amount of a compound according to claim 33.

35 38. A method according to claim 37 wherein the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowl syndrome (IBS), a respiratory disorder, asthma, chronic obstructive pulmonary disease (COPD), a



urological disorder, neuropathy, incontinence, interstitial cystitis or an inflammatory disorder.

39. A compound identified by the method according to claim 26.

40. A compound according to claim 39 for use in therapy.

41. The use of a compound according to claim 39 in the manufacture of a medicament for treatment or prophylaxis of a disorder which is responsive to the modulation of hVR activity in a human patient.

42. The use according to claim 41 wherein the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowl syndrome (IBS), a respiratory disorder, asthma, chronic obstructive pulmonary disease (COPD), a urological disorder, neuropathy, incontinence, interstitial cystitis or an inflammatory disorder.

43. A method of treatment or prophylaxis of a disorder which is responsive to modulation of hVR activity in a human patient which comprises administering to said patient an effective amount of a compound according to claim 39.

44. A method according to claim 43 wherein the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowl syndrome (IBS), a respiratory disorder, asthma, chronic obstructive pulmonary disease (COPD), a urological disorder, neuropathy, incontinence, interstitial cystitis or an inflammatory disorder.

45. A method of producing an hVR protein or a variant thereof according to any one of claims 1-5 comprising introducing into an appropriate cell line a suitable vector comprising a nucleotide sequence encoding for an hVR protein or

a variant thereof, under conditions suitable for obtaining expression of the hVR protein or variant thereof.

46. A method of producing an hVR1 protein or a variant thereof comprising introducing into an appropriate cell line a suitable vector comprising a nucleotide sequence encoding for an hVR1 protein or a variant thereof, under conditions suitable for obtaining expression of the hVR1 protein or variant thereof.

47. A method of producing an hVR3 protein or a variant thereof comprising introducing into an appropriate cell line a suitable vector comprising a nucleotide sequence encoding for an hVR3 protein or a variant thereof, under conditions suitable for obtaining expression of the hVR3 protein or variant thereof.

48. A human vanilloid receptor (hVR) protein or a variant thereof for use in a method of screening for agents useful in the treatment or prophylaxis of a disorder which is responsive to the modulation of hVR activity in a human patient

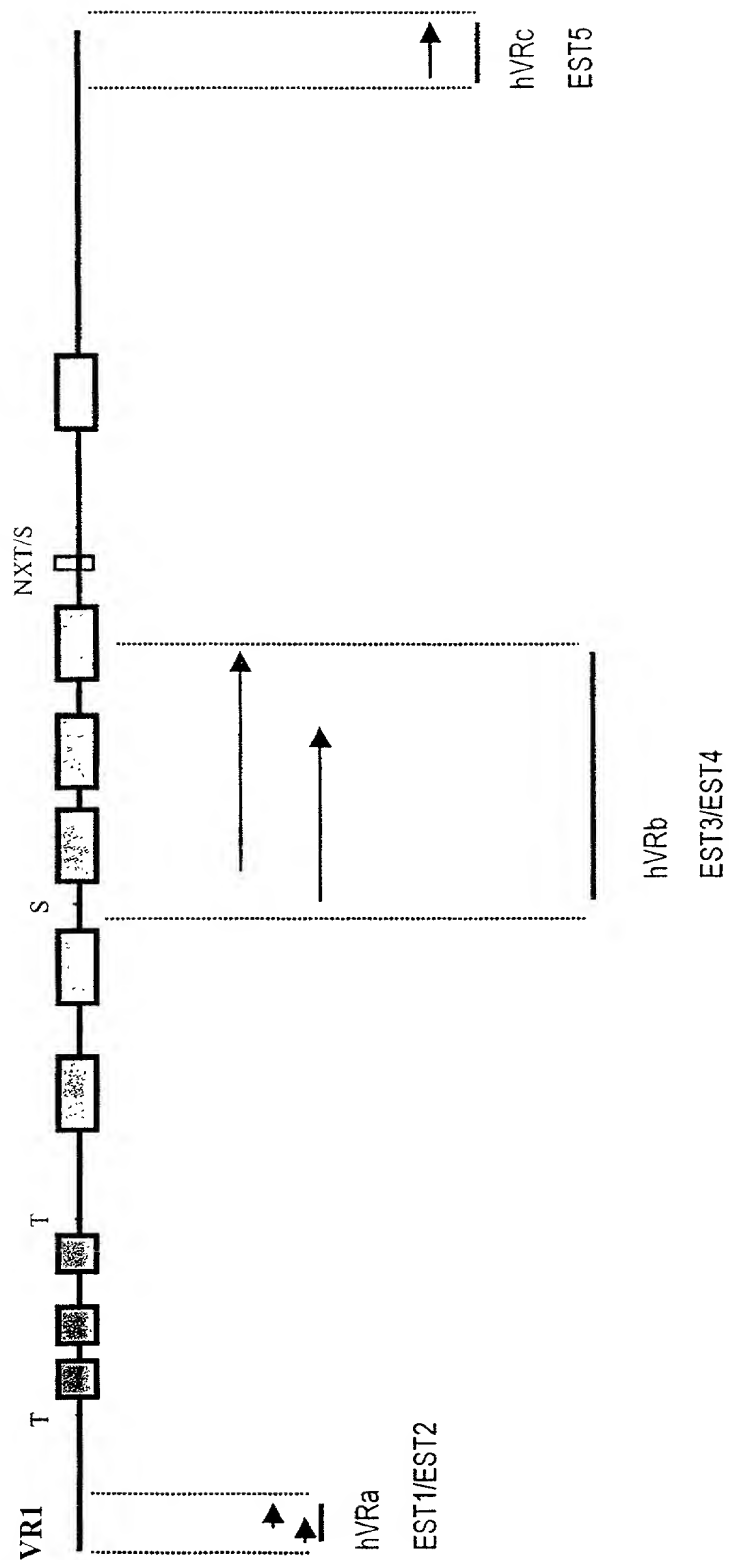
49. A human vanilloid receptor (hVR) protein according to claim 48 wherein the disorder is pain, neuropathic pain, inflammatory pain, chronic pain, post-operative pain, rheumatoid arthritic pain, neuropathies, neuralgia, algesia, neurodegeneration, nerve injury, stroke, ischaemia migraine, irritable bowel syndrome (IBS), a respiratory disorder, asthma, chronic obstructive pulmonary disease (COPD), a urological disorder, neuropathy, incontinence, interstitial cystitis or an inflammatory disorder.

50. A human vanilloid receptor (hVR) protein according to claim 48 or 49 which is hVR1 or a variant thereof.

51. A human vanilloid receptor (hVR) protein according to claim 48 or 49 which is hVR3 or a variant thereof.

FIG. 1

ALIGNMENT OF HUMAN VR1 IN SILICO DERIVED CLUSTERS WITH RAT VR1



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## FIG. 2

hVR1 SEQUENCE INCLUDING THE 5'UTR (nt -773 TO nt 0), CODING  
REGION (nt 1 TO 2517) AND 3'UTR (nt 2518 TO nt 3560)

-773 cccccagccacacacacacacgcacacacatacacacacacacacaggcttaaccattca -714  
-713 aaggccagaagcttgacagatggtgattcataaaaatgcaaaagccaaaatccaaaatct -654  
-653 tgtataagctcagtggtgtggcagcgaggttgaagagcaaaggcaggccggggcacctgg -594  
-593 ctgatgatgtgtggaccggttgacagcagggcccgagtgcggtgtgggtgtgggtggg -534  
-533 ccagtctctgcgctcaccctattccagggaacacagtccttggtcttcttgactgag -474  
-473 ccattctcatcacagagatcctcctgaattcagcccacgacagccaccccgccggtttt -414  
-413 ccttggtctgtgtgggaaggaggcagcgcggtggttatcaacctcaccctgcagaggag -354  
-353 gcacctgaggcccagagacgaggaggatgggtctaaccagaaccacagatgggtctga -294  
-293 gccggggggcctgtccaccctcccaggccgacgtcagtgccgcaggactgcctgggcct -234  
-233 gctaggcctgtctacaccttgaggcctctggggtgagaggttcagtcctggaaacacttca -174  
-173 gttctagggggctgggggcagcagcaagttggagttttggggtaccctgcttcacagggc -114  
-113 ccttggcaaggaggggcaggtggggtctaaggacaagcagtccttactttgggagtcaacc -54  
-53 ccggcggtggtggctgctgcaggttgacactggggccacagaggatccagcaaggATGAAG 6  
7 AAATGGAGCAGCACAGACTTGGGGGCAGCTGCGGACCCACTCCAAAAGGACACCTGCCCA 66  
67 GACCCCCTGGATGGAGACCCTAACTCCAGGCCACCTCCAGCCAAGCCCCAGCTCTCCACG 126  
127 GCCAAGAGCCGCACCCGGCTCTTTGGGAAGGGTGACTCGGAGGAGGCTTTCCCGGTGGAT 186  
187 TGCCCTCACGAGGAAGGTGAGCTGGACTCCTGCCCCGACCATCACAGTCAGCCCTGTTATC 246  
247 ACCATCCAGAGGCCAGGAGACGGCCCCACCGGTGCCAGGCTGCTGTCCAGGACTCTGTC 306

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307 GCCGCCAGCACCCGAGAAGACCCCTCAGGCTCTATGATCGCAGGAGTATCTTTGAAGCCGTT 366

367 GCTCAGAATAACTGCCAGGATCTGGAGAGCCTGCTGCTCTTCCTGCAGAAGAGCAAGAAG 426

427 CACCTCACAGACAACGAGTTCAAAGACCCTGAGACAGGGAAGACCTGTCTGCTGAAAGCC 486

487 ATGCTCAACCTGCACGACGGACAGAACACCACCATCCCCCTGCTCCTGGAGATCGCGCGG 546

547 CAAACGGACAGCCTGAAGGAGCTTGTCAACGCCAGCTACACGGACAGCTACTACAAGGGC 606

607 CAGACAGCACTGCACATCGCCATCGAGAGACGCAACATGGCCCTGGTGACCCTCCTGGTG 666

667 GAGAACGGAGCAGACGTCCAGGCTGCGGCCCATGGGGACTTCTTTAAGAAAACCAAAGGG 726

727 CGGCCTGGATTCTACTTCGGTGAAGTGGCCCTGTCCCTGGCCGCGTGACCAACCAGCTG 786

787 GGCATCGTGAAGTTCCTGCTGCAGAACTCCTGGCAGACGGCCGACATCAGCGCCAGGGAC 846

847 TCGGTGGGCAACACGGTGTGCACGCCCTGGTGGAGGTGGCCGACAACACGGCCGACAAC 906

907 ACGAAGTTTGTGACGAGCATGTACAATGAGATTCTGATCCTGGGGGCCAAACTGCACCCG 966

967 ACGCTGAAGCTGGAGGAGCTCACCAACAAGAAGGGAATGACGCCGCTGGCTCTGGCAGCT 1026

1027 GGGACCGGGAAGATCGGGGTCTTGGCCTATATTCTCCAGCGGGAGATCCAGGAGCCCGAG 1086

1087 TGCAGGCACCTGTCCAGGAAGTTCACCGAGTGGGCCTACGGGCCCGTGCACTCCTCGCTG 1146

1147 TACGACCTGTCCTGCATCGACACCTGCGAGAAGAACTCGGTGCTGGAGGTGATCGCCTAC 1206

1207 AGCAGCAGCGAGACCCCTAATCGCCACGACATGCTCTTGGTGGAGCCGCTGAACCGACTC 1266

1267 CTGCAGGACAAGTGGGACAGATTCGTCAAGCGCATCTTCTACTTCAACTTCCTGGTCTAC 1326

1327 TGCCTGTACATGATCATCTTCACCATGGCTGCCTACTACAGGCCCGTGGATGGCTTGCCT 1386

1387 CCCTTTAAGATGGAAAAAATTGGAGACTATTTCCGAGTTACTGGAGAGATCCTGTCTGTG 1446

FIG. 2<sub>CONT'D</sub>

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1447 TTAGGAGGAGTCTACTTCTTTTTCCGAGGGATTTCAGTATTTCTGTCAGAGGCGGCCGTCTG 1506

1507 ATGAAGACCCCTGTTTGTGGACAGCTACAGTGAGATGCTTTTCTTTCTGCAGTCACTGTTC 1566

1567 ATGCTGGCCACCGTGGTGCTGTACTTCAGCCACCTCAAGGAGTATGTGGCTTCCATGGTA 1626

1627 TTCTCCCTGGCCTTGGGCTGGACCAACATGCTCTACTACACCCGCGGTTTCCAGCAGATG 1686

1687 GGCATCTATGCCGTCATGATAGAGAAGATGATCCTGAGAGACCTGTGCCGTTTCATGTTT 1746

1747 GTCTACATCGTCTTCTTGTTCGGGTTTTCCACAGCGGTGGTGACGCTGATTGAAGACGGG 1806

1807 AAGAATGACTCCCTGCCGTCTGAGTCCACGTCGCACAGGTGGCGGGGGCCTGCCTGCAGG 1866

1867 CCCCCGATAGCTCCTACAACAGCCTGTACTCCACCTGCCTGGAGCTGTTCAAGTTCACC 1926

1927 ATCGGCATGGGCGACCTGGAGTTCACTGAGAACTATGACTTCAAGGCTGTCTTCATCATC 1986

1987 CTGCTGCTGGCCTATGTAATTCTCACCTACATCCTCCTGCTCAACATGCTCATCGCCCTC 2046

2047 ATGGGTGAGACTGTCAACAAGATCGCACAGGAGAGCAAGAACATCTGGAAGCTGCAGAGA 2106

2107 GCCATCACCATCCTGGACACGGAGAAGAGCTTCCTTAAGTGCATGAGGAAGGCCTTCCGC 2166

2167 TCAGGCAAGCTGCTGCAGGTGGGGTACACACCTGATGGCAAGGACGACTACCGGTGGTGC 2226

2227 TTCAGGGTGGACGAGGTGAACTGGACCACCTGGAACACCAACGTGGGCATCATCAACGAA 2286

2287 GACCCGGGCAACTGTGAGGGCGTCAAGCGCACCTGAGCTTCTCCCTGCGGTCAAGCAGA 2346

2347 GTTTCAGGCAGACACTGGAAGAACTTTGCCCTGGTCCCCCTTTTAAGAGAGGCAAGTGCT 2406

2407 CGAGATAGGCAGTCTGCTCAGCCCGAGGAAGTTTATCTGCGACAGTTTTTCAGGGTCTCTG 2466

2467 AAGCCAGAGGACGCTGAGGTCTTCAAGAGTCCTGCCGCTTCCGGGAGAAGtgaggacgt 2526

2527 cacgcagacagcactgtcaacactgggccttaggagaccccggtgccacggggggctgct 2586

FIG.2<sub>CONT'D</sub>

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2587 gagggaaacaccagtgtctgtcagcagcctggcctgggtctgtgcctgccagcatgttcc 2646  
2647 caaatctgtgtctggacaagctgtgggaagcgttcttgaagcatggggagtgtgtacat 2706  
2707 ccaaccgtcactgtccccaagtgaatctcctaacagactttcagggtttttactcacttta 2766  
2767 ctaaacagtttggttggtcagtctctactgggacatgttaggcccttgttttctttgatt 2826  
2827 ttattcttttctgtgagacagagttcactcttgttgcccaggctggagtgcagtgggtgtg 2886  
2887 atcttggtcactgcaacctctgtctcccggttcaagcgattcttctgtctcagttctccc 2946  
2947 aagtagcttggttacagggtgagcactaccacgcccggctaatttttgatttttaatag 3006  
3007 agacgggggtttcaccatgttggccaggctggtctcgaactcttgacctcagggtgatctgc 3066  
3067 ccgccttggcctcccaaagtgtctgggattacagggtgtgagccgctgcgctcggccttctt 3126  
3127 tgattttatattattagggagcaaaagttaaataagcccaggaaaacacctttgggaacaa 3186  
3187 actcttcctttgatggaaaatgcagaggcccttcctctctgtgcctgtgttgcctctt 3246  
3247 acctgcccgggtggtttgggggtgttggtgtttcctccctggagaagatgggggaggctg 3306  
3307 tccactcccagctctggcagaatcaagctgttgagcagtgcttcttcatccttctt 3366  
3367 acgatcaatcacagtctccagaagatcagctcaattgctgtgcagggttaaaactacagaa 3426  
3427 ccacatcccaaagggtacctggttaagaatgtttgaaagatcttccatttctaggaacccca 3486  
3487 gtctgtcttctccgcaatggcacatgcttccactccatccatactggcatcctcaaataa 3546  
3547 acagatatgtatacaaaaaaaaaaaaaaaaaaaaaaaaaaaaaa 3591

FIG. 2<sub>CONT'D</sub>

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## FIG. 3

NUCLEOTIDE AND AMINO ACID SEQUENCE OF hVR1 INCLUDING  
THE 5'UTR (nt -773 TO nt 0), CODING REGION (nt TO 2517) AND  
3'UTR (nt 2518 TO nt 3560)

-773	ccccagccacacacacacacgcacacacatacacacacacacacaggttaaccattca	-714
-713	aaggccagaagcttgacagatgttgattcataaaaaatgcaaaagccaaaatccaaaatct	-654
-653	tgtataagctcagtggctgtggcagcgaggttgaagagcaaaggcaggccgggcacctgg	-594
-593	ctgatgatgtgtggacccgttgacagcagggcccgagtgcggtgtgggtgtgggtggg	-534
-533	ccagtctctgccgctcacctattccagggacacagtcctgcttggtctctctggactgag	-474
-473	ccatcctcatcacogagatcctccctgaattcagccacgacagccaccccgccggtttt	-414
-413	ccttgttctgtgtgggaaggaggagcgcggtggttatcaacctcacctgcagaggag	-354
-353	gcacctgaggcccagagacgaggagggatgggtctaaccacagaaccacagatggctctga	-294
-293	gccgggggacctgtccaccctcccaggccgacgtcagtgcccgaggactgcctggggacct	-234
-233	gctaggacctgtcacctctgaggacctctgggggtgagaggttcagtcctggaaacacttca	-174
-173	gttctagggggctgggggcagcagcaagttggagttttggggtaccctgcttcacagggc	-114
-113	ccttggaaggagggcaggtgggggtctaaggacaagcagtccttactttgggagtcaacc	-54
-53	ccggcgtgggtggctgctgcaggttgcacactggggccacagaggatccagcaaggATGAAG	6
1		M K 2
7	AAATGGAGCAGCACAGACTTGGGGGCAGCTGCGGACCCACTCCAAAAGGACACCTGCCCA	66
3	K W S S T D L G A A A D P L Q K D T C P	22
67	GACCCCTGGATGGAGACCCTAACTCCAGGCCACCTCCAGCCAAGCCCCAGCTCTCCACG	126
23	D P L D G D P N S R P P P A K P Q L S T	42
127	GCCAAGAGCCGCACCCGGCTCTTTGGGAAGGGTGACTCGGAGGAGGCTTTCCCGGTGGAT	186
43	A K S R T R L F G K G D S E E A F P V D	62
187	TGCCCTCACGAGGAAGGTGAGCTGGACTCCTGCCCGACCATCACAGTCAGCCCTGTTATC	246
63	C P H E E G E L D S C P T I T V S P V I	82
247	ACCATCCAGAGGCCAGGAGACGGCCCCACCGGTGCCAGGCTGCTGTCCCAGGACTCTGTC	306
83	T I Q R P G D G P T G A R L L S Q D S V	102
307	GCCGCCAGCACCGAGAAGACCCTCAGGCTCTATGATCGCAGGAGTATCTTTGAAGCCGTT	366
103	A A S T E K T L R L Y D R R S I F E A V	122
367	GCTCAGAATAACTGCCAGGATCTGGAGAGCCTGCTGCTCTTCCTGCAGAAGAGCAAGAAG	426
123	A Q N N C Q D L E S L L L F L Q K S K K	142
427	CACCTCACAGACAACGAGTTCAAAGACCCTGAGACAGGGAAGACCTGTCTGCTGAAAGCC	486
143	H L T D N E F K D P E T G K T C L L K A	162
487	ATGCTCAACCTGCACGACGGACAGAACACCACCATCCCCCTGCTCCTGGAGATCGCGCGG	546



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163 M L N L H D G Q N T T I P L L L E I A R 182  
547 CAAACGGACAGCCTGAAGGAGCTTGTCAACGCCAGCTACACGGACAGCTACTACAAGGGC 606  
183 Q T D S L K E L V N A S Y T D S Y Y K G 202  
607 CAGACAGCACTGCACATCGCCATCGAGAGACGCAACATGGCCCTGGTGACCCTCCTGGTG 666  
203 Q T A L H I A I E R R N M A L V T L L V 222  
667 GAGAACGGAGCAGACGCTCCAGGCTGCGGCCCATGGGGACTTCTTTAAGAAAACCAAAGGG 726  
223 E N G A D V Q A A A H G D F F K K T K G 242  
727 CGGCCTGGATTCTACTTTCGGTGAAGTGGCCCTGTCCCTGGCCGCGTGCACCAACCAGCTG 786  
243 R P G F Y F G E L P L S L A A C T N Q L 262  
787 GGCATCGTGAAGTTCCTGCTGCAGAACTCCTGGCAGACGGCCGACATCAGCGCCAGGGAC 846  
263 G I V K F L L Q N S W Q T A D I S A R D 282  
847 TCGGTGGGCAACACGGTGCTGCACGCCCTGGTGGAGGTGGCCGACAACACGGCCGACAAC 906  
283 S V G N T V L H A L V E V A D N T A D N 302  
907 ACGAAGTTTGTGACGAGCATGTACAATGAGATTCTGATCCTGGGGGCCAAACTGCACCCG 966  
303 T K F V T S M Y N E I L I L G A K L H P 322  
967 ACGCTGAAGCTGGAGGAGCTCACCAACAAGAAGGGAATGACGCCGCTGGCTCTGGCAGCT 1026  
323 T L K L E E L T N K K G M T P L A L A A 342  
1027 GGGACCGGGAAGATCGGGGTCTTGGCCTATATTCTCCAGCGGGAGATCCAGGAGCCCGAG 1086  
343 G T G K I G V L A Y I L Q R E I Q E P E 362  
1087 TGCAGGCACCTGTCCAGGAAGTTCACCGAGTGGGCCTACGGGCCCGTGCACCTCCTCGCTG 1146  
363 C R H L S R K F T E W A Y G P V H S S L 382  
1147 TACGACCTGTCCTGCATCGACACCTGCCAGAAGAACTCGGTGCTGGAGGTGATCGCCTAC 1206  
383 Y D L S C I D T C E K N S V L E V I A Y 402  
1207 AGCAGCAGCGAGACCCCTAATCGCCACGACATGCTCTTGGTGGAGCCGCTGAACCGACTC 1266  
403 S S S E T P N R H D M L L V E P L N R L 422  
1267 CTGCAGGACAAGTGGGACAGATTTCGTCAAGCGCATCTTCTACTTCAACTTCCTGGTCTAC 1326  
423 L Q D K W D R F V K R I F Y F N F L V Y 442  
1327 TGCCTGTACATGATCATCTTCACCATGGCTGCCTACTACAGGCCCGTGGATGGCTTGCCT 1386  
443 C L Y M I I F T M A A Y Y R P V D G L P 462  
1387 CCCTTTAAGATGGAAAAAATTGGAGACTATTTCCGAGTTACTGGAGAGATCCTGTCTGTG 1446  
463 P F K M E K I G D Y F R V T G E I L S V 482  
1447 TTAGGAGGAGTCTACTTCTTTTTCCGAGGGATTTCAGTATTTCTGCAGAGGCGGCCGTCG 1506  
483 L G G V Y F F F R G I Q Y F L Q R R P S 502  
1507 ATGAAGACCCTGTTTGTGGACAGCTACAGTGAGATGCTTTTCTTCTGCAGTCACTGTTC 1566  
503 M K T L F V D S Y S E M L F F L Q S L F 522  
1567 ATGCTGGCCACCGTGGTGTGTAATTCAGCCACCTCAAGGAGTATGTGGCTTCCATGGTA 1626  
523 M L A T V V L Y F S H L K E Y V A S M V 542  
1627 TTCTCCCTGGCCTTGGGCTGGACCAACATGCTCTACTACACCCGCGGTTTCCAGCAGATG 1686

FIG. 3<sub>CONT'D</sub>

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543 F S L A L G W T N M L Y Y T R G F Q Q M 562

1687 GGCATCTATGCCGTCATGATAGAGAAGATGATCCTGAGAGACCTGTGCCGTTTCATGTTT 1746  
563 G I Y A V M I E K M I L R D L C R F M F 582

1747 GTCTACATCGTCTTCTTGTTCGGGTTTTCCACAGCGGTGGTGACGCTGATTGAAGACGGG 1806  
583 V Y I V F L F G F S T A V V T L I E D G 602

1807 AAGAATGACTCCCTGCCGTCTGAGTCCACGTCGCACAGGTGGCGGGGGCCTGCCTGCAGG 1866  
603 K N D S L P S E S T S H R W R G P A C R 622

1867 CCCCCGATAGCTCCTACAACAGCCTGTACTCCACCTGCCTGGAGCTGTTCAAGTTCACC 1926  
623 P P D S S Y N S L Y S T C L E L F K F T 642

1927 ATCGGCATGGGCGACCTGGAGTTCACTGAGAAGTATGACTTCAAGGCTGTCTTCATCATC 1986  
643 I G M G D L E F T E N Y D F K A V F I I 662

1987 CTGCTGCTGGCCTATGTAATTCTCACCTACATCCTCCTGCTCAACATGCTCATCGCCCTC 2046  
663 L L L A Y V I L T Y I L L L N M L I A L 682

2047 ATGGGTGAGACTGTCAACAAGATCGCACAGGAGAGCAAGAACATCTGGAAGCTGCAGAGA 2106  
683 M G E T V N K I A Q E S K N I W K L Q R 702

2107 GCCATCACCATCCTGGACACGGAGAAGAGCTTCCTTAAGTGCATGAGGAAGGCCTTCCGC 2166  
703 A I T I L D T E K S F L K C M R K A F R 722

2167 TCAGGCAAGCTGCTGCAGGTGGGGTACACACCTGATGGCAAGGACGACTACCGGTGGTGC 2226  
723 S G K L L Q V G Y T P D G K D D Y R W C 742

2227 TTCAGGGTGGACGAGGTGAACTGGACCACCTGGAACACCAACGTGGGCATCATCAACGAA 2286  
743 F R V D E V N W T T W N T N V G I I N E 762

2287 GACCCGGGCAACTGTGAGGGCGTCAAGCGCACCTGAGCTTCTCCCTGCGGTCAAGCAGA 2346  
763 D P G N C E G V K R T L S F S L R S S R 782

2347 GTTTCAGGCAGACACTGGAAGAACTTTGCCCTGGTCCCCCTTTTAAGAGAGGCAAGTGCT 2406  
783 V S G R H W K N F A L V P L L R E A S A 802

2407 CGAGATAGGCAGTCTGCTCAGCCCCGAGGAAGTTTATCTGCGACAGTTTTCAGGGTCTCTG 2466  
803 R D R Q S A Q P E E V Y L R Q F S G S L 822

2467 AAGCCAGAGGACGCTGAGGTCTTCAAGAGTCTGCCGCTTCCGGGAGAGtgaggacgt 2526  
823 K P E D A E V F K S P A A S G E K 839

2527 cacgcagacagcactgtcaacactgggccttaggagaccccggttgccaoggggggctgct 2586

2587 gaggggaacaccagtgtctgtgcagcagcctggcctggtctgtgcctgcccagcatgttcc 2646

2647 caaatctgtgctggacaagctgtgggaagcgttcttggaagcatggggagtgtgtacat 2706

2707 ccaacogtcaactgtccccaagtgaatctcctaacagactttcaggtttttactcacttta 2766

2767 ctaaacagtttggttggtcagtcctcactgggacatgttagggcccttggttttctttgatt 2826

2827 ttattcttttctgtgagacagagttcactcttgttgcccaggctggagtgcagtgggtgtg 2886

2887 atcttggtcactgcaacctctgctcccggttcaagcgattctctgtcttcagtcctccc 2946

FIG. 3CONT'D

2947 aagtagcttggattacaggtgagcactaccacgcccggctaatttttgatattttaatag 3006  
3007 agacgggggtttcaccaatggtggccaggctggtctcgaactcttgacctcaggtgatctgc 3066  
3067 ccgccttggcctcccaaagtgcctgggattacaggtgtgagccgctgcgctcggccttctt 3126  
3127 tgattttatattattagggagcaaaagtaaatgaagcccaggaaaaacacctttgggaacaa 3186  
3187 actcttcctttgatggaaaatgcagaggcccttcctctctgtgccgtgcttgctcctctt 3246  
3247 acctgcccgggtggtttgggggtgttggtgtttcctccctggagaagatgggggaggctg 3306  
3307 tcccaactcccagctctggcagaatcaagctggtgcagcagtgccctcttcacaccttcctt 3366  
3367 acgatcaatcacagtctccagaagatcagctcaattgctgtgcaggttaaaactacagaa 3426  
3427 ccacatcccaaaggtacctggtgaagaatgtttgaaagatcttccatttctaggaacccca 3486  
3487 gtctgtcttctccgcaatggcacatgcttccactccatccatactggcatcctcaaataa 3546  
3547 acagatatgtatacaaaaaaaaaaaaaaaaaaaaaaaaaaaaaa 3591

FIG. 3<sub>CONT'D</sub>

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## FIG. 4


## AMINO ACID SEQUENCE OF hVR1

1 MKKWSSTDLG AAADPLQKDT CPDPLDGDPN SRPPPAKPQL STAKSRTLRF  
 51 GKGDSSEAFP VDCPHEEGEL DSCPTITVSP VITIQRPGDG PTGARLLSQD  
 101 SVAASTEKTL RLYDRRSIFE AVAQNNCQDL ESLLLFLQKS KKHLTDNEFK  
 151 DPETGKTCLL KAMLNLDGQ NTTIPLLEI ARQTDSELKEL VNASYTDSYY  
 201 KGQ TALHIAI ERRNMALVTL LVENGADVQA AAHGDFFKKT KGRPGFYFGE  
 251 LPLSLAACTN QLGIVKFLQ NSWQTADISA RDSVGN TVLH ALVEVADNTA  
 301 DNTKFVTSMY NEILILGAKL HPTLKEELT NKKGMTPLAL AAGTGKIGVL  
 351 AYILQREIQE PECHLSRKF TEWAYGPVHS SLYDLSCIDT CEKNSVLEVI  
 401 AYSSSETPNR HDMLLVEPLN RLLQDKWDRF VKR IETENELVYCIYMIEEL  
 451 MAAYLRPVDG LPPFKMEKIG DYFRVTGEI LSVLGGVYFFESRGIQY FLQRR  
 501 PSMKTLFVLSYSEMIFFELQSLEMLATVVLYESILKEYVAS MVESEFAGWT  
 551 NMDFYLRGEQOMGIEYAVMIE KMILRIICREMEVYIVFEEGTFSTAVVTLIE  
 601 DGKNDSLPSE STSHRWGPA CRPPDSSYNS LYSTCLELFK FTIGMGDLEF  
 651 TENYDEKAVEELIETIYAVIEIYNIIFGANMIEALMGETVNKI AQESKNIWKL  
 701 QRAITILDTE KSFLKCMRKA FRSGKLLQVG YTPDGKDDYR WCFRVDEVNWN  
 751 TTWNTNVGII NEDPGNCXGV KRTLSFSLRS SRVSGRHWKN FALVPLLREA  
 801 SARDRQSAQP EEVYLRQFSG SLKPDAEVF KSPAASGEK\*

## Key

T/S predicted phosphorylation sites

 Transmembrane domains

 Ankyrin binding domains

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## FIG. 5

COMPARISON OF THE AMINO ACID SEQUENCE OF THE RAT (VR1)  
AND HUMAN (hVR1) VANILLOID PROTEINS.

	10	20	30	40	50
VR1	MEQRASLDSESESP	QENSCLDPPDRD	PNCKPPV	KPHIFTTRS	RTLRF
hVR1	MKKWSSTD	LGAADPLQK	DTCPDPLD	GPNSRPP	PAKPQLSTAKSRTLRF
	60	70	80	90	100
VR1	GKGDSE	EASPLDCPYEE	GGLASCP	IIITVSSV	LTIQRPDGPASVRPSSQD
hVR1	GKGDSE	EAFVDCPHEE	GELDSCPT	ITITVSPV	ITIQRPDGP
	110	120	130	140	150
VR1	SVSAG	EKPPRLYDRRS	IFDAVAQ	SNCQELES	LLPFLQ
hVR1	SVAAS	TEKTLRLYDRRS	IFEAVAQ	NNCQDLES	LLLFLQ
	160	170	180	190	200
VR1	DPETGKT	CLLKAM	LNHN	GNDTIAL	LLDVARKTDSL
hVR1	DPETGKT	CLLKAM	LNH	DGNTTIP	LLLEIARQ
	210	220	230	240	250
VR1	KGQTAL	HIAIERN	MTLV	TLLVENG	ADVQAAANGDFF
hVR1	KGQTAL	HIAIERN	MTLV	TLLVENG	ADVQAAAHGDFF
	260	270	280	290	300
VR1	LPLSLA	ACTNQLA	IVKELLQ	NSWQPA	DISARD
hVR1	LPLSLA	ACTNQLG	IVKELLQ	NSWQTAD	ISARD
	310	320	330	340	350
VR1	DNTKFV	TSMYNEI	LILGAK	LHPTLK	LEEITNR
hVR1	DNTKFV	TSMYNEI	LILGAK	LHPTLK	LEELTNK
	360	370	380	390	400
VR1	AYILQRE	IHEPECR	HLSRK	FTWAYG	VPVHSSLY
hVR1	AYILQRE	IQEPECR	HLSRK	FTWAYG	VPVHSSLY
	410	420	430	440	450
VR1	AYSSSE	TPNRHDM	LLVEPL	NRL	LQDKWDRFV
hVR1	AYSSSE	TPNRHDM	LLVEPL	NRL	LQDKWDRFV
	460	470	480	490	500
VR1	AAAYYR	PVEGLPPY	KLKWT	VG	DYFRVTGEILSV
hVR1	MAAYYR	PVDGLPP	FKMEK	IGDYFRVTGEILSV	LG
	510	520	530	540	550
VR1	RPSLKS	LFVDSYSE	ILFEVQ	SLEF	MLVSVVLYFS
hVR1	RPSMKT	LFVDSYSE	MLF	FLQSLF	MLATVVLYF
	560	570	580	590	600
VR1	TNMLYY	TRGFQOM	GIYAV	MI	EKMILRDL
hVR1	TNMLYY	TRGFQOM	GIYAV	MI	EKMILRDL
	610	620	630	640	650
VR1	EDGKNS	SLPMEST	PHKCRGS	ACK	PGNSYNSLY
hVR1	EDGKNS	SLPSEST	SHRWRGP	ACRPPD	SSYNSLY
	660	670	680	690	700
VR1	FTENYD	EKAVFI	ILLAYV	ILTYI	ILLNMLIAL
hVR1	FTENYD	EKAVFI	ILLAYV	ILTYI	ILLNMLIAL
	710	720	730	740	750
VR1	LQRAIT	ILDTEKS	FLKCMR	KAF	RS
hVR1	LQRAIT	ILDTEKS	FLKCMR	KAF	RS
	760	770	780	790	800
VR1	WTTWNT	TVNGI	NEDPG	NCEGV	KRTLSF
hVR1	WTTWNT	TVNGI	NEDPG	NCEGV	KRTLSF
	810	820	830		
VR1	ASTDRH	ATQOE	EVOLK	HYTG	SLKPEDAEV
hVR1	ASARDR	QSAQPEE	VYLRQ	FSGSLK	PEDAEVFKSPAASGEK

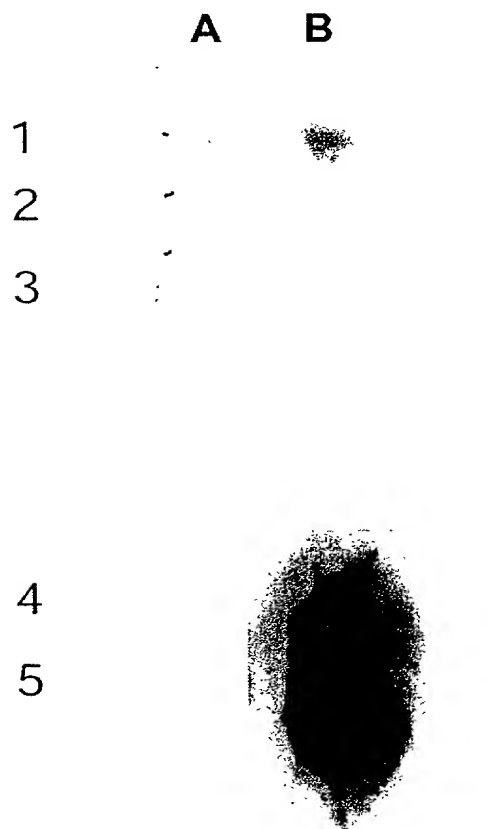
FULL-LENGTH hRV1 CLONED INTO (A) pBLUESCRIPT SK(+) (hVR1pBSK) AND (B) pCIN5-NEW (hVR1pCIN5) VIA NotI/EcoRI RESTRICTION SITES.



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## FIG. 7

SLOT HYBRIDISATION WITH hVR1 PROBE



Well

1A hDRG

2A rDRG

3A Water

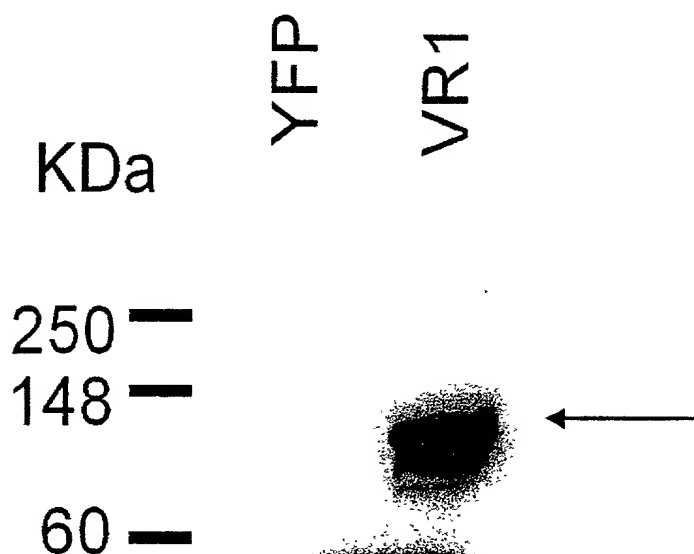
4B EST3 clone

5B 260bp Amplicon from Brain cDNA

1B hDRG

## FIG. 8

WESTERN BLOT PROBED WITH ANTI-hVR1 ANTIBODIES.  
ARROW POINTS TO hVR1 SPECIFIC BAND





**FIG. 9**

IN SITU LOCALISATION OF VR1 IN RAT DRG TISSUE SECTIONS.  
ARROW POINTS TO A VR1 EXPRESSING SMALL DIAMETER  
( $<25\mu\text{m}$ ) NEURONE CELL BODY, MAGNIFICATION USED 147x10.

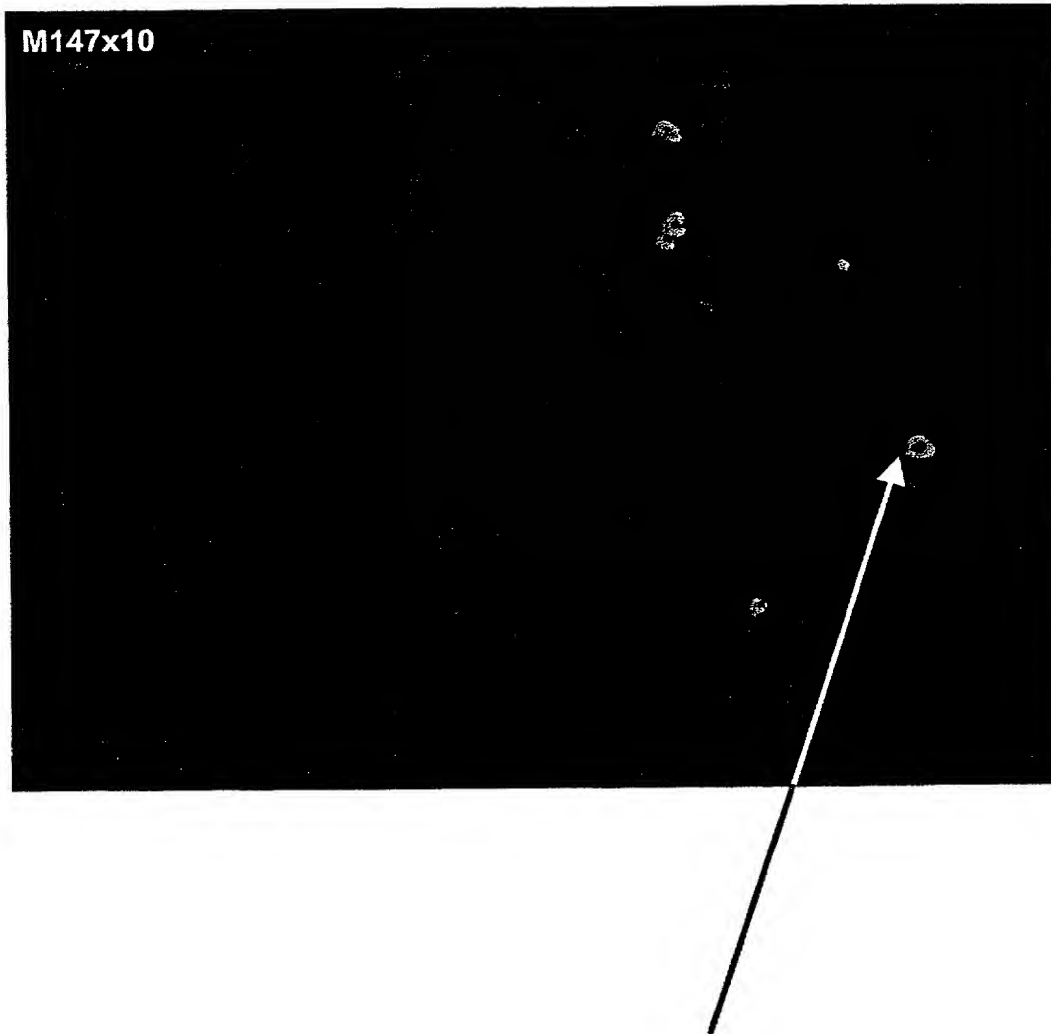


FIG. 10A

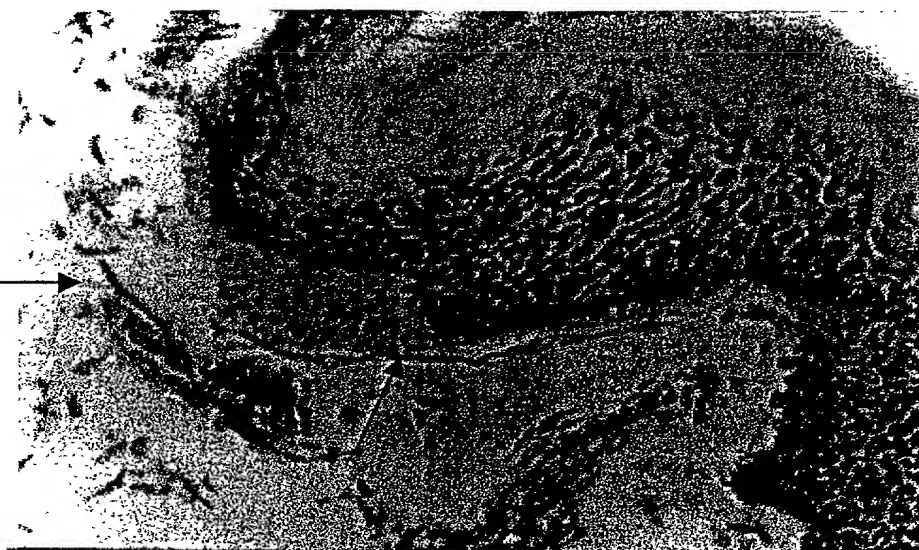
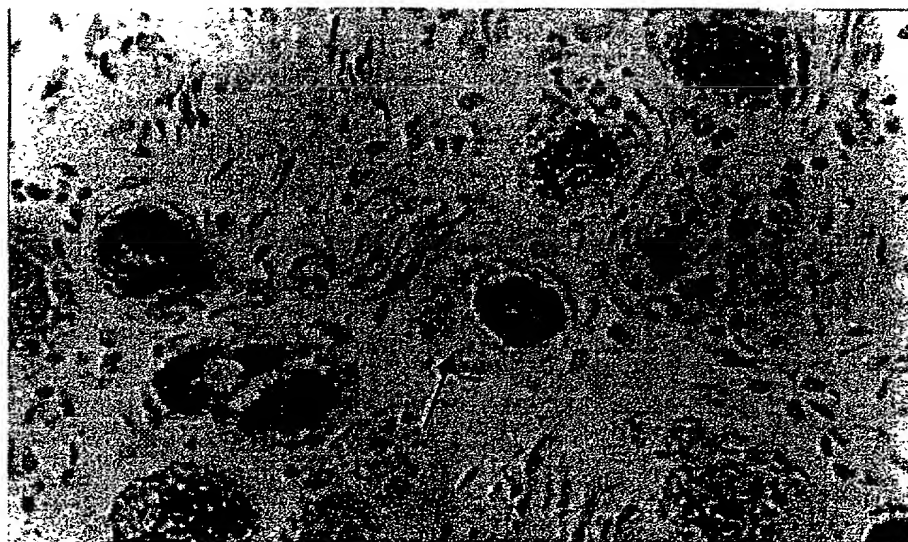


FIG. 10B

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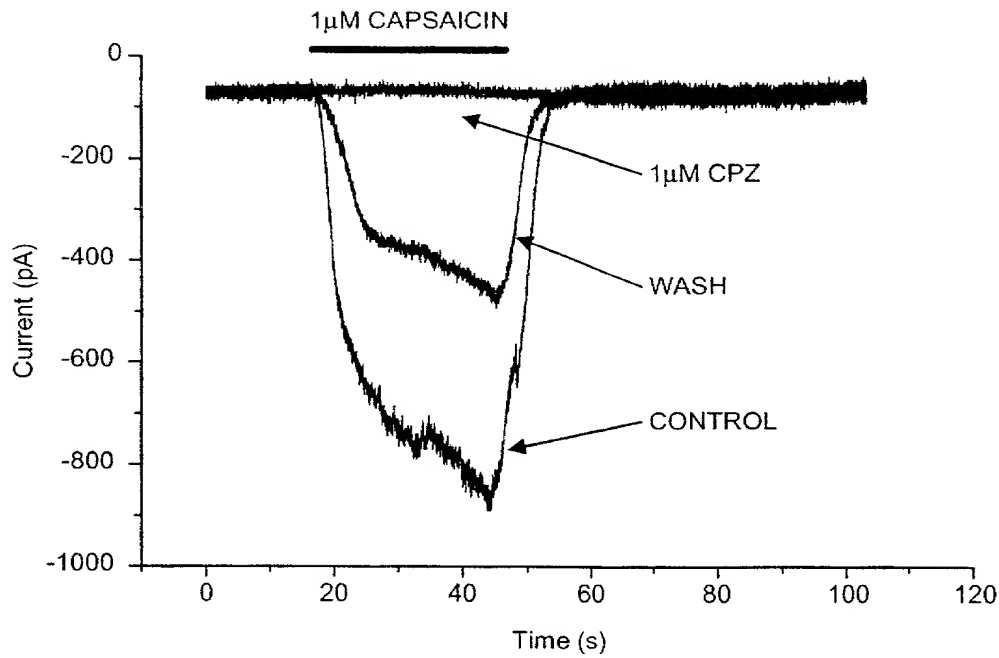


FIG. 11A

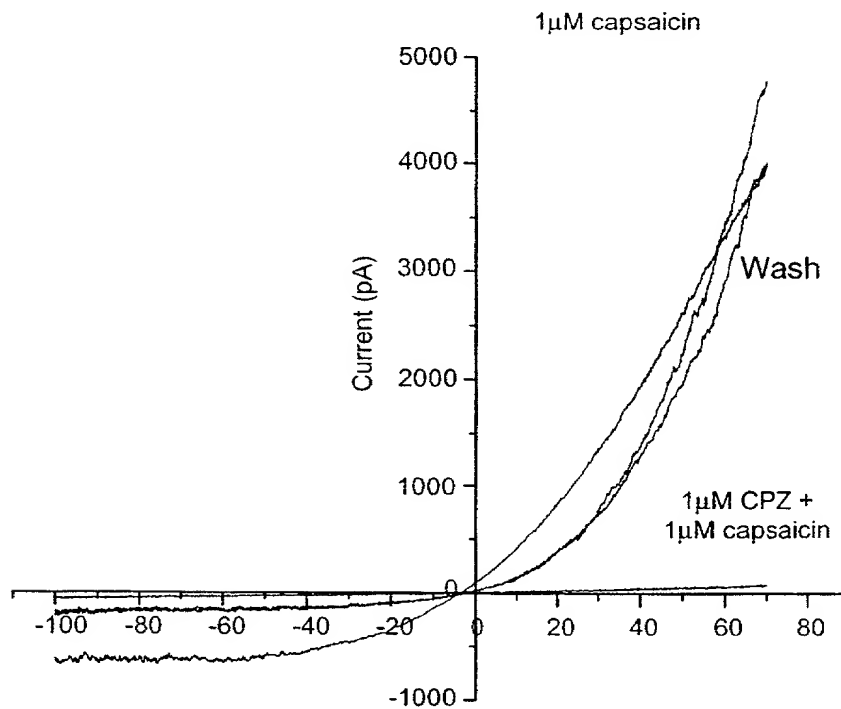


FIG. 11B

SOLUTIONS  
OUTSIDE 140mM Na<sup>+</sup> 2mM Ca<sup>2+</sup>  
INSIDE 140mM Cs<sup>+</sup>

FIG. 12A

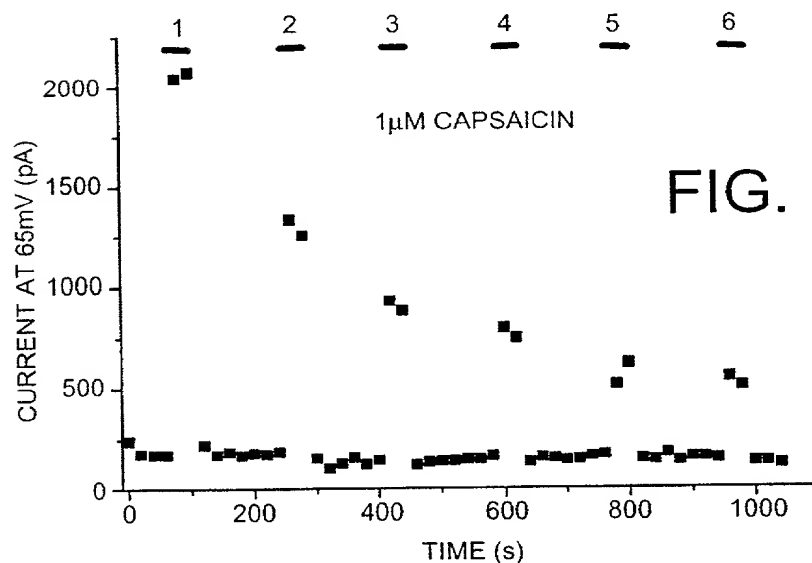
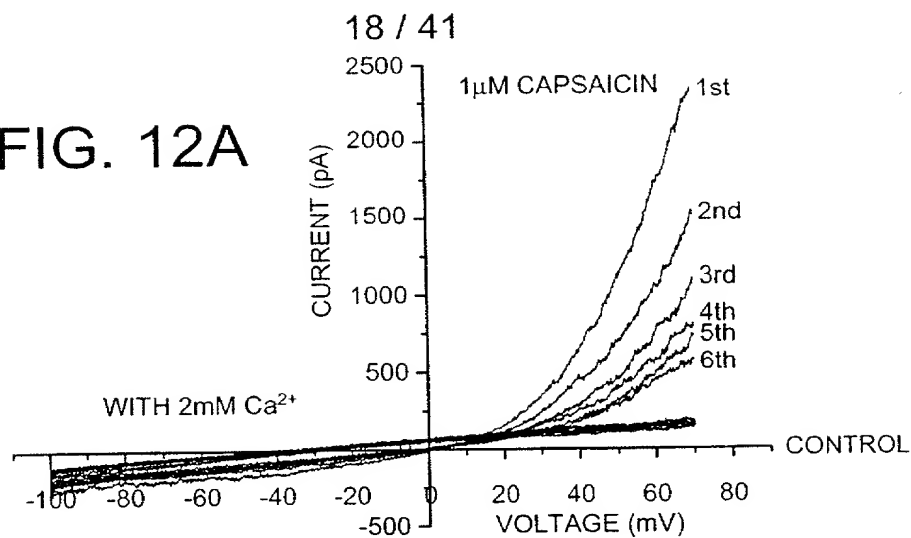


FIG. 12B

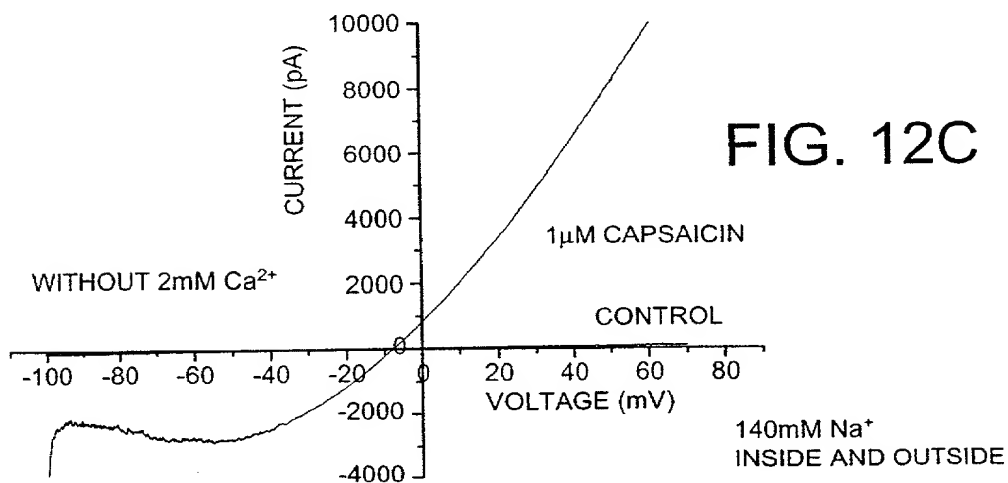
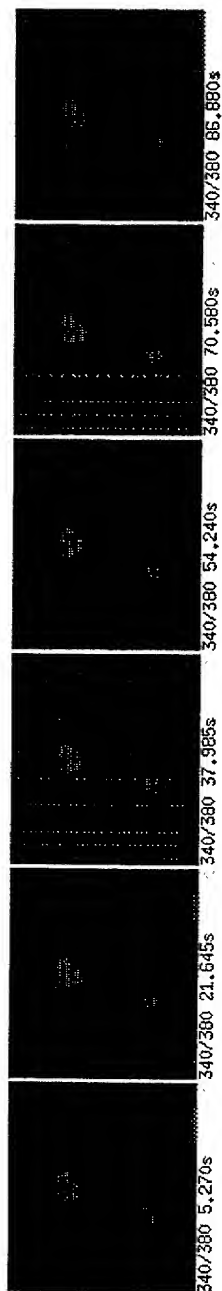
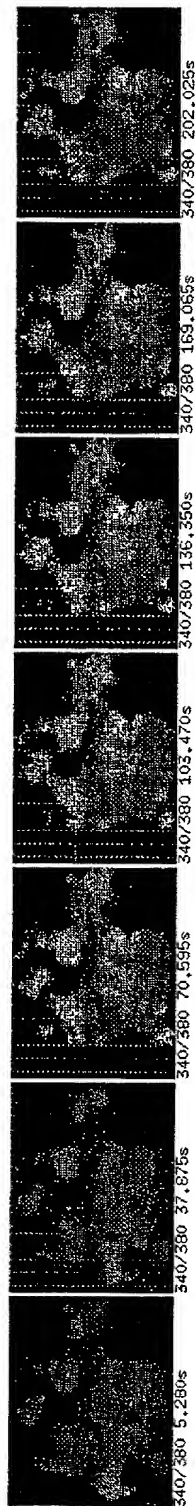


FIG. 12C

13A pCIN5-new in HEK293T, 24hr transient expression, stimulated with 3 $\mu$ M capsaicin at time point 52 secs of time course



13B hVR1pCIN5 in HEK293T, 24hr expression, stimulated with 1 $\mu$ M capsaicin at time point 52 seconds



13C hVR1pCIN5 in HEK293T, 24hr transient expression, 20 min pre-incubation with 10  $\mu$ M capsazepine, stimulated with 1  $\mu$ M capsaicin at time point 52 seconds of time course

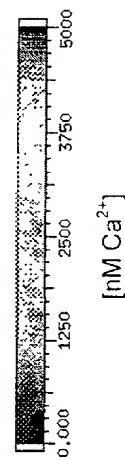
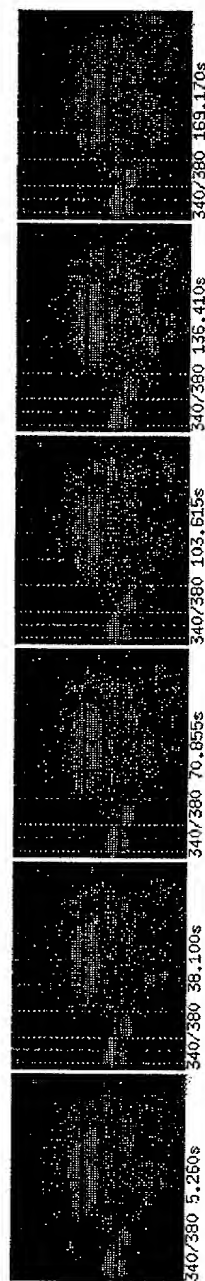
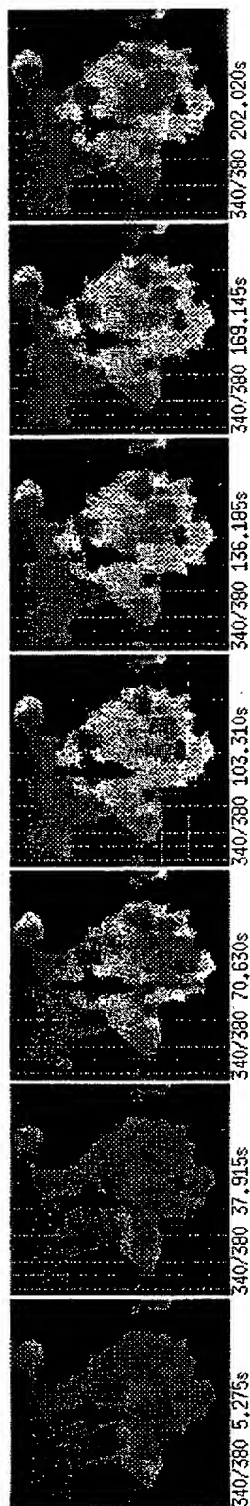


FIG. 13

13D hVR1pCIN5 in HEK293T, 24hr transient expression, stimulated with 10uM anandamide at time point 52 seconds



13E hVR1pCIN5 in HEK293T, 24hr transient expression, 20 min pre-incubation in 10uM capsaizpine, stimulated with 10uM anandamide at time point 52 sec

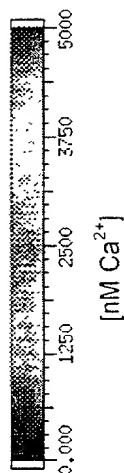
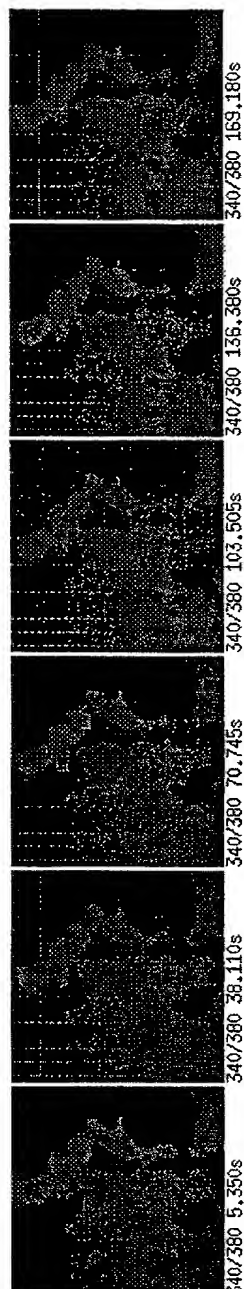
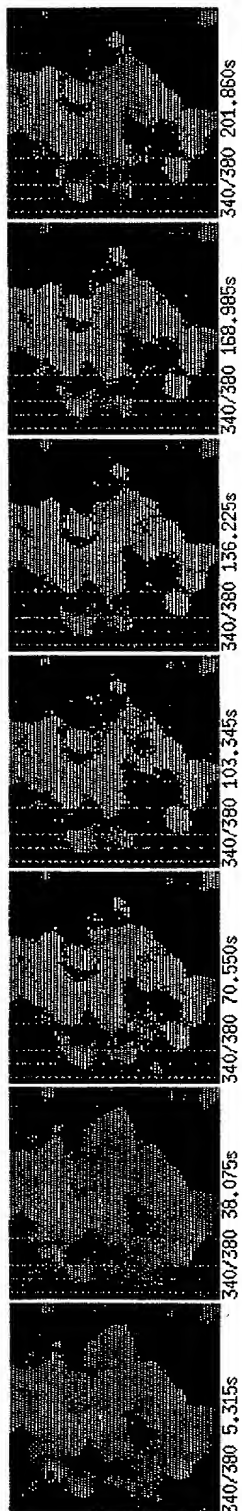


FIG. 13CONT'D

13F hVR1pCIN5 in HEK293T cells, 24hr transient expression, stimulated with 1uM Resiniferatoxin at time point 52 seconds



13G hVR1pCIN5 in HEK293T, 24hr transient expression, 20 min pre-incubation with 10 uM capsaizepine, stimulated with 1 uM Resiniferatoxin at time point 52 seconds

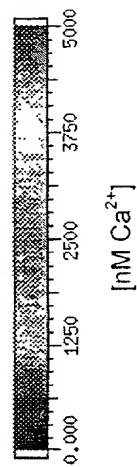
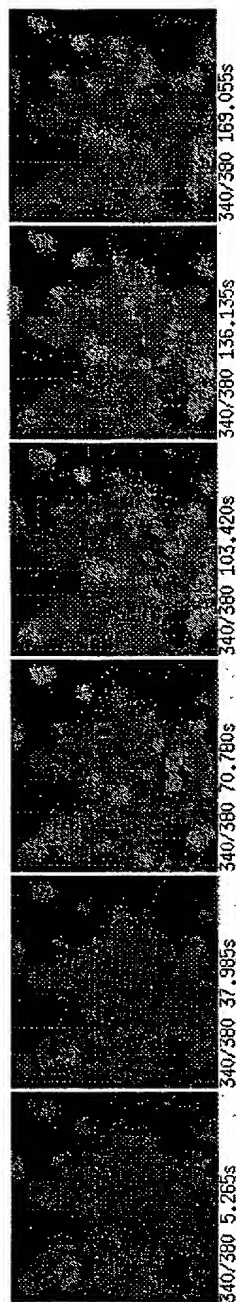


FIG. 13<sub>CONT'D</sub>

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## FIG. 14

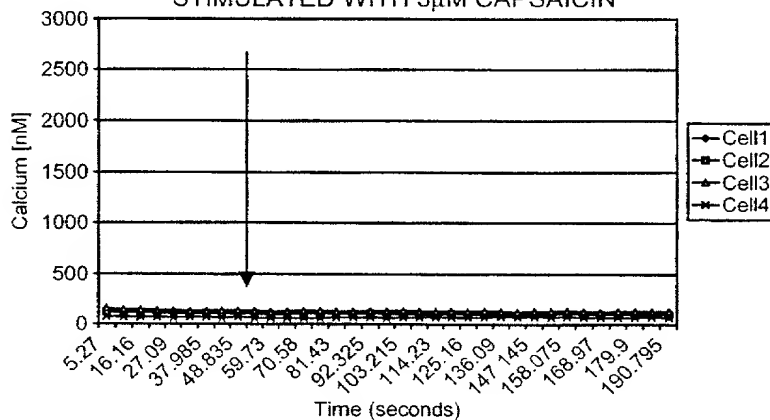
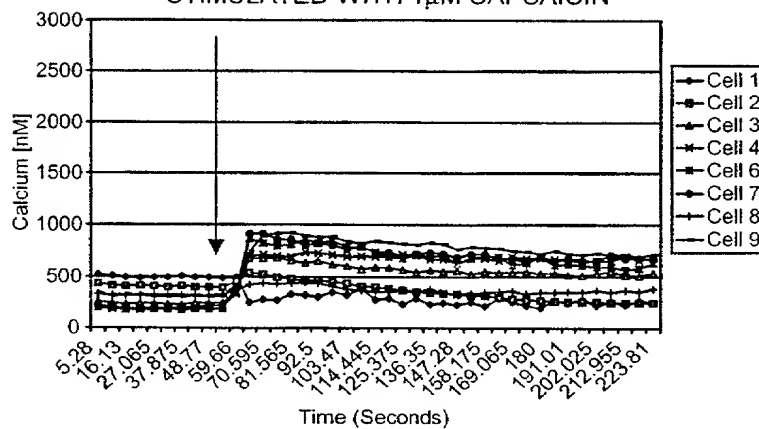
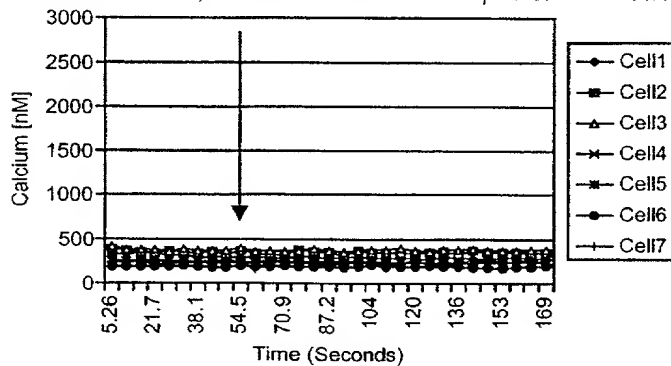
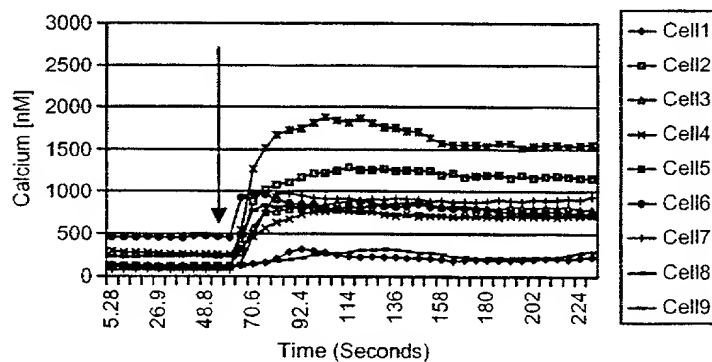
EXPOSURE OF TRANSFECTED CELLS TO AGONISTS  
(ADDITION INDICATED BY ARROW).14A: pCIN5-NEW IN HEK293T, 24hr TRANSIENT EXPRESSION,  
STIMULATED WITH  $3\mu\text{M}$  CAPSAICIN14B: hVR1pCIN5 IN HEK293T, 24hr EXPRESSION,  
STIMULATED WITH  $1\mu\text{M}$  CAPSAICIN14C: hVR1pCIN5 IN HEK293T, 24hr TRANSIENT  
EXPRESSION, 20 MIN PRE-INCUBATION WITH  $10\mu\text{M}$   
CAPSAZEPINE, STIMULATION WITH  $1\mu\text{M}$  CAPSAICIN



FIG. 14<sub>CONT'D</sub>

14D: hVR1pCIN5 IN HEK293T, 24hr TRANSIENT  
EXPRESSION, STIMULATION WITH 10 $\mu$ M ANANDAMIDE



14E: hVR1pCIN5 IN HEK293T, 24hr TRANSIENT  
EXPRESSION, 20 MIN PRE-INCUBATION IN 10 $\mu$ M  
CAPAZEPINE, STIMULATED WITH 10 $\mu$ M ANANDAMIDE

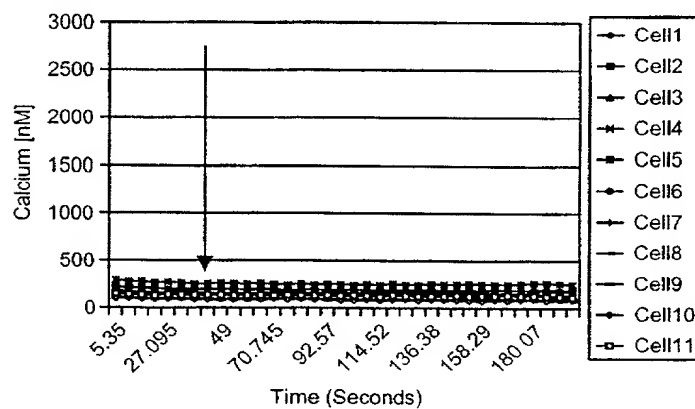
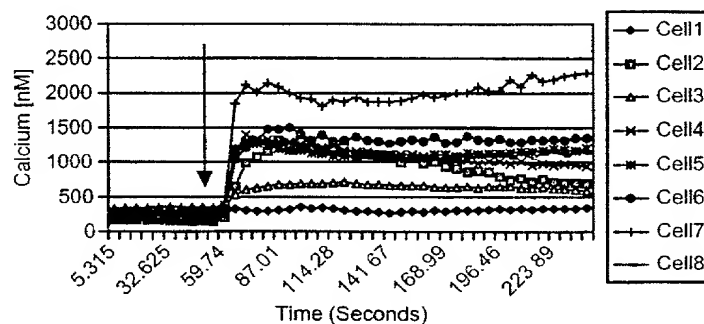
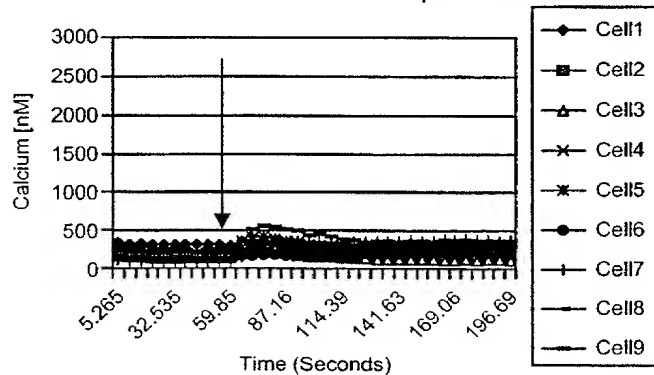


FIG. 14<sub>CONT'D</sub>

14F: hVR1pCIN5 IN HEK293T CELLS, 24hr TRANSIENT  
EXPRESSION, STIMULATED WITH 1 $\mu$ M RESINIFERATOXIN



14G: hVR1pCIN5 IN HEK293T, 24hr TRANSIENT  
EXPRESSION, 20 MIN PRE-INCUBATION WITH 10 $\mu$ M  
CAPSAZEPINE, STIMULATED WITH 1 $\mu$ M RESINIFERATOXIN



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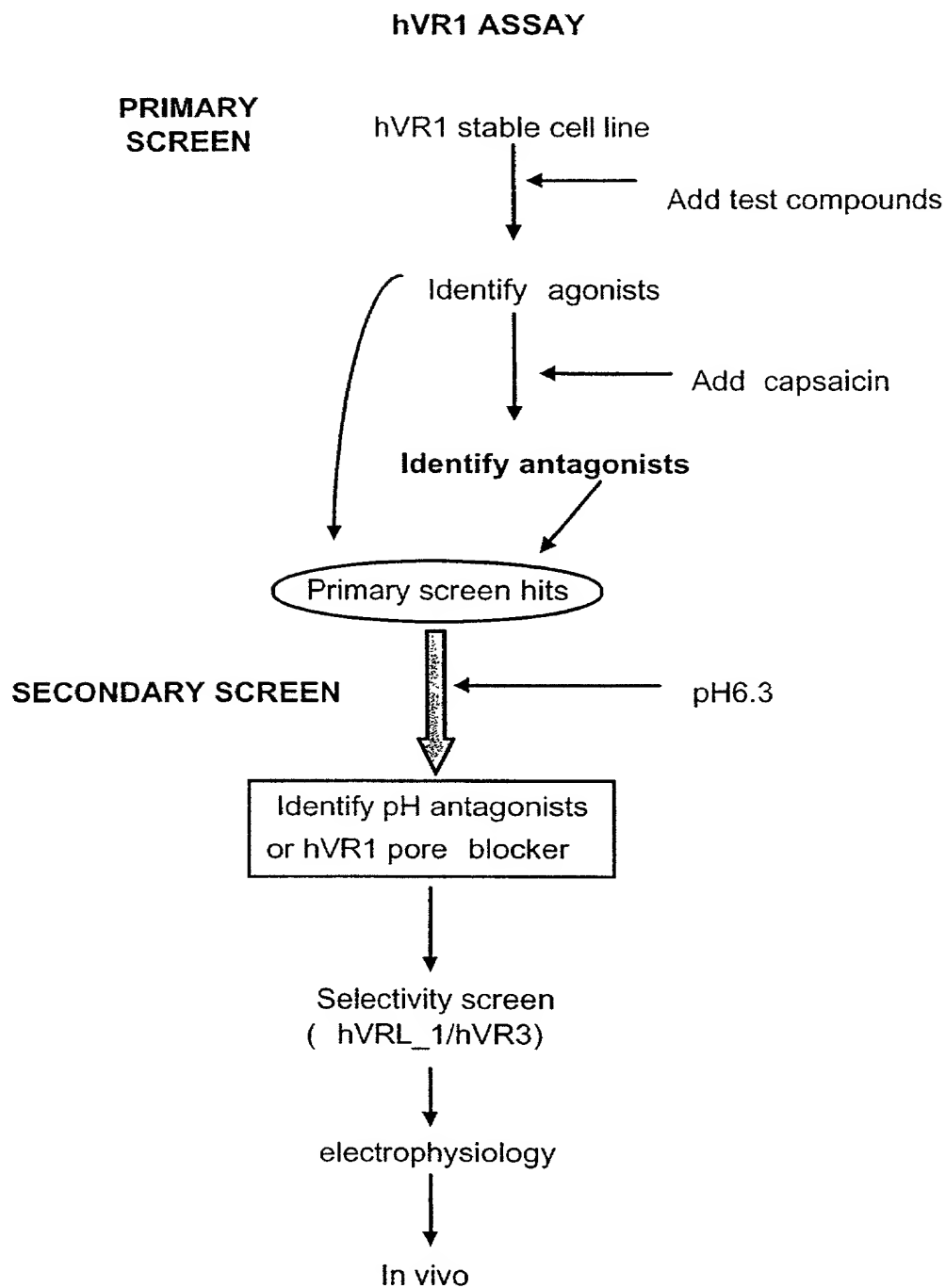


FIG. 15

FIG. 16



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## FIG. 17

hVR3 SEQUENCE INCLUDING 5' UTR (nt -686 TO nt 0) CODING  
REGION (nt1 TO nt 2889), 3'UTR (nt 2890 TO nt 3418)

-684	ttacgcgtaagaaatacccaagcttatgcatcaagcttggtacccagctcggatccact	-625
-624	agtaccgcccggccagtgtgctggaattcaaggtgaggagaggagcatggatcctgggagc	-565
-564	gagtgtgtgcaggccaggaggggctttccagaggagcccagttgagctggaacaccagt	-505
-504	gggaggagttgaccagcaaaggtgcaggaggaggatcagcaactttgcactggggagcagag	-445
-444	tttgtgcactggggaagtcaactcaagtattggagcctcagtttctgttctgtaaaatg	-385
-384	ggttcatcatgacagtgtttgatgaggaaaaggactgccggcctacacagcaagtcacaca	-325
-324	tggattttctgagccctcctgtgcctgaagcccacgggttaatggttctgccttagcagg	-265
-264	tgcttaccacgtgcaggcactgcactgcactggccactggactgcatgttctgtccatg	-205
-204	aggcttggatatcccatcttacagatcaggaagctgaggctatgaaatgtcgacttgct	-145
-144	caatgtcatggaatgactaagtgtggagcctggatttgaacttggtctctctggggctcca	-85
-84	aagctggctttcttggtcagcagtagggtctgggatccaagtatgggggtcccagcttgac	-25
-24	cctgaagtccaccctctttcagctaATGCCCAGGCTAGTTGGAACCTGGGGCCAATTTGTG	35
36	TTTCCAGGTTTCGTGAAAGAGGCTCCTGTTGCAGTTCCCGCCTGAGGCTGGCGGCCAACCA	95
96	CATCTGGGAGTGGCCTCCCTGTGCCCCCTGTCATTACAACGGTGGCTTTGAAGCAGCTGGC	155
156	AGCACTGCTGCTTGTCCACGTGGGAGGGGGCTTCTGGAGCCCCCGCCCCCTGGCCGGGTT	215
216	CTGCCCTGACTCCCCTTTCATTCCCCTTGCAGGCTGAGCAGTGCAGACGGGCCTGGGGCAGG	275
276	CATGGCGGATTCCAGCGAAGGCCCCCGCGCGGGGCCCGGGGAGGTGGCTGAGCTCCCCGG	335
336	GGATGAGAGTGGCACCCCAGGTGGGGAGGCTTTTCTCTCTCTCTCCCTGGCCAATCTGTT	395

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396 TGAGGGGGAGGATGGCTCCCTTTTCGCCCTCACCGGCTGATGCCAGTCGCCCTGCTGGCCC 455

456 AGGCGATGGGCGACCAAATCTGCGCATGAAGTTCCAGGGCGCCTTCCGCAAGGGGGTGCC 515

516 CAACCCCATCGATCTGCTGGAGTCCACCCTATATGAGTCCTCGGTGGTGCCTGGGCCCCAA 575

576 GAAAGCACCCATGGACTCACTGTTTGACTACGGCACCTATCGTCACCACTCCAGTGACAA 635

636 CAAGAGGTGGAGGAAGAAGATCATAGAGAAGCAGCCGCAGAGCCCCAAAGCCCCTGCCCC 695

696 TCAGCCGCCCCCATCCTCAAAGTCTTCAACCGGCCTATCCTCTTTGACATCGTGTCCCG 755

756 GGGCTCCACTGCTGACCTGGACGGGCTGCTCCCATTCTTGCTGACCCACAAGAAACGCCT 815

816 AACTGATGAGGAGTTTCGAGAGCCATCTACGGGAAGACCTGCCTGCCCAAGGCCCTTGCT 875

876 GAACCTGAGCAATGGCCGCAACGACACCATCCCTGTGCTGCTGGACATCGCGGAGCGCAC 935

936 CGGCAACATGCGGGAGTTCATTAACTCGCCCTTCCGTGACATCTACTATCGAGGTCAGAC 995

996 AGCCCTGCACATCGCCATTGAGCGTCGCTGCAAACACTACGTGGAACCTTCTCGTGCCCCA 1055

1056 GGGAGCTGATGTCCACGCCCAGGCCCGTGGGCGCTTCTTCCAGCCCAAGGATGAGGGGGG 1115

1116 CTACTTCTACTTTGGGGAGCTGCCCCTGTCGCTGGCTGCCTGCACCAACCAGCCCCACAT 1175

1176 TGTCAACTACCTGACGGAGAACCCCCACAAGAAGGCGGACATGCGGCGCCAGGACTCGCG 1235

1236 AGGCAACACAGTGCTGCATGCGCTGGTGGCCATTGCTGACAACACCCGTGAGAACACCAA 1295

1296 GTTTGTTACCAAGATGTACGACCTGCTGCTGCTCAAGTGTCGCCGCTCTTCCCCGACAG 1355

1356 CAACCTGGAGGCCGTGCTCAACAACGACGGCCTCTCGCCCCCTCATGATGGCTGCCAAGAC 1415

1416 GGGCAAGATTGGGATCTTTCAGCACATCATCCGGCGGGAGGTGACGGATGAGGACACACG 1475

1476 GCACCTGTCCCGCAAGTCCAAGGACTGGGCCTATGGGCCAGTGTATTCTCGCTTTATGA 1535

FIG. 17<sub>CONT'D</sub>

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1536 CCTCTCCTCCCTGGACACGTGTGGGGAAGAGGCCTCCGTGCTGGAGATCCTGGTGTACAA 1595  
1596 CAGCAAGATTGAGAACCGCCACGAGATGCTGGCTGTGGAGCCCATCAATGAACTGCTGCG 1655  
1656 GGACAAGTGGCGGAAGTTCGGGGCCGTCTCCTTCTACATCAACGTGGTCTCCTACCTGTG 1715  
1716 TGCCATGGTTATCTTCACTCTCACCGCCTACTACCAGCCGCTGGAGGGCACACCGCCGTA 1775  
1776 CCCTTACCGCACCACGGTGGACTACCTGCGGCTGGCTGGCGAGGTCATTACGCTCTTCAC 1835  
1836 TGGGGTCTGTCTTCTTCACTCAACATCAAAGACTTGTTTCATGAAGAAATGCCCTGGAGT 1895  
1896 GAATTCTCTCTTCATTGATGGCTCCTTCCAGCTGCTCTACTTCATCTACTCTGTCTGGT 1955  
1956 GATCGTCTCAGCAGCCCTCTACCTGGCAGGGATCGAGGCCTACCTGGCCATGATGGTCTT 2015  
2016 TGCCCTGGTCTGGGCTGGATGAATGCCCTTTACTTCACCCGTGGGCTGAAGCTGACGGG 2075  
2076 GACCTATAGCATCATGATCCAGAAGATTCTCTTCAAGGACCTTTTCCGATTCTGCTCGT 2135  
2136 CTACTTGCTCTTCATGATCGGCTACGCTTCAGCCCTGGTCTCCCTCCTGAACCCGTGTGC 2195  
2196 CAACATGAAGGTGTGCAATGAGGACCAGACCAACTGCACAGTGCCCACTTACCCCTCGTG 2255  
2256 CCGTGACAGCGAGACCTTCAGCACCTTCCTCCTGGACCTGTTTAAGCTGACCATCGGCAT 2315  
2316 GGGCGACCTGGAGATGCTGAGCAGCACCAAGTACCCCGTGGTCTTCATCATCCTGCTGGT 2375  
2376 GACCTACATCATCCTCACCTCTGTGCTGCTCCTCAACATGCTCATTGCCCTCATGGGCGA 2435  
2436 GACAGTGGGCCAGGTCTCCAAGGAGAGCAAGCACATCTGGAAGCTGCAGTGGGCCACCAC 2495  
2496 CATCCTGGACATTGAGCGCTCCTTCCCCGTATTCTGAGGAAGGCCTTCCGCTCTGGGGA 2555  
2556 GATGGTCACCGTGGGCAAGAGCTCGGACGGCACTCCTGACCGCAGGTGGTGCTTCAGGGT 2615  
2616 GGATGAGGTGAACTGGTCTCACTGGAACCAGAACTTGGGCATCATCAACGAGGACCCGGG 2675

FIG. 17 CONT'D

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2676 CAAGAATGAGACCTACCAGTATTATGGCTTCTCGCATACCGTGGGCCGCTCCGCAGGGA 2735

2736 TCGCTGGTCCTCGGTGGTACCCCGCGTGGTGGAAGTGAACAAGAAGTGAACCCGGACGA 2795

2796 GGTGGTGGTGCCTCTGGACAGCATGGGGAACCCCGCTGCGATGGCCACCAGCAGGGTTA 2855

2856 CCCCCGCAAGTGGAGGACTGATGACGCCCGCTCtagggactgcagcccagcccagctt 2915

2916 ctctgcccaactcattttctagtccagccgcatttcagcagtgcccttctggggtgtcccccc 2975

2976 acaccctgctttggccccagagggcgagggaccagtgagggtgccagggaggccccaggac 3035

3036 cctgtgggtcccctggctctgcctccccaccctggggtgggggctcccggccacctgtctt 3095

3096 gctcctatggagtcacataagccaacgccagagccctccacctcaggccccagcccctg 3155

3156 cctctccattattttatttgcctctgctctcaggaagcgacgtgaccctgccccagctgga 3215

3216 acctggcagaggccttaggaacccggttccaagtgcactgcccggccaagccccagcctca 3275

3276 gcctgcgcctgagctgcatgogccaccatttttggcagcgtggcagctttgcaaggggct 3335

3336 ggggcccctcggcgtggggccatgccttctgtgtgttctgtagtgtctgggatttgccggt 3395

3396 gctcaataaatgtttatttcattgaaaaaaaaaaaaaa 3433

FIG. 17 CONT'D



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## FIG. 18

NUCLEOTIDE AND AMINO ACID SEQUENCE OF hVR3  
INCLUDING THE 5'UTR (nt -684 TO nt 0), CODING REGION (nt1  
TO 2889) AND 3'UTR (nt 2890 TO nt 3418)

-684	ttacgcgttaagaaatacccaagcttatgcatcaagcttggtaccgagctcggatccact	-625
-624	agtaccgccggccagtgctgctggaattcaaggtgaggagaggagcatggatcctgggagc	-565
-564	gagtgtgtgcaggccagggagggcctttccagaggagcccagttgagctggaacaccagt	-505
-504	gggaggagttgaccagcaaaggtgcaggggaggatcagcactttgcactggggagcagag	-445
-444	tttgtgcactggggaagtcaactcaagtattggagcctcagtttctgttctgtaaaatg	-385
-384	ggttcatcatgacagtggttgatgaggaaaaggactgcccgcctacacagcaagtccaca	-325
-324	tggattttctgagccccctcctgtgcctgaagccccacgggttaatggttctgccttagcagg	-265
-264	tgettaccacgtgccaggcactgcactgcactggccactggactgcatgttctgtccatg	-205
-204	aggcttgatataccccatcttacagatcaggaagctgaggctatgaaatgtcgacttgct	-145
-144	caatgtcatggaatgactaagtgtggagcctggatttgaacttggctctctggggctcca	-85
-84	aagctggctttcttggtcagcagtagggtctgggatccaagtatgggggtcccagcttgac	-25
-24	cctgaagtccaccctctttcagctaATGCCAGGGTAGTTGGACCTGGGGCCAATTTGTG	35
1	M P R V V G P G A N L C	12
36	TTTCCAGGTTTCGTGAAAGAGGCTCCTGTGTCAGTTCCCCGCCTGAGGCTGGCGGCCAACCA	95
13	F Q V R E R G S C C S S R L R L A A N H	32
96	CATCTGGGAGTGGCCTCCCTGTGCCCCCTGTCATTACAACGGTGGCTTTGAAGCAGCTGGC	155
33	I W E W P P C A P V I T T V A L K Q L A	52
156	AGCACTGCTGCTTGTCCACGTGGGAGGGGGCTTCCTGGAGCCCCCGCCCCTGGCCGGGTT	215
53	A L L L V H V G G G F L E P P P L A G F	72
216	CTGCCTGACTCCCCTTTTCATTCCCTTGCAGGCTGAGCAGTGCAGACGGGCCTGGGGCAGG	275
73	C L T P L S F P C R L S S A D G P G A G	92
276	CATGGCGGATTCCAGCGAAGGCCCGCGCGGGGGCGGGGAGGTGGCTGAGCTCCCCGG	335
93	M A D S S E G P R A G P G E V A E L P G	112
336	GGATGAGAGTGGCACCCCAGGTGGGGAGGCTTTTCTCTCTCCTCCCTGGCCAATCTGTT	395
113	D E S G T P G G E A F P L S S L A N L F	132
396	TGAGGGGGAGGATGGCTCCCTTTTCGCCCTCACCGGCTGATGCCAGTCGCCCTGCTGGCCC	455
133	E G E D G S L S P S P A D A S R P A G P	152
456	AGGCGATGGGCGACCAAATCTGCGCATGAAGTTCCAGGGCGCCTTCCGCAAGGGGGTGCC	515
153	G D G R P N L R M K F Q G A F R K G V P	172
516	CAACCCCATCGATCTGCTGGAGTCCACCCTATATGAGTCCTCGGTGGTGCCTGGGCCCAA	575
173	N P I D L L E S T L Y E S S V V P G P K	192

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576 GAAAGCACCCATGGACTCACTGTTTGACTACGGCACCTATCGTCACCACTCCAGTGACAA 635  
193 K A P M D S L F D Y G T Y R H H S S D N 212

636 CAAGAGGTGGAGGAAGAAGATCATAGAGAAGCAGCCGAGAGCCCCAAAGCCCCTGCCCC 695  
213 K R W R K K I I E K Q P Q S P K A P A P 232

696 TCAGCCGCCCCCATCCTCAAAGTCTTCAACCGGCCTATCCTCTTTGACATCGTGTCCCG 755  
233 Q P P P I L K V F N R P I L F D I V S R 252

756 GGGCTCCACTGCTGACCTGGACGGGCTGCTCCCATTTCTTGCTGACCCACAAGAAACGCCT 815  
253 G S T A D L D G L L P F L L T H K K R L 272

816 AACTGATGAGGAGTTTCGAGAGCCATCTACGGGAAGACCTGCCTGCCCAAGGCCTTGCT 875  
273 T D E E F R E P S T G K T C L P K A L L 292

876 GAACCTGAGCAATGGCCGCAACGACACCATCCCTGTGCTGCTGGACATCGCGGAGCGCAC 935  
293 N L S N G R N D T I P V L L D I A E R T 312

936 CGGCAACATGCGGGAGTTCATTAAGTTCGCGCTTCCGTGACATCTACTATCGAGGTCAGAC 995  
313 G N M R E F I N S P F R D I Y Y R G Q T 332

996 AGCCCTGCACATCGCCATTGAGCGTCGCTGCAAACACTACGTGGAACCTTCTCGTGGCCCA 1055  
333 A L H I A I E R R C K H Y V E L L V A Q 352

1056 GGGAGCTGATGTCCACGCCCAGGCCCGTGGGCGCTTCTTCCAGCCCAAGGATGAGGGGGG 1115  
353 G A D V H A Q A R G R F F Q P K D E G G 372

1116 CTACTTCTACTTTGGGGAGCTGCCCCTGTGCTGCTGCTGACCAACCAGCCCCACAT 1175  
373 Y F Y F G E L P L S L A A C T N Q P H I 392

1176 TGTCAACTACCTGACGGAGAACCCCCACAAGAAGGCGGACATGCGGCGCCAGGACTCGCG 1235  
393 V N Y L T E N P H K K A D M R R Q D S R 412

1236 AGGCAACACAGTGTGTCATGCGCTGGTGGCCATTGCTGACAACACCCGTGAGAACACCAA 1295  
413 G N T V L H A L V A I A D N T R E N T K 432

1296 GTTTGTTACCAAGATGTACGACCTGCTGCTGCTCAAGTGTGCCCCGCTCTTCCCCGACAG 1355  
433 F V T K M Y D L L L L K C A R L F P D S 452

1356 CAACCTGGAGGCCGTGCTCAACAACGACGGCCTCTCGCCCCTCATGATGGCTGCCAAGAC 1415  
453 N L E A V L N N D G L S P L M M A A K T 472

1416 GGGCAAGATTGGGATCTTTTCAGCACATCATCCGGCGGGAGGTGACGGATGAGGACACACG 1475  
473 G K I G I F Q H I I R R E V T D E D T R 492

1476 GCACCTGTCCCGCAAGTCCAAGGACTGGGCCTATGGGCCAGTGATTCTCTCGCTTTATGA 1535  
493 H L S R K S K D W A Y G P V Y S S L Y D 512

1536 CCTCTCCTCCCTGGACACGTGTGGGGAAGAGGCCTCCGTGCTGGAGATCCTGGTGTACAA 1595  
513 L S S L D T C G E E A S V L E I L V Y N 532

1596 CAGCAAGATTGAGAACCGCCACGAGATGCTGGCTGTGGAGCCCCATCAATGAACTGCTGCG 1655  
533 S K I E N R H E M L A V E P I N E L L R 552

1656 GGACAAGTGGCGGAAGTTCGGGGCCGTCTCCTTCTACATCAACGTGGTCTCCTACCTGTG 1715  
553 D K W R K F G A V S F Y I N V V S Y L C 572

FIG. 18<sub>CONT'D</sub>

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1716 TGCCATGGTTATCTTCACTCTCACCGCCTACTACCAGCCGCTGGAGGGCACACCGCCGTA 1775  
573 A M V I F T L T A Y Y Q P L E G T P P Y 592

1776 CCCTTACCGCACCACGGTGGACTACCTGCGGCTGGCTGGCGAGGTCATTACGCTCTTCAC 1835  
593 P Y R T T V D Y L R L A G E V I T L F T 612

1836 TGGGGTCCTGTTCTTCTTACCAACATCAAAGACTTGTTTCATGAAGAAATGCCCTGGAGT 1895  
613 G V L F F F T N I K D L F M K K C P G V 632

1896 GAATTCTCTCTTCATTGATGGCTCCTTCCAGCTGCTCTACTTCATCTACTCTGTCTGGT 1955  
633 N S L F I D G S F Q L L Y F I Y S V L V 652

1956 GATCGTCTCAGCAGCCCTCTACCTGGCAGGGATCGAGGCCTACCTGGCCATGATGGTCTT 2015  
653 I V S A A L Y L A G I E A Y L A M M V F 672

2016 TGCCCTGGTCTCTGGGCTGGATGAATGCCCTTTACTTCACCCGTGGGCTGAAGCTGACGGG 2075  
673 A L V L G W M N A L Y F T R G L K L T G 692

2076 GACCTATAGCATCATGATCCAGAAGATTCTCTTCAAGGACCTTTTCCGATTCTGCTCGT 2135  
693 T Y S I M I Q K I L F K D L F R F L L V 712

2136 CTACTTGCTCTTCATGATCGGCTACGCTTCAGCCCTGGTCTCCCTCCTGAACCCGTGTGC 2195  
713 Y L L F M I G Y A S A L V S L L N P C A 732

2196 CAACATGAAGGTGTGCAATGAGGACCAGACCAACTGCACAGTGCCCACTTACCCCTCGTG 2255  
733 N M K V C N E D Q T N C T V P T Y P S C 752

2256 CCGTGACAGCGAGACCTTCAGCACCTTCTCCTGGACCTGTTTAAGCTGACCATCGGCAT 2315  
753 R D S E T F S T F L L D L F K L T I G M 772

2316 GGGCGACCTGGAGATGCTGAGCAGCACCAAGTACCCCGTGGTCTTCATCATCCTGCTGGT 2375  
773 G D L E M L S S T K Y P V V F I I L L V 792

2376 GACCTACATCATCCTCACCTCTGTGCTGCTCCTCAACATGCTCATTGCCCTCATGGGCGA 2435  
793 T Y I I L T S V L L L N M L I A L M G E 812

2436 GACAGTGGGCCAGGTCTCCAAGGAGAGCAAGCACATCTGGAAGCTGCAGTGGGCCACCAC 2495  
813 T V G Q V S K E S K H I W K L Q W A T T 832

2496 CATCCTGGACATTGAGCGCTCCTTCCCCGTATTCTCTGAGGAAGGCCTTCCGCTCTGGGGA 2555  
833 I L D I E R S F P V F L R K A F R S G E 852

2556 GATGGTCACCGTGGGCAAGAGCTCGGACGGCACTCCTGACCGCAGGTGGTGCTTCAGGGT 2615  
853 M V T V G K S S D G T P D R R W C F R V 872

2616 GGATGAGGTGAACTGGTCTCACTGGAACCAGAACTTGGGCATCATCAACGAGGACCCGGG 2675  
873 D E V N W S H W N Q N L G I I N E D P G 892

2676 CAAGAATGAGACCTACCAGTATTATGGCTTCTCGCATACCGTGGGCCCGCTCCGCAGGGA 2735  
893 K N E T Y Q Y Y G F S H T V G R L R R D 912

2736 TCGCTGGTCTCGGTGGTACCCCGCGTGGTGGAACTGAACAAGAACTCGAACCCGGACGA 2795  
913 R W S S V V P R V V E L N K N S N P D E 932

2796 GGTGGTGGTGCTCTGGACAGCATGGGGAACCCCGCTGCGATGGCCACCAGCAGGGTTA 2855  
933 V V V P L D S M G N P R C D G H Q Q G Y 952

FIG. 18<sub>CONT'D</sub>

2856 CCCCCGCAAGTGGAGGACTGATGACGCCCCGCTCtagggactgcagcccagcccagctt 2915  
953 P R K W R T D D A P L 963

2916 ctctgcccactcatttctagtccagccgcatttcagcagtgcccttctgggggtgtcccccc 2975

2976 acaccctgctttggccccagaggcgagggaccagtgagggtgccagggaggccccaggac 3035

3036 cctgtgggtcccctggctctgcctccccaccctgggggtgggggtccccggccacctgtctt 3095

3096 gctcctatggagtcacataagccaacgccagagcccctccacctcaggccccagcccctg 3155

3156 cctctccattatttatttgctctgctctcaggaagcgacgtgacccctgccccagctgga 3215

3216 acctggcagaggccttaggaccccggtccaagtgcactgcccggccaagccccagcctca 3275

3276 gcctgcgcctgagctgcatgcgccaccatTTTTGGCAGCGTGGCAGCTTTGCAAGGGGCT 3335

3336 ggggccctcggcgtggggccatgccttctgtgtgttctgtagtgtctgggatttgccggt 3395

3396 gctcaataaaatgtttatttcattgaaaaaaaaaaaaaaaaa 3433

FIG. 18<sub>CONT'D</sub>


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## FIG. 19

## AMINO ACID SEQUENCE OF hVR3

1 MPRVVGPAN LCFQVRERGS CCSSRLRLAA NHIWEWPPCA PVITTVALKQ  
 51 LAALLLVHVG GGFLEPPPLA GFCLTPLSFP CRLSSADGPG AGMADSSEGP  
 101 RAGPGEVAEL PGDESGTPGG EAFPLSSIAN LFEGEDGSLS PSPADASRPA  
 151 GPGDGRPNLR MKFQGAFRKG VPNPIDLLES TLYESSVVPK PKKAPMDSL  
 201 DYGTYRHHSS DNKRWRKKII EKQPQSPKAP APQPPPIKV FNRPIFLDIV  
 251 SRGSTADLDG LLPFLLTHKK RLTDEEFREP STGKTCLPKA LLNLSNGRND  
 301 TIPVLLDIAE RTGNMREFIN SPFRDIYYRG QTALHIAIER RCKHYVELLV  
 351 AQQADVHAQA RGRFFQPKDE GGYFYFEGELP LSLAACTNQP HIVNYLTENP  
 401 HKKADMRRQD SRGNTVLHAL VAIADNTREN TKFVTKMYDL LLLKCARLFP  
 451 DSNLEAVLNN DGLSPLMAA KTGKIGIFQH IIRREVTDED TRHLSRKS  
 501 WAYGPVYSSL YDLSSLDTCG EEASVLEILV YNSKIENRHE MLAVEPINEL  
 551 LRDKWRKFGA VSEYINVVSYSICAMVIEETITAYVOPLEGTP PYPYRTTVDY  
 601 LRLAGEVETITETIGVIEETITETIKDLFMKKCP GVNSLFIDGSFQHHYETISV  
 651 IVIVSAALYD AGIEAYLAMM VEAIVIGWMNSALYETRCIKITCTLYSIMEOK  
 701 ILFKDIREFITIVYIEEMIGYASALVSLNLP CANMKVCNED QTNCTVPTYP  
 751 SCRDSSETFST FLDDLFLKTI GMGDLEMLSS TKYPAVETITIVYIETTSV  
 801 LILNMITALMAGETVGQVSKE SKHIWKLQWA TTILDIERSF PVFLRKAFRS  
 851 GEMVTVGKSS DGTPDRRWCF RVDEVNWSHW NQNLGIINED PGKNETYQYY  
 901 GFSHTVGRLR RDRWSSVVR VVELNKNSNP DEVVVPLDSM GNPRCDGHQO  
 951 GYPRKWRTDD APL

## Key

 Transmembrane domains


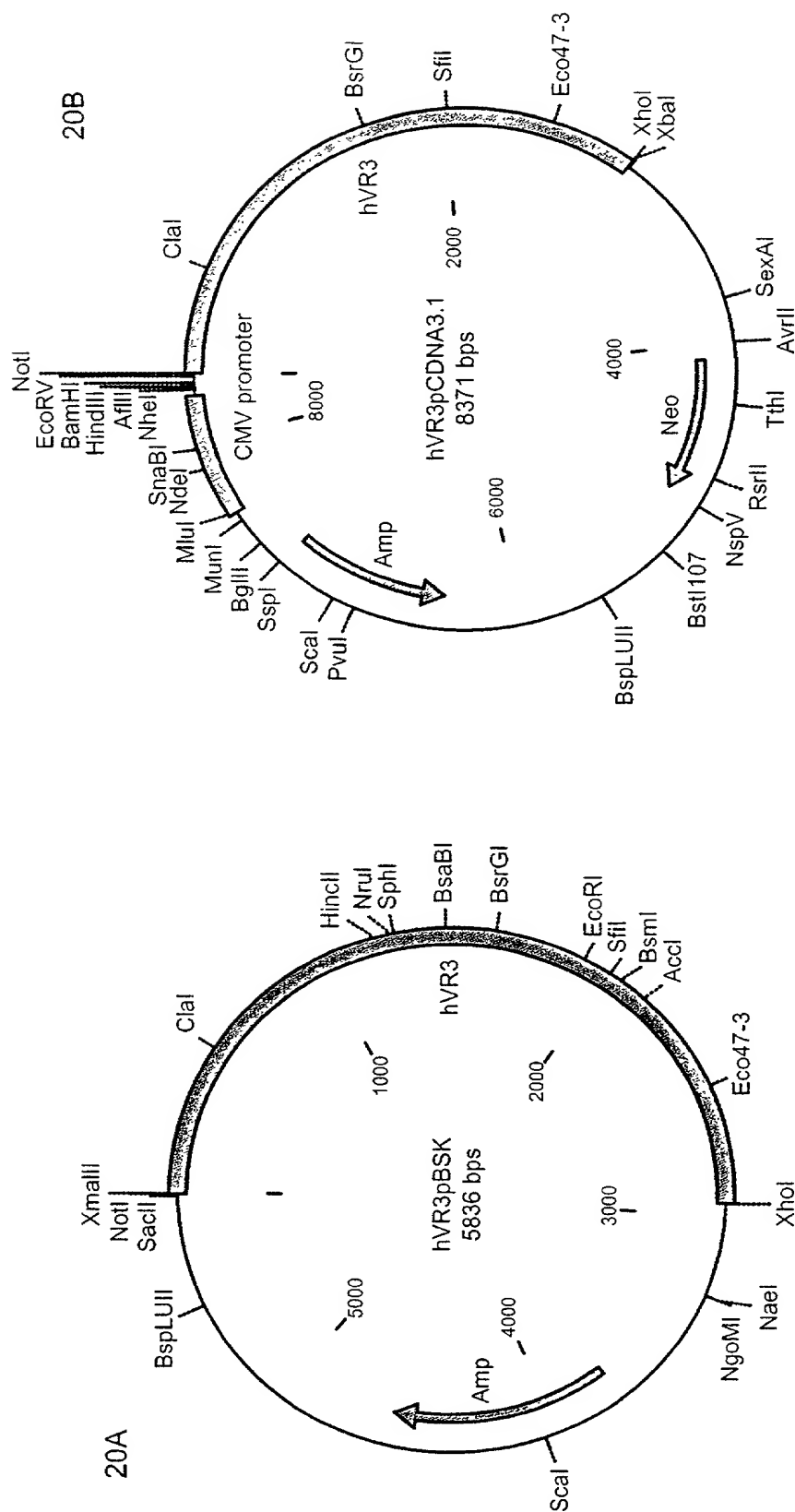
 Ankyrin binding domains

FIG. 20

FULL-LENGTH hVR3 CLONED INTO (A) pBLUESCRIPT SK(+) (hVR3pBSK) AND  
(B) pCDNA3.1(+) (hVR1pCDNA3.1) VIA NotI/XhoI RESTRICTION SITES.



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## FIG. 21

A MULTIPLE COMPARISON OF THE AMINO ACID SEQUENCES OF THE RAT VR1 AND THE HUMAN VANILLOID RECEPTORS, hVR1, hVRL-1 AND hVR3

	10	20	30	40	50
VR1	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
hVR1	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
hVRL-1	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
hVR3	MPRVVGP	GANLCFQV	RERGSCCSSRLRLA	ANHIWEWPPCAPVIT	TVALKQ
	60	70	80	90	100
VR1	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
hVR1	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
hVRL-1	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
hVR3	LAALLLVH	VGGGFLEPP	PLAGFCLT	PLSFPCRLSSADG	PGAGMADSSEGP
	110	120	130	140	150
VR1	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
hVR1	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
hVRL-1	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
hVR3	RAGPGEVA	ELPGDESGT	PGGEAFPLSS	LANLFEGEDGSL	SPSPADASRPA
	160	170	180	190	200
VR1	DPPDRDP	NCNCKPPV	KPHIFTT	RSRTRLF	... KGDSEEASPLDCPYEEG
hVR1	DPLDGD	PNSRPPAK	PQLSTAK	SRTRLFG	... KGDSEEAFVDCPHEEG
hVRL-1	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
hVR3	GGGGRN	LRMKFQGA	FRKGVNP	.... IDLLESTLY	ESSVVP
	210	220	230	240	250
VR1	GLASCP	ITVSSVLT	IQRPDGP	ASVREPS	QDSVSAG.EKP.PR
hVR1	ELDSCP	ITVSPVIT	IQRPDGP	TGARLLS	QDSVAASTEKT.LR
hVRL-1	GSGLPP	..ESQFQ	GEDRKFA	PQIRVNL	NYRKGTGASQPD
hVR3	MDSLED	YGYRHH	SSDNKR	WRKKIIE	KQPQSPKAPAPQ
	260	270	280	290	300
VR1	IFDAVA	QSNCOE	LESLLP	FLQSKK	RLTDSEETD
hVR1	IFEAVA	QNNCOD	LESLLL	FLQSKK	HLTDNEFKD
hVRL-1	LENVSR	GVPEDL	AGLPEY	LSKTSK	YLTDSYTE
hVR3	LEDIVS	RGSTAD	LDGLLP	FLTHKK	RLTDEEF
	310	320	330	340	350
VR1	NGNDT	IALLLD	VARKT	DSLKQ	FVNASYTDSY
hVR1	DGONT	TIPILL	IEARQ	TDSEL	VNASYTDSY
hVRL-1	DGVNA	CILPLL	QIDR	SGNPQ	PLVNAQCTD
hVR3	NGRND	TIPVLL	IAERT	GNMRE	FINSPPFDI
	360	370	380	390	400
VR1	VTLIV	ENGADV	QAAANG	DFFKTK	GRPGFYF
hVR1	VTLIV	ENGADV	QAAAHG	DFFKTK	GRPGFYF
hVRL-1	VKLLV	ENGANV	HARACG	RFFQK	GQG.TCFYF
hVR3	VELLVA	QGADV	HQAARG	RFFQPK	DEGGYFYF
	410	420	430	440	450
VR1	LLONSW	OPADIS	ARDSV	GNITV	LHALVE
hVR1	LLONSW	QADIS	ARDSV	GNITV	LHALVE
hVRL-1	LLENPH	QASLQ	ATDSQ	GNITV	LHALVMI
hVR3	LTENPH	KKADM	RRODS	RGNITV	LHALVA
	460	470	480	490	500
VR1	AKLHPT	LKLEE	ITNRK	GLTPLA	LAASSG
hVR1	AKLHPT	LKLEE	ITNRK	GLTPLA	LAAGTG
hVRL-1	ARLCPT	VOLED	IRNLQ	DLTPLK	LAKEG
hVR3	ARLFPS	DNLEA	VLNND	GLSPLM	MAAKT

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510 520 530 540 550  
 VR1 RKFTWAYGPVHSSLYDLSCIDTC.EKNSVLEVIAYSSSETPNRHDMMLV  
 hVR1 RKFTWAYGPVHSSLYDLSCIDTC.EKNSVLEVIAYSSSETPNRHDMMLV  
 hVRL-1 RKFTWCYGPVRVSLYDLASVDSC.EENSVEIIAF.HCKSPHRHRMVVL  
 hVR3 RKSKDWAYGPVYSSLYDLSSLDTCGEEASVLEILVY.WSKIENRHEMLAV  
 560 570 580 590 600  
 VR1 EPLNRLLQDKWDRFVKRIFYNFVYCLYMIIFTAAAYYRPV..EGLPPY  
 hVR1 EPLNRLLQDKWDRFVKRIFYNFVYCLYMIIFTMAAYYRPV..DGLPPF  
 hVRL-1 EPLNKLQAKWDLIPK.FFLNFLCNLIYMFIFTAVAYHQPTLKKQAAPH  
 hVR3 EPINELLRDKWRKFGAVSFYINVVSYLCAVIFTLTAYYQPL..EGTPPY  
 610 620 630 640 650  
 VR1 KLKNTVGDYFRVTGEILSVSGGVYFFFRGIQ.YFLQRRPSLKSLEFVDSYS  
 hVR1 KMEN.IGDYFRVTGEILSVLGGVYFFFRGIQ.YFLQRRPSMKTLEFVDSYS  
 hVRL-1 .LNAEVGNSMLLTGHILILLGGIYLLVGQLW.YFWRRHVFIWISFIDSYF  
 hVR3 PYRTTV.DYLRAGEVITLFTGVLEFFFTNIKDLFMKKCPGVNSLFIDGSE  
 660 670 680 690 700  
 VR1 EILFFVQSLFMLVSVVLYFSQRKEYVASMVFSLAMGWTNMLYYTRGFQOM  
 hVR1 EMLFFLQSLFMLATVVLYFSLHKEYVASMVFSALGWTNMLYYTRGFQOM  
 hVRL-1 EILFLFQALLTVVSQVLCFLAIEWYPLLVSALVGLWLNLLYYTRGFQHT  
 hVR3 QLLFYIYSVLIVVSAALYLAGIEAYLAMVFAVLGWNALYFTRGLKLT  
 710 720 730 740 750  
 VR1 GIYAVMIEKMILRDLCREMFVYLVFLEGFSTAVVTLIEDGKNSLP....  
 hVR1 GIYAVMIEKMILRDLCREMFVYIVFLEGFSTAVVTLIEDGKNSLP....  
 hVRL-1 GIYSVMIQKVLRLDLLRFLLIYLVFLEGFFAVALVSLSQEAWRPEAPTGP  
 hVR3 GTYSIMIQLFKDLFRFLLVYLLEMIGYASALVSLNPCANMKVCNEDQ  
 760 770 780 790 800  
 VR1 MESTPHKCRGSACK.PGNSYNSLYSTCLELEFKFTIGMGDLEFTENYDFKA  
 hVR1 SESTSHRWGPACRPDPSSYNSLYSTCLELEFKFTIGMGDLEFTENYDFKA  
 hVRL-1 ATESVQPMEGQDEGNGAQYRGILEASLELEFKFTIGMGELAFQEQHLFRG  
 hVR3 TNCTVPTY..PSCR.DSETFSTFL...LDLFLKTIGMGDLEMLSSTKYPV  
 810 820 830 840 850  
 VR1 VFIILLAYVILTYILLNMLIALMGETVNKIAQESKNIWKLOQRAITILD  
 hVR1 VFIILLAYVILTYILLNMLIALMGETVNKIAQESKNIWKLOQRAITILD  
 hVRL-1 MVLLLLAYVILTYILLNMLIALMSETVNSVATDSWSIWKLOKAISVLE  
 hVR3 VFIILLVTYIILTSVILLNMLIALMGETVGVQSKE SKHIWKLOWATTILD  
 860 870 880 890 900  
 VR1 TEKSFLKCMRKAFRSGKLLQVGETPDGKDDYRWCFRVDEVNWTWNTNVG  
 hVR1 TEKSELKCMRKAFRSGKLLQVGYTPDGKDDYRWCFRVDEVNWTWNTNVG  
 hVRL-1 MENGYYWC.RKKORAGVMLTVGTPDGSPDERWCFRVEEVNWSWEQTLF  
 hVR3 IERSFPVFLRKAFRSGEMVTVGKSSDGTDDRWCFRVDEVNWSHWNQNLG  
 910 920 930 940 950  
 VR1 IINEDPGNCE.....GVKRTLSFSLSRSG.....RVSGRNWKNFALV  
 hVR1 IINEDPGNCE.....GVKRTLSFSLSRSS.....RVSGRHWKNFALV  
 hVRL-1 TLCEDPSGA.....GVPRTLNPNVLAS.....PPKEDDGASEENYVPV  
 hVR3 IINEDPGKWETYQYYGFSHTVGRLLRRDRWSSVVPVVELNKNNSNPDEVVV  
 960 970 980 990  
 VR1 PLLRDASTDRRHATQOEVOVKHYTGSLKPEDAEVFKDSMVPGEN  
 hVR1 PLLREASARDRQSAQPEEVYLRQFSGSLKPEDAEVFKSPAASGEN  
 hVRL-1 QLLQSN~~~~~  
 hVR3 PLDSMGNPRCDGHQGYPRKWRTDDAPL~~~~~

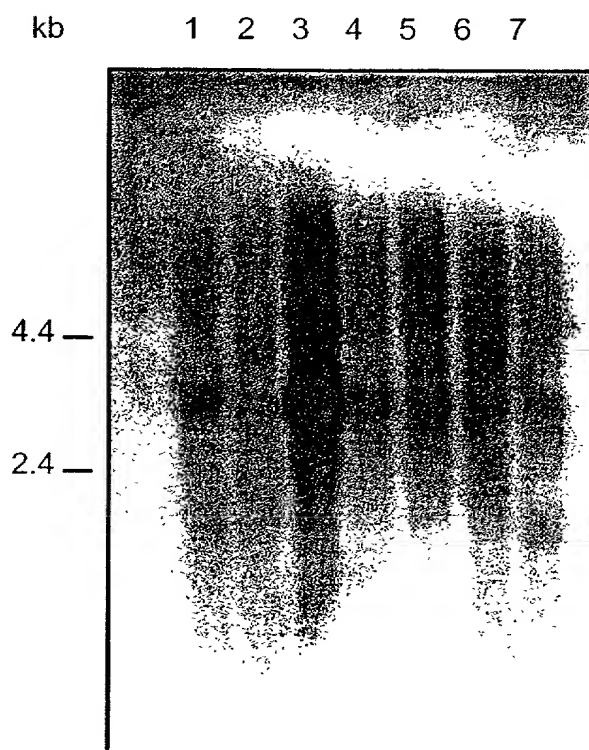
FIG. 21 CONT'D



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**FIG. 22A**

HYBRIDISATION OF A NORTHERN BLOT WITH hVR3



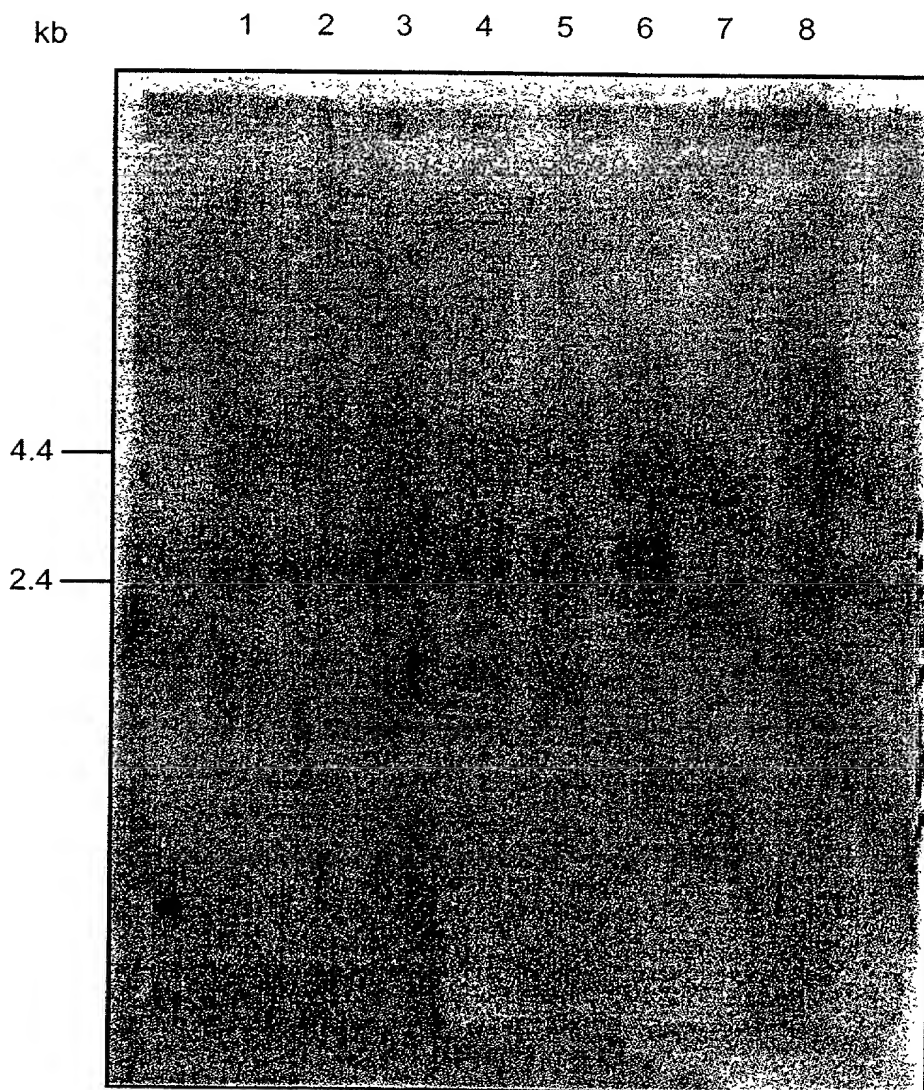
LANE 1: BONE MARROW  
LANE 2: ADRENAL GLAND  
LANE 3: TRACHEA  
LANE 4: LYMPH NODE

LANE 5: SPINAL CORD  
LANE 6: THYROID  
LANE 7: STOMACH

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## FIG. 22B

HYBRIDISATION OF NORTHERN BLOT WITH hVR3 PROBE



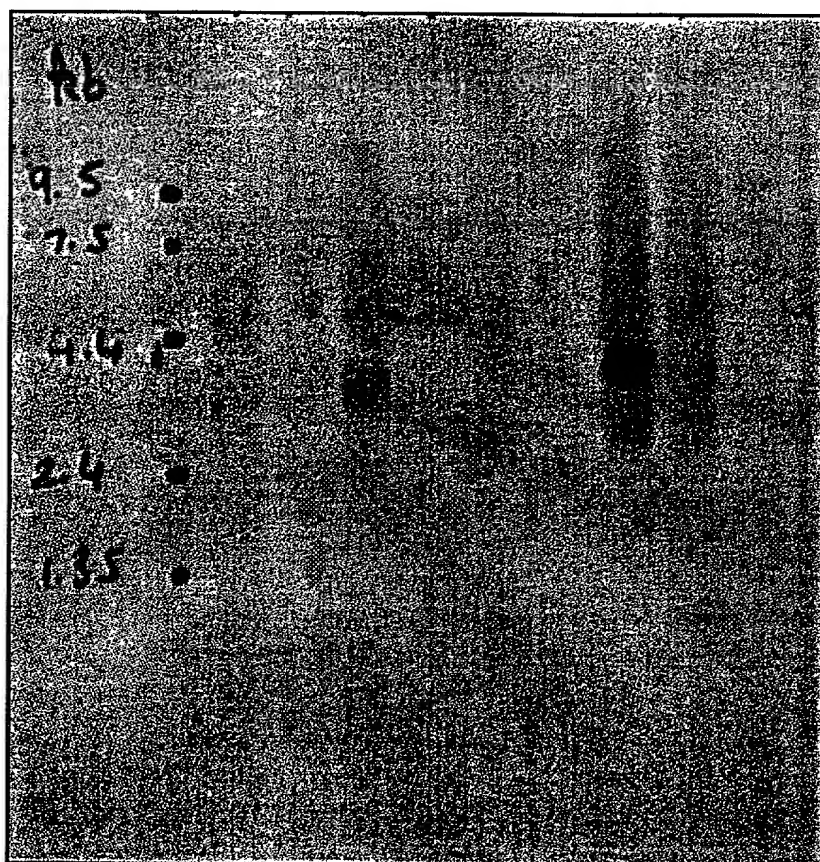
LANE 1: PERIPHERAL BLOOD  
LEUKOCYTE  
LANE 2: COLON  
LANE 3: SMALL INTESTINE  
LANE 4: UTERUS  
LANE 5: TESTIS  
LANE 6: PROSTATE  
LANE 7: THYROID  
LANE 8: SPLEEN

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## FIG. 22C

HYBRIDISATION OF A MULTI-TISSUE NORTHERN  
BLOT WITH THE hVR3 PROBE

1 2 3 4 5 6 7 8



LANE 1: HEART  
LANE 2: BRAIN  
LANE 3: PLACENTA  
LANE 4: LUNG

LANE 5: LIVER  
LANE 6: SKELETAL MUSCLE  
LANE 7: KIDNEY  
LANE 8: PANCREAS

## SEQUENCE LISTING

<110> Glaxo Group Ltd  
Tate, Simon N  
Delany, Natalie S  
Sanseau, P

<120> Novel Receptors

<130> PG3606

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<150> GB 9826359.3

<151> 1998-12-01

<160> 40

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tgtataagct cagtggctgt ggcagcgagg ttgaagagca aaggcaggcc gggcacctgg 180

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 Lys Lys Trp Ser Ser Thr Asp Leu Gly Ala Ala Ala Asp Pro Leu Gln  
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 Lys Asp Thr Cys Pro Asp Pro Leu Asp Gly Asp Pro Asn Ser Arg Pro  
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 Pro Pro Ala Lys Pro Gln Leu Ser Thr Ala Lys Ser Arg Thr Arg Leu  
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 Ile Thr Ile Gln Arg Pro Gly Asp Gly Pro Thr Gly Ala Arg Leu Leu  
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                     115                    120                    125

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 Asp Asn Glu Phe Lys Asp Pro Glu Thr Gly Lys Thr Cys Leu Leu Lys  
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 Ala Met Leu Asn Leu His Asp Gly Gln Asn Thr Thr Ile Pro Leu Leu  
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 230 235 240

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 Gly Arg Pro Gly Phe Tyr Phe Gly Glu Leu Pro Leu Ser Leu Ala Ala  
 245 250 255

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cac gcc ctg gtg gag gtg gcc gac aac acg gcc gac aac acg aag ttt 1689  
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 Pro Thr Leu Lys Leu Glu Glu Leu Thr Asn Lys Lys Gly Met Thr Pro  
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1593 1641 1689 1737 1785 1833

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aga cac tgg aag aac ttt gcc ctg gtc ccc ctt tta aga gag gca agt 3177  
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gct cga gat agg cag tct gct cag ccc gag gaa gtt tat ctg cga cag 3225  
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gcc gct tcc ggg gag aag tga ggacgtcacg cagacagcac tgtcaacact 3324  
Ala Ala Ser Gly Glu Lys  
835

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&lt;211&gt; 839

&lt;212&gt; PRT

&lt;213&gt; Homo sapiens

&lt;400&gt; 2

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Leu Phe Gly Lys Gly Asp Ser Glu Glu Ala Phe Pro Val Asp Cys Pro  
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His Glu Glu Gly Glu Leu Asp Ser Cys Pro Thr Ile Thr Val Ser Pro  
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Gly Ala Asp Val Gln Ala Ala Ala His Gly Asp Phe Phe Lys Lys Thr  
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Trp Gln Thr Ala Asp Ile Ser Ala Arg Asp Ser Val Gly Asn Thr Val  
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Phe Val Thr Ser Met Tyr Asn Glu Ile Leu Ile Leu Gly Ala Lys Leu  
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His Pro Thr Leu Lys Leu Glu Glu Leu Thr Asn Lys Lys Gly Met Thr  
325 330 335

Pro Leu Ala Leu Ala Ala Gly Thr Gly Lys Ile Gly Val Leu Ala Tyr  
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Glu Pro Leu Asn Arg Leu Leu Gln Asp Lys Trp Asp Arg Phe Val Lys  
420 425 430

Arg Ile Phe Tyr Phe Asn Phe Leu Val Tyr Cys Leu Tyr Met Ile Ile  
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Phe Thr Met Ala Ala Tyr Tyr Arg Pro Val Asp Gly Leu Pro Pro Phe  
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Leu Gln Arg Arg Pro Ser Met Lys Thr Leu Phe Val Asp Ser Tyr Ser  
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Glu Met Leu Phe Phe Leu Gln Ser Leu Phe Met Leu Ala Thr Val Val  
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Leu Ala Leu Gly Trp Thr Asn Met Leu Tyr Tyr Thr Arg Gly Phe Gln  
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Gln Met Gly Ile Tyr Ala Val Met Ile Glu Lys Met Ile Leu Arg Asp  
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Leu Cys Arg Phe Met Phe Val Tyr Ile Val Phe Leu Phe Gly Phe Ser  
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Thr Ala Val Val Thr Leu Ile Glu Asp Gly Lys Asn Asp Ser Leu Pro  
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Asp Ser Ser Tyr Asn Ser Leu Tyr Ser Thr Cys Leu Glu Leu Phe Lys  
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Phe Thr Ile Gly Met Gly Asp Leu Glu Phe Thr Glu Asn Tyr Asp Phe  
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Gln Glu Asn Ser Cys Leu Asp Pro Pro Asp Arg Asp Pro Asn Cys Lys  
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Tyr Glu Glu Gly Gly Leu Ala Ser Cys Pro Ile Ile Thr Val Ser Ser  
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Val Leu Thr Ile Gln Arg Pro Gly Asp Gly Pro Ala Ser Val Arg Pro  
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Ser Ser Gln Asp Ser Val Ser Ala Gly Glu Lys Pro Pro Arg Leu Tyr  
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Asp Arg Arg Ser Ile Phe Asp Ala Val Ala Gln Ser Asn Cys Gln Glu  
115 120 125

Leu Glu Ser Leu Leu Pro Phe Leu Gln Arg Ser Lys Lys Arg Leu Thr  
130 135 140

Asp Ser Glu Phe Lys Asp Pro Glu Thr Gly Lys Thr Cys Leu Leu Lys  
145 150 155 160

Ala Met Leu Asn Leu His Asn Gly Gln Asn Asp Thr Ile Ala Leu Leu  
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Leu Asp Val Ala Arg Lys Thr Asp Ser Leu Lys Gln Phe Val Asn Ala  
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Ser Tyr Thr Asp Ser Tyr Tyr Lys Gly Gln Thr Ala Leu His Ile Ala  
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Ile Glu Arg Arg Asn Met Thr Leu Val Thr Leu Leu Val Glu Asn Gly  
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Ala Asp Val Gln Ala Ala Ala Asn Gly Asp Phe Phe Lys Lys Thr Lys  
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485 490 495

Leu Gln Arg Arg Pro Ser Leu Lys Ser Leu Phe Val Asp Ser Tyr Ser  
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Glu Ile Leu Phe Phe Val Gln Ser Leu Phe Met Leu Val Ser Val Val  
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Leu Tyr Phe Ser Gln Arg Lys Glu Tyr Val Ala Ser Met Val Phe Ser  
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Leu Ala Met Gly Trp Thr Asn Met Leu Tyr Tyr Thr Arg Gly Phe Gln  
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Gln Met Gly Ile Tyr Ala Val Met Ile Glu Lys Met Ile Leu Arg Asp  
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Leu Cys Arg Phe Met Phe Val Tyr Leu Val Phe Leu Phe Gly Phe Ser  
580 585 590

Thr Ala Val Val Thr Leu Ile Glu Asp Gly Lys Asn Asn Ser Leu Pro  
595 600 605

Met Glu Ser Thr Pro His Lys Cys Arg Gly Ser Ala Cys Lys Pro Gly  
610 615 620

Asn Ser Tyr Asn Ser Leu Tyr Ser Thr Cys Leu Glu Leu Phe Lys Phe  
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Asp Glu Ser Gly Thr Pro Gly Gly Glu Ala Phe Pro Leu Ser Ser Leu  
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Gln Ser Pro Lys Ala Pro Ala Pro Gln Pro Pro Pro Ile Leu Lys Val  
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Thr Asp Glu Glu Phe Arg Glu Pro Ser Thr Gly Lys Thr Cys Leu Pro  
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Lys Ala Leu Leu Asn Leu Ser Asn Gly Arg Asn Asp Thr Ile Pro Val  
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Leu Leu Asp Ile Ala Glu Arg Thr Gly Asn Met Arg Glu Phe Ile Asn  
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340 345 350

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Ala Cys Thr Asn Gln Pro His Ile Val Asn Tyr Leu Thr Glu Asn Pro  
385 390 395 400

His Lys Lys Ala Asp Met Arg Arg Gln Asp Ser Arg Gly Asn Thr Val  
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Leu His Ala Leu Val Ala Ile Ala Asp Asn Thr Arg Glu Asn Thr Lys  
420 425 430

Phe Val Thr Lys Met Tyr Asp Leu Leu Leu Leu Lys Cys Ala Arg Leu  
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Phe Pro Asp Ser Asn Leu Glu Ala Val Leu Asn Asn Asp Gly Leu Ser  
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Pro Leu Met Met Ala Ala Lys Thr Gly Lys Ile Gly Ile Phe Gln His  
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Ile Ile Arg Arg Glu Val Thr Asp Glu Asp Thr Arg His Leu Ser Arg  
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Lys Ser Lys Asp Trp Ala Tyr Gly Pro Val Tyr Ser Ser Leu Tyr Asp  
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Leu Val Tyr Asn Ser Lys Ile Glu Asn Arg His Glu Met Leu Ala Val  
530 535 540

Glu Pro Ile Asn Glu Leu Leu Arg Asp Lys Trp Arg Lys Phe Gly Ala  
545 550 555 560

Val Ser Phe Tyr Ile Asn Val Val Ser Tyr Leu Cys Ala Met Val Ile  
565 570 575

Phe Thr Leu Thr Ala Tyr Tyr Gln Pro Leu Glu Gly Thr Pro Pro Tyr  
580 585 590

Pro Tyr Arg Thr Thr Val Asp Tyr Leu Arg Leu Ala Gly Glu Val Ile  
595 600 605

Thr Leu Phe Thr Gly Val Leu Phe Phe Phe Thr Asn Ile Lys Asp Leu  
610 615 620

Phe Met Lys Lys Cys Pro Gly Val Asn Ser Leu Phe Ile Asp Gly Ser  
625 630 635 640

Phe Gln Leu Leu Tyr Phe Ile Tyr Ser Val Leu Val Ile Val Ser Ala  
645 650 655

Ala Leu Tyr Leu Ala Gly Ile Glu Ala Tyr Leu Ala Met Met Val Phe  
660 665 670

Ala Leu Val Leu Gly Trp Met Asn Ala Leu Tyr Phe Thr Arg Gly Leu  
675 680 685

Lys Leu Thr Gly Thr Tyr Ser Ile Met Ile Gln Lys Ile Leu Phe Lys  
690 695 700

Asp Leu Phe Arg Phe Leu Leu Val Tyr Leu Leu Phe Met Ile Gly Tyr  
705 710 715 720

Ala Ser Ala Leu Val Ser Leu Leu Asn Pro Cys Ala Asn Met Lys Val  
725 730 735

Cys Asn Glu Asp Gln Thr Asn Cys Thr Val Pro Thr Tyr Pro Ser Cys  
740 745 750

Arg Asp Ser Glu Thr Phe Ser Thr Phe Leu Leu Asp Leu Phe Lys Leu  
755 760 765



Ala Pro Leu

&lt;210&gt; 6

&lt;211&gt; 764

&lt;212&gt; PRT

&lt;213&gt; Homo sapiens

&lt;400&gt; 6

Met Thr Ser Pro Ser Ser Ser Pro Val Phe Arg Leu Glu Thr Leu Asp  
1 5 10 15

Gly Gly Gln Glu Asp Gly Ser Glu Ala Asp Arg Gly Lys Leu Asp Phe  
20 25 30

Gly Ser Gly Leu Pro Pro Met Glu Ser Gln Phe Gln Gly Glu Asp Arg  
35 40 45

Lys Phe Ala Pro Gln Ile Arg Val Asn Leu Asn Tyr Arg Lys Gly Thr  
50 55 60

Gly Ala Ser Gln Pro Asp Pro Asn Arg Phe Asp Arg Asp Arg Leu Phe  
65 70 75 80

Asn Ala Val Ser Arg Gly Val Pro Glu Asp Leu Ala Gly Leu Pro Glu  
85 90 95

Tyr Leu Ser Lys Thr Ser Lys Tyr Leu Thr Asp Ser Glu Tyr Thr Glu  
100 105 110

Gly Ser Thr Gly Lys Thr Cys Leu Met Lys Ala Val Leu Asn Leu Lys  
115 120 125

Asp Gly Val Asn Ala Cys Ile Leu Pro Leu Leu Gln Ile Asp Arg Asp  
130 135 140

Ser Gly Asn Pro Gln Pro Leu Val Asn Ala Gln Cys Thr Asp Asp Tyr  
145 150 155 160

Tyr Arg Gly His Ser Ala Leu His Ile Ala Ile Glu Lys Arg Ser Leu  
165 170 175

Gln Cys Val Lys Leu Leu Val Glu Asn Gly Ala Asn Val His Ala Arg  
180 185 190

Ala Cys Gly Arg Phe Phe Gln Lys Gly Gln Gly Thr Cys Phe Tyr Phe  
195 200 205

Gly Glu Leu Pro Leu Ser Leu Ala Ala Cys Thr Lys Gln Trp Asp Val  
210 215 220

Val Ser Tyr Leu Leu Glu Asn Pro His Gln Pro Ala Ser Leu Gln Ala  
225 230 235 240

Thr Asp Ser Gln Gly Asn Thr Val Leu His Ala Leu Val Met Ile Ser  
245 250 255

Asp Asn Ser Ala Glu Asn Ile Ala Leu Val Thr Ser Met Tyr Asp Gly  
260 265 270

Leu Leu Gln Ala Gly Ala Arg Leu Cys Pro Thr Val Gln Leu Glu Asp  
275 280 285

Ile Arg Asn Leu Gln Asp Leu Thr Pro Leu Lys Leu Ala Ala Lys Glu  
290 295 300

Gly Lys Ile Glu Ile Phe Arg His Ile Leu Gln Arg Glu Phe Ser Gly  
305 310 315 320

Leu Ser His Leu Ser Arg Lys Phe Thr Glu Trp Cys Tyr Gly Pro Val  
325 330 335

Arg Val Ser Leu Tyr Asp Leu Ala Ser Val Asp Ser Cys Glu Glu Asn  
340 345 350

Ser Val Leu Glu Ile Ile Ala Phe His Cys Lys Ser Pro His Arg His  
355 360 365

Arg Met Val Val Leu Glu Pro Leu Asn Lys Leu Leu Gln Ala Lys Trp  
370 375 380

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Pro Met Glu Gly Gln Glu Asp Glu Gly Asn Gly Ala Gln Tyr Arg Gly  
580 585 590

Ile Leu Glu Ala Ser Leu Glu Leu Phe Lys Phe Thr Ile Gly Met Gly  
595 600 605

Glu Leu Ala Phe Gln Glu Gln Leu His Phe Arg Gly Met Val Leu Leu  
610 615 620

Leu Leu Leu Ala Tyr Val Leu Leu Thr Tyr Ile Leu Leu Leu Asn Met  
625 630 635 640

Leu Ile Ala Leu Met Ser Glu Thr Val Asn Ser Val Ala Thr Asp Ser  
645 650 655

Trp Ser Ile Trp Lys Leu Gln Lys Ala Ile Ser Val Leu Glu Met Glu  
660 665 670

Asn Gly Tyr Trp Trp Cys Arg Lys Lys Gln Arg Ala Gly Val Met Leu  
675 680 685

Thr Val Gly Thr Lys Pro Asp Gly Ser Pro Asp Glu Arg Trp Cys Phe  
690 695 700

Arg Val Glu Glu Val Asn Trp Ala Ser Trp Glu Gln Thr Leu Pro Thr  
705 710 715 720

Leu Cys Glu Asp Pro Ser Gly Ala Gly Val Pro Arg Thr Leu Glu Asn  
725 730 735

Pro Val Leu Ala Ser Pro Pro Lys Glu Asp Glu Asp Gly Ala Ser Glu  
740 745 750

Glu Asn Tyr Val Pro Val Gln Leu Leu Gln Ser Asn  
755 760

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<212> DNA

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18

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<211> 20

<212> DNA

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<223> Description of Artificial Sequence: Primer

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taatacgact cactataggg

20

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<211> 19

<212> DNA

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ggaaacagct atgaccatg

19

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17

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19

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20

[illegible]



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<211> 21

<212> DNA

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<400> 16

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21

<210> 17

<211> 20

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence: Primer

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<210> 18

<211> 21

<212> DNA

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<220>

<223> Description of Artificial Sequence: Primer

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ctggagttag ggtctccatc c

21

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<210> 19

<211> 43

<212> DNA

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<220>

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gtcatagcgg ccgcgccgcc accatgaaga aatggagcag cac

43

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<211> 20

<212> DNA

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aggcccactc ggtgaacttc

20

<210> 21

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<223> Description of Artificial Sequence: Primer

<400> 21

gacgagcatg tacaatgaga

20

<210> 22

<211> 20

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<400> 22

gtcaccaccg ctgtggaaaa

20

<210> 23

<211> 20

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence: Primer

<400> 23

tgtggacagc tacagtgaga

20

<210> 24

<211> 32

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<400> 24

tgcaactgaat tcgagcactg gtgttcctc ag

32

<210> 25

<211> 20

<212> DNA

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<223> Description of Artificial Sequence: Primer

<400> 25

tgtggacagc tacagtgaga

20

<210> 26

<211> 19

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence: Primer

<400> 26

gtggaaaacc cgaacaaga

19

<210> 27

<211> 23

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Cys His Ile Phe Thr Thr Arg Ser Arg Thr Arg Leu Phe Gly Lys Gly

1

5

10

15

Asp Ser Glu Glu Ala Ser Cys

20

<210> 28

<211> 21

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<220>

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sequence

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Cys Gly Ser Leu Lys Pro Glu Asp Ala Glu Val Phe Lys Asp Ser Met  
1 5 10 15

Val Pro Gly Glu Lys  
20

<210> 29

<211> 20

<212> DNA

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<220>

<223> Description of Artificial Sequence: Primer

<400> 29

atggccacca gcagggttac

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<210> 30

<211> 18

<212> DNA

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<223> Description of Artificial Sequence: Primer

<400> 30

tctgccaggt tccagctg

18

<210> 31

<211> 41

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gtcatagcgg ccgcgcgcca ccatgccag gtagttgga c

41

<210> 32

<211> 20

<212> DNA

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cacctcttgt tgtcactgga

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<210> 33

<211> 23

<212> DNA

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<220>

<223> Description of Artificial Sequence: Primer

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caaattctgcg catgaagttc cag

23

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<210> 34

<211> 23

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 34

gccacgagaa gttccacgta gtg

23

<210> 35

<211> 20

<212> DNA

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<223> Description of Artificial Sequence: Primer

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gctgctccca ttcttgctga

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<210> 36

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<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 36

tgcaactctg agaaatgagt gggcagagaa gc

32

Publ. No. 97/01252B2

<210> 37

<211> 20

<212> DNA

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<223> Description of Artificial Sequence: Primer

<400> 37

atggccacca gcagggttac

20

<210> 38

<211> 18

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence: Primer

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tctgccaggt tccagctg

18

<210> 39

<211> 20

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

<400> 39

acaagaaggc ggacatgcgg

20

Publ. No. WO/00/32766



<210> 40

<211> 20

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Primer

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atctcgtggc gggttctcaat

20

ABSTRACT

**HUMAN VANILLOID RECEPTORS AND THEIR USES**

The invention provides novel human vanilloid receptor (hVR) proteins, in particular hVR1 and hVR3, nucleotide sequences encoding for the novel hVR proteins, and hVR proteins for use in a method for screening for agents useful in the treatment or prophylaxis of disorders which are responsive to modulation of hVR activity in a human patient. The invention also provides expression vectors comprising said nucleotide sequences, stable cell lines comprising said expression vectors, antibodies specific for the novel hVR proteins, methods for the identification of compounds which exhibit hVR modulating activity, compounds identifiable and identified by such methods, and methods of treatment or prophylaxis of disorders which are responsive to modulation of hVR activity in a human patient.

09/857123

# COMBINED DECLARATION FOR UTILITY OR DESIGN PATENT APPLICATION WITH POWER OF ATTORNEY

ATTORNEY'S DOCKET  
PG3606USWFirst Names Inventor:  
**DELANY**Complete if known:  
App No.:Filing Date  
Concurrently herewith

Group Art Unit:

( ) Declaration submitted with initial filing or

(X) Declaration submitted after initial filing (surcharge required 37CFR1.16(e))



23347

As below named inventor. I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name. UNITED STATES TRADEMARK OFFICE

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

## HUMAN VANILLOID RECEPTORS AND THEIR USES

the specification of which (check only one item below):

[ ] is attached hereto.

OR

[X] was filed on \_\_\_\_\_ as United States application Serial No. \_\_\_\_\_ or PCT International

Application Number EP99/09284 filed 11/30/1999 and was amended on (MM/DD/YYYY) \_\_\_\_\_ (if applicable)

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment specifically referred to above.

I acknowledge the duty to disclose information which is material to patentability as defined in 37 CFR §1.56.

I hereby claim foreign priority benefits under 35, U.S.C. §119 (a)-(d) or §365(b) of any foreign applications(s) for patent or inventor's certificate or 365(a) of any PCT international application which designated at least one country other than the United States of America, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate or of any PCT international application having a filing date before that of the application on which priority is claimed:

### PRIOR FOREIGN AND ANY PRIORITY CLAIMS UNDER 35 U.S.C. 119:

Prior Foreign Application Number (s)	Country	Foreign Filing Date (MM/DD/YYYY)	PRIORITY CLAIMED
1. GB9826359.3	GB	12/01/1998	x
2.			
3.			
4.			
5.			

I hereby claim the benefit under Title 35, United States Code §119(e) of any United States provisional application(s) listed below:

Application No.	Filing Date (MM/DD/YYYY)	

## DECLARATION FOR "371" APPLICATION

**COMBINED DECLARATION FOR UTILITY or DESIGN  
PATENT APPLICATION WITH POWER OF ATTORNEY** ContinuedATTORNEY'S DOCKET NUMBER  
**PG3606USW**

I hereby claim the benefit under 35, U.S.C. §120 of any United States application or §365(c) of any PCT international application designating the United States of America that is listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of 35 U.S.C. §112, I acknowledge the duty to disclose information which is material to patentability as defined in 37 C.F.R. §1.56 which became available between the filing date of the prior application(s) and the national or PCT international filing date of this application:

**PRIOR U.S. PARENT APPLICATION or PCT PARENT APPLICATION**

U.S. Parent Application or PCT Parent Number	Parent Filing Date (MM/DD/YYYY)	STATUS (Check one)		
		PATENTED	PENDING	ABANDONED

**POWER OF ATTORNEY:** As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the U.S. Patent and Trademark Office connected therewith. (List name and registration number)

David J. Levy Reg. No. <u>27,655</u>	James P. Riek Reg. No. <u>39,009</u>	Bonnie L. Deppenbrock Reg. No. <u>28,209</u>
Charles E. Dadswell Reg. No. <u>35,851</u>	Virginia C. Bennett Reg. No. <u>37,092</u>	John L. Lemanowicz Reg. No. <u>37,380</u>
Karen L. Prus Reg. No. <u>39,337</u>	Frank P. Grassler Reg. No. <u>31,164</u>	Amy H. Fix Reg. No. <u>42,616</u>
Robert H. Brink Reg. No. <u>36,094</u>	Christopher P. Rogers Reg. No. <u>36,334</u>	
Elizabeth Selby Reg. No. <u>38,298</u>	Lorie Ann Morgan Reg. No. <u>38,181</u>	

**Send Correspondence to:**

David J. Levy, Patent Counsel  
Corporate Intellectual Property Department  
GlaxoSmithKline,  
Five Moore Drive, PO Box 13398  
Research Triangle Park, NC 27709



23347

PATENT TRADEMARK OFFICE

**Direct Telephone Calls to:**

Frank Grassler  
919-483-2482

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under 18 U.S.C. 1001, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

1-00	FULL NAME OF INVENTOR	FAMILY NAME	FIRST GIVEN NAME	SECOND GIVEN NAME/INITIAL
	INVENTOR'S SIGNATURE	DELANY	Natalie	Samantha
	RESIDENCE & CITIZENSHIP	CITY	STATE OR FOREIGN COUNTRY	COUNTRY OF CITIZENSHIP
	POST OFFICE ADDRESS	POST OFFICE ADDRESS	CITY	STATE & ZIP CODE/COUNTRY
0		Stevenage	Hertfordshire GB	GB
1		GlaxoSmithKline Five Moore Drive, PO Box 13398	Research Triangle Park	NC 27709 US
2-00	FULL NAME OF INVENTOR	FAMILY NAME	FIRST GIVEN NAME	SECOND GIVEN NAME/INITIAL
	INVENTOR'S SIGNATURE	SANSEAU	Philippe	
	RESIDENCE & CITIZENSHIP	CITY	STATE OR FOREIGN COUNTRY	COUNTRY OF CITIZENSHIP
	POST OFFICE ADDRESS	POST OFFICE ADDRESS	CITY	STATE & ZIP CODE/COUNTRY
0		Stevenage	Hertfordshire GB	FR
2		GlaxoSmithKline, Inc. Five Moore Drive, PO Box 13398	Research Triangle Park	NC 27709 US
3-00	FULL NAME OF INVENTOR	FAMILY NAME	FIRST GIVEN NAME	SECOND GIVEN NAME/INITIAL
	INVENTOR'S SIGNATURE	TATE	Simon	Nicholas
	RESIDENCE & CITIZENSHIP	CITY	STATE OR FOREIGN COUNTRY	COUNTRY OF CITIZENSHIP
	POST OFFICE ADDRESS	POST OFFICE ADDRESS	CITY	STATE & ZIP CODE/COUNTRY
0		Stevenage	Hertfordshire GB	GB
3		GlaxoSmithKline Five Moore Drive, PO Box 13398	Research Triangle Park	NC 27709 US